

Doctoral Thesis

Molecular Phylogeny of Junglefowls, genus *Gallus*
and
Monophyletic Origin of Domestic Fowls

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A b s t r a c t

With the aim of elucidating the evolutionary origin of junglefowls and their domestication processes, I conducted molecular evolutionary analyses of mitochondrial DNAs for various kinds of birds belonging to the subfamily Phasianinae. I then found that the real matriarchic origin of all the domestic fowls examined in the present study was an Asian continental population of *Gallus gallus gallus*. The phylogenetic analysis conducted in this study also suggested that the continental population of *Gallus gallus gallus* is the monophyletic ancestor of all domestic fowls. These findings resolve the long-time controversy concerning monophyletic versus polyphyletic origin theories of domestic fowls.

The present thesis is composed of four chapters. In Chapter 1, as an introduction, I described the evolutionary significance of the domestication processes of junglefowls and the overview of taxonomical problems of birds within the subfamily Phasianinae, particularly junglefowls and domestic fowls.

In Chapter 2, attention is focused on the molecular phylogeny of the subfamily Phasianinae. Comparisons of DNA sequences for mitochondrial control regions among 16 avian species belonging to the subfamily Phasianinae, revealed the following: (1) Generalized perdicine birds (quails and partridges) are descended from ancient lineages. Even the closest pair,

the common quail of the Japanese subspecies (*Coturnix coturnix japonica*) and the Chinese bamboo partridge (*Bambusicola thoracica*), maintained only a 85.71% identity. (2) The 12 species of phasianine birds previously and presently studied belong to three distinct branches. The first branch is made up exclusively of members of the genus *gallus*, while the second branch is made up of pheasants of the genera *Phasianus*, *Chrysolophus* and *Syrmaticus*. Gallopheasants of the genus *Lophura* are distant cousins to these pheasants. The great argus (*Argusianus argus*) and peafowls of the genus *Pavo* constitute the third branch. Members of the fourth phasianine branch, such as tragopans and monals, were not included in the present study. (3) The one perdicine species, *Bambusicola thoracica*, is more closely related to the phasianine genera *Gallus* and *Pavo* than to members of other perdicine genera. The above might indicate that *Bambusicola* belongs to one stem of the perdicine lineage which later split into two sublineages that yielded phasianine birds; one evolving to *Gallus*, while the other differentiated toward *Pavo* and related genera. (4) Tandem duplication of the 60-base unit was established as a trait unique to the genus *Gallus*, which is shared neither by pheasant nor by quail.

In Chapter 3, I discuss evolutionary relationships between red and green junglefowls. The noncoding control region of the mitochondrial DNA of various gallinaceous birds was studied with regard to its RFLP (restriction fragment length polymorphism) and sequences of the first 400 bases. Unlike

its close ally green junglefowl, the red junglefowl *Gallus gallus* is a genetically very diverse species; a 7.0% sequence divergence was seen between those from Thailand (*Gallus gallus gallus* and *Gallus gallus spadiceus*) and that of the Indonesian island of Java (*Gallus gallus bankiva*). Furthermore, the divergence increased to 27.83% when each transversion was regarded as an equivalent of 10 transitions. On the other hand, a mere 0.5-3.0% difference (all transitions) separated various domestic breeds of chicken from two subspecies of *Gallus gallus gallus* of Thailand, thus indicating a single domestication event in the area inhabited by this subspecies, with the red junglefowl being the origin of all domestic breeds. Only transitions separated six diverse domesticated breeds. Nevertheless, a 2.75% difference was seen between RFLP type I breeds (white leghorn and nagoya) and a RFLP type VIII breed (*ayam pelung*). The above data suggest that although the mitochondrion of RFLP type V is the main contributor to domestication, hens of other RFLP types also contributed to this event.

