

氏名	坂本敏夫
学位(専攻分野)	博士(理学)
学位記番号	総研大甲第92号
学位授与の日付	平成6年3月24日
学位授与の要件	生命科学研究科 分子生物機構論専攻 学位規則第4条第1項該当
学位論文題目	Fatty acid desaturases of cyanobacteria and the modification of membrane lipid unsaturation.
論文審査委員	主査教授 村田紀夫 教授 藤田善彦 教授 西村幹夫 助教授 岡田清孝

## 論文内容の要旨

The physical properties of biological membranes depend on the fatty acid unsaturation of membrane lipids. Cyanobacteria respond to a decrease in ambient temperature by desaturating fatty acids of membrane lipids to compensate the decrease in membrane fluidity at low temperature. Fatty acid desaturases are enzymes that introduce double bonds into hydrocarbon chain of fatty acids. The enzymes play an important role to control the fluidity of membrane lipids in the acclimation of cyanobacterial cells to low temperature.

In the cyanobacterium, *Synechocystis* PCC6803, polyunsaturated fatty acids are synthesized by sequential introduction of double bonds into C<sub>18</sub> fatty acids esterified to the *sn*-1 position of glycerolipids by four distinct desaturases, and each of the desaturases catalyzes the desaturation of fatty acids at a specific position: the  $\Delta 6$ ,  $\Delta 9$ ,  $\Delta 12$  or  $\omega 3$  position. At the beginning of the present studies, the *desA* gene that encodes the  $\Delta 12$  desaturase had been isolated from *Synechocystis* PCC6803 [Wada, H., Gombos, Z. and Murata, N. (1990) *Nature* 347: 200-203], but no other desaturase genes of cyanobacteria, or of higher plants had been reported.

In order to resolve the role of unsaturation of membrane lipids, it is necessary to control the degree of membrane lipid unsaturation by genetic manipulation of the fatty acid desaturases. In addition, the structural information of desaturases is requisite to understand the reaction mechanism of the fatty acid desaturation.

The author isolated cyanobacterial desaturase genes: the  $\Delta 12$  desaturase genes (*desA*) from three strains of cyanobacteria, i.e.,

*Synechocystis* PCC6714, *Synechococcus* PCC7002 and *Anabaena variabilis*; the  $\omega$ 3 desaturase gene (*desB*) from the cyanobacterium, *Synechocystis* PCC6803; and the  $\Delta$ 9 desaturase genes (*desC*) from *A. variabilis* and *Synechocystis* PCC6803. The membrane-lipid unsaturation of cyanobacteria was altered by manipulating the isolated desaturase genes.

Cyanobacterial *desA* genes were isolated from *Synechocystis* PCC6714, *Synechococcus* PCC7002 and *A. variabilis* by cross-hybridization with DNA probes derived from the *desA* gene of *Synechocystis* PCC6803. It appeared that the *desA* genes of *Synechocystis* PCC6714, *Synechococcus* PCC7002 and *A. variabilis* encode proteins of 349, 347 and 350 amino acid residues, respectively. The amino acid sequences of the products of the *desA* genes revealed the presence of four conserved domains. The transformation of *Synechococcus* PCC7942 with the *desA* genes from the three strains was associated with acquisition of the ability to introduce the second double bond at the  $\Delta$ 12 position of fatty acids.

Cyanobacteria respond to a decrease in temperature by desaturating fatty acids of membrane lipids to compensate the decrease in membrane fluidity. Among various desaturation reactions in cyanobacteria, the desaturation at the  $\omega$ 3 position of fatty acids is the most sensitive to the change in ambient temperature. The author isolated a gene for the  $\omega$ 3 desaturase, *desB*, from the cyanobacterium, *Synechocystis* PCC6803, using a heterologous hybridization probe derived from the *desA* gene of the same cyanobacterium. The *desB* gene encodes a protein of 359 amino-acid residues and of molecular mass of 41.9 kDa. In order to manipulate the fatty acid unsaturation of membrane lipids, the *desB* gene in *Synechocystis* PCC6803 was mutated by insertion of the

kanamycin-resistance gene cartridge. The resultant mutant was unable to desaturate fatty acids at the  $\omega$ 3 position. On the other hand, the *desA* and *desB* genes were introduced into cells of another cyanobacterium, *Synechococcus* PCC7942, which can desaturate fatty acids of membrane lipids only at the  $\Delta$ 9 position. The resultant transformant was capable of desaturating fatty acids at the  $\Delta$ 9,  $\Delta$ 12 and  $\omega$ 3 positions. These results demonstrate that the disruption and the introduction of the *desB* gene enable us to genetically manipulate the fatty acid unsaturation of membrane lipids in cyanobacteria.