Finally, in Chapter 4, the evolutionary origin and dispersal patterns of domestic fowls are discussed from various aspects of molecular evolution and human history. With the aim of elucidating in more detail, the genealogical origin of the present domestic fowls of the world, I determined mitochondrial DNA (mtDNA) sequences of the D-loop regions for a total of 21 birds which belong to the red junglefowl (*Gallus gallus*) comprising three subspecies (6 *Gallus gallus gallus*, 3 *Gallus gallus spadiceus* and 3 *Gallus*

gallus bankiva) and 9 birds representing diverse domestic breeds (*Gallus gallus domesticus*). I also sequenced mtDNAs from 4 green junglefowl (*Gallus varius*), 2 Cingalese junglefowl (*Gallus lafayettei*) and 1 grey junglefowl (*Gallus sonnerati*). I then constructed a phylogenetic tree for these birds using nucleotide sequences, choosing the Japanese quail (*Coturnix coturnix japonica*) as an outgroup. Moreover, I found that a continental population of *Gallus gallus gallus* was the real matriarchic origin of all the domestic species examined in the present study. It is also of particular interest to note that there were no discernible differences among *Gallus gallus* subspecies; *Gallus gallus bankiva* being a notable exception. This was because *Gallus gallus spadiceus* and a continental population of *Gallus gallus gallus* formed a single cluster in the phylogenetic tree. One obvious and distinct feature that customarily separates *Gallus gallus gallus* from *Gallus gallus spadiceus* is the color of their ear lobes: white for the former and red for the latter. The fact that domestic breeds of the chicken are of two kinds as to color of ear lobes is a clear reflection of the contribution made by *Gallus gallus gallus* as well as by *Gallus gallus spadiceus* to their ancestry. *Gallus gallus bankiva*, on the other hand, is a distinct entity, thus, deserving its subspecies status. This implies that a continental population of *Gallus gallus gallus* was the monophyletic ancestor of all domestic breeds. I also discuss the possible significance of the initial dispersal pattern of present domestic fowls, utilizing the phylogenetic tree.

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Chapter 1

Introduction

In our ever increasingly mechanized society, the fond memory of intimate tie that existed between man and his domesticated animals is rapidly fading. Yet the lasting impact that the domestication event exerted upon our history can easily be realized by recalling 16th century tragedies that befell the two great empires of the new world with wondrous architectural skills; Aztec in the middle and Inca to the south.

Horses of the genus *Equus* originally evolved in the new world and eventually their range expanded to the central Asia. This expansion was probably through the Bering landbridge during the initial Gunz period of the great ice age. It is a historical injustice of extreme proportion that by the time, the first wave of migrants from the Asia began to arrive in the new world some 30,000 years ago also through the Bering landbridge, horses were already extinct from the continent of their origin. At the same time, the domestication of wild horses in the central Asian plains was yet to take place in the ancestral land of these migrants. Because of the above two tragic coincidences, ameridians were condemned to remain *gentis sans*

equus. Gazing at the magnificent temple city ruins of Teotihuacan and Machu Picchu, one can not help but wonder how it was possible to move and pile such enormous stones without help from beasts of the burden.

It is this condemnation which enabled two spaniards, Hernando Cortes and Francisco Pizarro, to conquer the two great new world empires possessing well organized armies with less than 200 horsemen each.

It now appears that the domestication of junglefowls was no less a historical significance. Only man-made artefacts recognizable from high-orbiting manned spacecrafts is said to be the Great Wall of China. This enormous architectural project was started around 220 B.C. by Shi-huangdi, the first emperor of Qin. Many archeologists now believe that labors that built the Great Wall of China were sustained by proteins furnished by the domesticated chicken.

Needless to say, only modern man *Homo sapiens* managed to domesticate wild animals. Accordingly, the recent revelation that although our predecessors such as *Homo erectus* inhabited outside of Africa as well as Africa itself (e.g., Peking man and Java man), modern man originated in Africa only half a million years ago or so and that a small band of them moved out to the near East about 100,000 years ago or thereabout and began migration toward West, North and East was a precious gift to the students of all domestication events. This is because 100,000 years became the upper time limit of all domestication events that occurred aside of Africa.

With the aim of elucidating the evolutionary origin of junglefowls and their domestication processes, I conducted molecular evolutionary analyses of mitochondrial DNAs for various kinds of birds belonging to subfamily Phasianinae. In particular, I sequenced the noncoding D-loop region of the mitochondrial genome for the birds.

In the case of the noncoding D-loop region of the mitochondrial genome, 2% sequence difference became the upper limit marker, for this is the difference seen among all ethnic groups of man. Accordingly, one is not expected to see greater than 2% sequence difference among domestic breeds of any species, as far as the D-loop region is concerned.

The present thesis is composed of four chapters. Except Introduction in Chapter 1, three chapters from Chapter 2 to Chapter 4 describes the results of my studies. In Chapter 2, I discuss molecular phylogeny of the subfamily Phasianinae and its implications to evolution of junglefowls. In particular, I focus the discussion on phylogenetic relationships between the tribes perdicine and phasianine.

First of all, duplication of the 60-base-long unit within the D-loop region was established as a trait unique to the genus *Gallus*. If in the future, a particular perdicine (quail or partridge) species is identified that has this duplication, that quail or partridge indisputably belongs to the perdicine lineage that gave rise to the genus *Gallus* and to no phasianine birds. Inasmuch as I have already identified the Chinese bamboo partridge as a

close relative of this lineage, it will be possible to identify the species soon.

In Chapter 3, I would clarify the evolutionary relationships between red and green junglefowls, the noncoding control region of the mitochondrial DNA with regard to its restriction fragment length polymorphism (RFLP) and sequences of the first 400 bases.

In Chapter 4, I would make an attempt to elucidate the evolutionary origin and dispersal patterns of domestic fowls, in order to understand the domestication processes of junglefowls.

In this chapter, I make clarification of subspecies status within the red junglefowl (*Gallus gallus*). Although it has generally been accepted that diverse domestic chicken breeds originated from the red junglefowl, this species is thought to be comprised of five subspecies. Accordingly, the question of which is the ancestral subspecies remained.

When the famous 4000-year-old Mohenjo-Doro site was thought of as the original site of chicken domestication, the expectation fell on one particular subspecies *Gallus gallus murghi* whose range included the Indus valley. When 8000-year-old remains of domesticated chicken were found along the Huang He (yellow river) in northern China which has never been the habitat of junglefowls, the expectation moved further to East and South and fell upon a number of red junglefowl subspecies inhabiting the area comprised of south China, Thailand and Indochinese regions.

I have shown that *Gallus gallus spadiceus* inhabiting the Malay

peninsula bisects the habitat of *Gallus gallus gallus*. *Gallus gallus gallus* of Indochinese regions in the Asian mainland was separated from *Gallus gallus gallus* of the Sumatra island. Furthermore, I have shown that continental *Gallus gallus gallus* and *Gallus gallus spadiceus* was a single genetic entity quite distinct from the Sumatran *Gallus gallus gallus*, and that diverse breeds of the domestic chicken fall into this newly recognized combined continental entity. Only discernible feature separating *Gallus gallus gallus* from *Gallus gallus spadiceus* is white ear lobes versus red ear lobes. Understandably, this dichotomy has been transmitted to domestic breeds of the chicken, for they are either white ear lobed or red ear lobed.

Among the red junglefowls, excluded from the ancestry of the domestic chicken were the Sumatran *Gallus gallus gallus* population as well as *Gallus gallus bankiva* inhabiting Indonesian islands. Although *Gallus gallus murghi* has not been studied, it can qualify as an ancestor, only if it is genetically indistinct from the afore-mentioned new combined entity.

At the end, I believe that these noted above are very important for understanding the phylogeny of junglefowl and the evolutionary origin of domestic fowls and their dispersal patterns.

Chapter 2.