In cyanobacteria, oleic acid is synthesized by introducing a double bond into stearic acid bound to polar glycerolipids. This biosynthetic pathway in cyanobacteria is distinct from the other organisms. The author found an open reading frame in the nucleotide sequence of the 5' upstream region of the *desA* gene of *A. variabilis*, and its deduced amino-acid sequence showed similarity to stearyl-CoA desaturases of rat, mouse and *Saccharomyces cerevisiae*. Thus, the author postulated that the open reading frame of *A. variabilis* encodes the cyanobacterial  $\Delta$ 9 desaturase. The gene was designated *desC*. Using a DNA fragment derived from the *desA* gene of *A. variabilis*, another *desC* gene was isolated from *Synechocystis* PCC6803. Upon expression of the *desC* gene of *Synechocystis* PCC6803 in *Escherichia coli*, the *E. coli* cells accumulated oleic acid, which is not synthesized in the wild-type cells of *E. coli*. The deduced amino-acid sequence of the *desC* gene of *Synechocystis* PCC6803 shows significant similarity to the stearyl-CoA desaturases of rat (25%), mouse (24%) and yeast (25% in the internal region corresponding to the *desC*).

Recently, the  $\Delta$ 6 desaturase gene was isolated from *Synechocystis* PCC6803 [Reddy, A.S., Nuccio, M.N., Gross, L.M. and

Thomas, T.L. (1993) *Plant Mol. Biol.* 27: 283-300]. Thus, the author has obtained the structural information of all desaturases of *Synechocystis* PCC6803. Similarity in terms of the amino acid sequence between the  $\Delta 12$  desaturase (*desA*) and the  $\omega 3$  desaturase (*desB*) is 27%. A short region of consecutive amino-acid identity was found in the  $\omega 3$  desaturase and the  $\Delta 12$  desaturase. By contrast, the extents of similarity in terms of amino acid sequence between the  $\Delta 6$  desaturase and the other desaturases, and those between the  $\Delta 9$  desaturase (*desC*) and the other desaturase are very small, namely 11-16%. However, striking similarities were found among the hydrophobic characteristics of these four desaturases of *Synechocystis* PCC6803. The predicted products of these genes contain two hydrophobic region, and, therefore, it is likely that they are membrane-bound proteins. The comparison of the local structures of the four desaturases of *Synechocystis* PCC6803 disclosed the conserved amino-acid residues among these desaturases. It is likely that the conserved amino-acid residues are responsible for the introduction of a double bond into fatty acids bound to glycerolipids.

## 論文の審査結果の要旨

生物は低温に曝されると膜脂質の脂肪酸の不飽和結合を増加させるが、これは温度低下に伴う膜の流動性の低下を補償して、低温下における生理活性を一定の水準に保つための適応現象である。脂肪酸不飽和化酵素は、膜脂質の脂肪酸に不飽和結合を導入する酵素であり、低温適応における流動性の調節において重要な役割を担っている。本研究では、ラン藻から脂肪酸不飽和化酵素の遺伝子を単離し、さらにその遺伝子操作によって膜脂質の不飽和度を改変した形質転換体を作成している。

先ず、ラン藻 *Synechocystis* PCC6803 の $\Delta$ 12位の脂肪酸不飽和化酵素の遺伝子 (*desA*) を用いて、他の3種のラン藻 (*Synechocystis* PCC6714, *Synechococcus* PCC7002, *Anabaena variabilis*) より *desA* 遺伝子を単離した。この *desA* 遺伝子を、シャトルベクター-pUC303 を用いて *Synechococcus* PCC7942 に導入し、得られた形質転換体において $\Delta$ 12位での不飽和化が進行することにより、*desA* 遺伝子の機能を同定した。続いて、*Synechocystis* PCC6803 より $\omega$ 3位の脂肪酸不飽和化酵素の遺伝子 (*desB*) を単離しその構造を明かにした。*desB* の機能は、*desB* を破壊した *Synechocystis* PCC6803 の形質転換体において、 $\omega$ 3位での脂肪酸の不飽和化が進行しないことにより同定した。また、*Synechococcus* PCC7942 に *desA* と *desB* を同時に導入することにより、 $\Delta$ 12位および $\omega$ 3位での脂肪酸の不飽和化を進行させることができた。さらに、*Anabaena variabilis* および *Synechocystis* PCC6803 より、 $\Delta$ 9位の脂肪酸不飽和化酵素の遺伝子 (*desC*) を単離し、その構造を明かにした。*desC* の機能は、不飽和化酵素を持たない大腸菌内での発現により $\Delta$ 9位での不飽和化がおこることにより同定した。

この研究の特徴は、従来単離されていなかったラン藻のアシル脂質不飽和化酵素の2種類の遺伝子を単離できたことであり、その成果はラン藻および高等植物の低温適応における膜脂質の役割を解明する研究、および不飽和化反応のメカニズム（二重結合の導入部位を決定するメカニズム）を分子レベルで解明する研究に大いに貢献するものと思われる。なおこの成果の一部は既に *Plant Molecular Biology* 誌に採択されている。

以上の内容により、審査会は当博士論文に対して合格の判断を下した。