Molecular Phylogeny of the Subfamily Phasianinae and its Implications to Evolution of Junglefowls, Genus Gallus

2.1 Introduction

2.1.1 Purpose of Study

In this study, I established duplication of a 60-base-long unit within the mitochondrial control (D-Loop) region characteristically unique to the genus *Gallus* among phasianine birds. Furthermore, base sequence comparison of this maternally derived noncoding region revealed that as diverse as domestic breeds of chicken are, they could only have been derived from a continental subspecies (e.g., *Gallus gallus gallus*, *Gallus gallus spadiceus*) but not an island subspecies (*Gallus gallus bankiva*) of the red junglefowl, *Gallus gallus* (Fumihito *et al*, 1994). Excluded from the ancestry of domesticated chicken were three other species of junglefowls; the green (*Gallus varius*), the grey (*Gallus sonneratii*) and the Cingalese (*Gallus lafayettei*).

In this study, I wish to define the position of the genus *Gallus* within the subfamily Phasianinae in relation to other members.

2.1.2 Systematic Classifications of the Subfamily

Phasianinae

Because of their ornamental value, large pheasant-like birds of the subfamily, Phasianinae are widely kept and propagated in various zoological gardens of the world as well as by private fanciers. This is not the case with diverse generalized perdicine birds (quails and partridges) of the same subfamily. When a few of them are kept and propagated, they are maintained only as exotic game birds. Accordingly, I was able to secure blood samples from only five species, albeit from four (five) different genera, of the generalized perdicine birds which are said to consist of 103 species belonging to 22 different genera (Johnsgard, 1986). They were the common quail of the Japanese subspecies, *Coturnix coturnix japonica*, the blue-breasted quail (*Coturnix chinensis*, also known as *Excalfactoria chinensis*) the Chinese bamboo partridge, *Bambusicola thoracica* the chukar partridge, *Alectoris graeca chukar* and the European grey partridge, *Perdix perdix perdix*. In sharp contrast, there were no difficulties in securing 12 species of the large pheasant-like birds of the subfamily Phasianinae representing 8 different genera, thus, 11 of the 49 extant species were

sampled (Johnsgard, 1986). In addition to the red (*Gallus gallus*) and green (*Gallus varius*) previously reported (Fumihito *et al.*, 1994), the genus *Gallus* was represented by two additional junglefowl species; the grey (*Gallus sonneratii*) and Cingalese (*Gallus lafayettei*) junglefowls. Pheasants, as such, were represented by three species; the green (*Phasianus versicolor*), the golden (*Chrysolophus pictus*) and Mrs. Hume's (*Syrmaticus huminae*). Other large bodied birds of this subfamily that were also called pheasants for want of more descriptive words, were the silver gallopheasant (*Lophura nycthemera*), the Burmese peacock-pheasant (*Polyplectron bicalcaratum*), and the great argus pheasant (*Argusianus argus*) which is as large as a peafowl. The peafowl of the genus *Pavo* is represented by the common peafowl; (*Pavo cristatus*) and the green peafowl (*Pavo muticus*).

Unfortunately, I was unable to secure samples of one important group of phasianine birds represented by tragopans, monals and others. Also, in this study, I did not deal with members of the four other subfamilies that together with Phasianinae constitute the family Phasianidae. They were turkeys of the new-world subfamily Meleagridinae, guineafowls of the African subfamily Numidinae and toothed quails and partridges of the new-world subfamily Odontophorinae. Grouses of the old-world subfamily Tetraoninae were also excluded.

2.2 Materials and Methods

Preparation of Cell Lysate and Extraction of DNA.

DNA were extracted from blood samples obtained from live birds with no apparent harm to them.

At least 5 μ l of peripheral blood was blotted on a small piece of filter paper (approximately 5 \times 5 mm) and kept dry during transportation. Blood was eluted from a filter paper in 500 μ l of phosphate-buffered saline. After centrifugation at 5,000 rpm for 2 min, cell pellets were suspended in 100 μ l of 10 mM Tris · HCl (pH 8.3) buffer containing 50 mM KCl, 1.5 mM MgCl₂ gelatin, 0.45% Nonidet P-40, 0.45% Tween 20, and 200 μ g of proteinase K per ml. The suspension was incubated for 30 min at 60 ° C and was heated at 94 ° C for 15 min to stop the reaction. The cell lysate was then extracted twice with 400 μ l of phenol/chloroform/isoamyl alcohol (25:24:1) and total DNA was dissolved in 200 μ l of 10 mM Tris · HCl (pH7.5) buffer containing 1 mM EDTA.

Polymerase Chain Reaction(PCR).

A conserved primer pair, H1255 (5' -CATCTTGGCATCTTCAGTGCCC-3') and L16750 (5' -AGGACTACGGCTTGAAAAGC-3'), was used to amplify the control region for RFLP analysis. L and H refer to light and heavy chains and the number designates the position of the 3' end of the primer in the

reference sequence (Desjardins & Morais, 1990).

Two microliters of total DNA or cell lysates was subjected to 35 amplification cycles using *Taq* (*Thermus aquaticus*) DNA polymerase (Takara Shuzo Co. Ltd.) according to the manufactures' instructions, with denaturation at 94° for 1 min, annealing at 55° C for 1 min, and extension at 72° C for 2 min.

Nucleotide Sequencing.

Because of the presence of an EcoRI site within, the primer H1254 (5'-ATGAATTCTGGCATCTTCAGTGCCA-3') was used instead of H1255 to obtain PCR products for cloning. The base sequence of another primer already given, L16775, did contain a *Hind*III site. When the above H1254/L16775 pair was used for PCR amplification, 3.0 mM MgCl₂ replaced the 1.5mM concentration recommended. PCR products were digested with *Eco*RI and *Hind*III and purified by agarose gel electrophoresis. Ligation of the DNA segments into the *Eco*RI/*Hind*III site of the cloning vector pUC118, transformation of *Escherichia coli* JM109, and single-strand DNA preparation using the helper phage M13K07 were performed as previously described (Desjardins & Morais, 1991). To minimize errors introduced by *Taq* DNA polymerase during PCR, two or three clones obtained from each sample were used for sequencing. Sequencing was carried out with the BcaBEST dideoxynucleotide sequencing kit (Takara Shuzo Co. Ltd.) using a fluorescein

isothiocyanate labeled M13 forward primer (Shimazu Co. Ltd.) and DNA sequencer DSQ1 (Shimazu Co. Ltd.).

Phylogenetic tree reconstruction

The nucleotide sequences obtained were aligned with each other (Figure 2-1), and the number of nucleotide substitutions were estimated by the six-parameter method (Gojobori et al., 1982). A phylogenetic tree was constructed by the neighbor-joining method, using the substitution numbers estimated (Saitou & Nei, 1987). Statistical significance was evaluated by the bootstrapping method (Felsenstein, 1985).

2.3 Results and Discussion

2.3.1 Evolutionary relationships between perdicine and phasianine birds

Sequences at the 392 positions of the mitochondrial control region of 14 species were aligned and are shown in three parts in Figures 2-1 and 2-2 (Fumihito *et al.*, 1994). The 60-base-long unit containing the nearly invariant tetradecamer, AACTATGAATGGTT in its center is duplicated only in members of the genus *Gallus* and no other. Accordingly, the interspecific sequence comparison presently carried out only involved the original of this 60-base-long unit. Further, the roughly 50-base-long region immediately in front of this 60-base-long region or its duplicated copy is hypervariable in all species (Fumihito *et al.*, 1994). In as much as liberal introduction of gaps was needed to maximize interspecific homology of this hypervariable region, it was thought advisable to eliminate this region from the present sequence comparison. Accordingly, the dendograms of perdicine and phasianine birds shown in Figures 2-1 and 2-2 were drawn based upon differences at a total of 392 positions; positions 1 to 192 and then positions 241 to 441 (307 to 507 or 370 to 570) in the case of *Gallus*, depending upon numbers of copies of the 60-base-long unit individual birds possessed.

2.3.2 The antiquity of generalized perdicide lineages

The antiquity of generalized perdicide lineages (quails and partridges).

The differences at 392 positions of the mitochondrial noncoding control region of five perdicide species are shown at the top of Figure 2-1 and the dendrogram drawn based upon these differences, is shown immediately below. It should be noted that the European grey partridge (*Perdix perdix*) was separated from the rest by the greatest distance. The second in remoteness was the chukar partridge (*Alectoris graeca chukar*). Although the remaining three appeared to be cohorts of one cohesive group, the closest relationship was seen within a pair comprised of representatives of two separate tribes; *Coturnix coturnix japonica* representing the tribe Corturnicini and *Bambusicola thoracica* of the tribe Perdicini (Johnsgard, 1986). Regardless of whether the common quail of the Japanese subspecies and the blue-breasted quail belong to the same genus *coturnix* or to two separate genera, *Coturnix* and *Excalfctoria*, the two belonged to the same tribe. Yet, the distance separating the two appeared slightly greater than that between the common quail and the Chinese bamboo partridge (*Bambusicola thoracica*) in the perdicide dendrogram of Figure 2-1. At any rate, even the closest pair of perdicide species shared identical bases at only 336 of the 392 positions (a mere 85.71% identity), as shown at the bottom of Figure 2-2.

2.3.3 Three distinct branches of phasianine birds

The base sequences of the nine phasianine species are aligned in four rows from the bottom half of Figure 2-1 to the top of Figure 2-2. Adding the base sequences of red and green junglefowls (*Gallus gallus* and *Gallus varius*) previously published (Fumihito *et al.*, 1994), as well as the sequence of the green peafowl (*Pavo muticus*) shown separately at the bottom of Figure 2-2, to the present comparison, the dendrogram of 12 phasianine species was drawn and is shown at the middle of Figure 2-2. It can immediately be seen that the first major division separated the branch made of peafowls of the genus *Pavo*, the great argus and the peacock-pheasant, from the rest of phasianine birds. The remaining birds were further subdivided into two branches. One was made exclusively of members of the genus *Gallus*, while the other was a pheasant branch. In addition to those belonging to genera *Phasianus*, *Chrysolophus* and *Syrmaticus*, gallopheasants of the genus *Lophura* were included in this branch. My phasianine dendrogram shown in the middle of Figure 2-2 is in essential agreement with that previously produced on the basis of classical taxonomic studies (Figure 2-3; Johnsgard, 1986). Unfortunately, those that are thought to be members of the fourth independent branch were not included in the present study. They were tragopans, monals (*Lophophorus*) and the koklass (*Pucrasia*) (Johnsgard, 1986).

2.3.4 Perdicine versus phasianine lineages

I made a number of attempts to construct a combined dendrogram of perdicine and phasianine species studied. In the process, it was found that when two rather distantly related groups are combined, the relative positions of individual species in a dendrogram tend to shift by an addition to or a subtraction from that diagram, of a few species. In as much as my survey is by no means comprehensive, lacking representation from a few key groups as already noted, I concluded that the construction of a combined diagram at this time would be inadvisable.

2.3.5 Paradoxical genetic link between the Chinese bamboo partridge and members of the genera *Gallus* and *Pavo*

In spite of the extreme meagerness of perdicine species in the present survey, one tangible link emerged between perdicine and phasianine lineages, but this link was a paradoxical one. At the bottom of Figure 2-2, the mitochondrial DNA base sequence of *Bambusicola thoracia* is aligned with that of the green peafowl (*Pavo muticus*) on the one hand and that of the green junglefowl (*Gallus varius*) on the other. Pertinent information extracted from these alignments are shown immediately below them. It can be seen that the Chinese bamboo partridge is genetically far closer to phasianine *Gallus* (87.75% identity) and *Pavo* (89.03% identity) than to its

closest perdicine ally, *Coturnix coturnix japonica* (85.71% identity).

A paradox is found in the fact that *Gallus* and *Pavo* belong to two very divergent phasianine lineages as evident in the dendrogram of Figure 2-2 as well as in the fact that there is only 85.02 % sequence identity between *Gallus varius* and *Pavo muticus* (Figure 2-2, bottom). While nearly equal affinities toward *Bambusicola thoracica* were shown by all four species of *Gallus* as well as by both species of *Pavo*, no hint of a close link was seen between *Bambusicola* and other members of the peafowl branch; i.e., *Argusianus argus* and *Polyplectron bicalcaratum*.

2.3.6 Evolutionary features of Chinese bamboo partridge

Poul A. Johnsgard, in his authoritative treatise on the subject "The Pheasants of the World" follows the customary subdivision of the subfamily Phasianinae to Perdicini and Phasianini (Johnsgard, 1986). Yet, he is of the opinion that four branches of phasianine birds sprang independently from generalized perdicine ancestors. This view is succinctly illustrated in his dendrogram shown in Figure 2-3 (Johnsgard, 1986). Indeed, the present study established one direct link between a particular perdicine lineage represented by *Bambusicola thoracica* and two phasianine genera *Gallus* and *Pavo*. The only problem was that *Gallus* and *Pavo* belong to two very divergent phasianine branches. It is quite conceivable that *Bambusicola*

with only two extant species, Chinese bamboo partridge (*B. thoracica*) and Bamboo partridge (*B. fytchii*) belongs to one stem of the perdicine lineage which later split into two phasianine-yielding sublineages; one being ancestral to *Gallus*, with the other giving rise to *Pavo* and related genera. If so, one might expect to locate a species or species belonging to one or the other of the above phasianine-yielding sublineages among close relatives of *Bambusicola*. The most attractive in this respect are three species of *Galloperdix*. If the implication from its generic name holds true, they might show even closer affinity to *Gallus* than *Bambusicola*, while manifesting little affinity to *Pavo*. Unfortunately, all attempts to secure samples of *Galloperdix* which inhabit India and Sri Lanka failed.

There is however an independent line of evidence that links *Bambusicola* with *Gallus*. In 1949, Yamashina classified the chromosome complements of members of the subfamily, *Phasianinae*, to four major types. Although the karyotype of *Bambusicola*, *Coturnix* and *Gallus* were found to belong to the same type, *Pavo* was found to belong to a different type (Figure 2-4; Yamashina, 1949).

2.4 Summary

Further comparison of mitochondrial control region DNA sequences of 16 avian species belonging to the subfamily, *Phasianinae* revealed the following: (1) Generalized perdicine birds (quails and partridges) are descended from ancient lineages. Even the closest pair, the common quail of the Japanese subspecies (*Coturnix coturnix japonica*) and the Chinese bamboo partridge (*Bambusicola thoracica*), maintained only an 85.71% identity. (2) The 12 species of phasianine birds previously and presently studied belonged to three distinct branches. The first branch was made exclusively of members of the genus *gallus*, while the second branch was made of pheasants of the genera *Phasianus*, *Chrysolophus* and *Syrmaticus*. Gallopheasants of the genus *Lophura* were distant cousins to these pheasants. The great argus (*Argusianus argus*) and peafowls of the genus *Pavo* constituted the third branch. Members of the fourth phasianine branch, such as tragopans and monals were not included in the present study. (3) The one perdicine species, *Bambusicola thoracica* was more closely related to the phasianine genera *Gallus* and *Pavo* than to members of other perdicine genera. The above might indicate that *Bambusicola* belongs to one stem of the perdicine lineage which later split into two sublineages that yielded phasianine birds; one evolving to *Gallus*, with the other differentiating toward *Pavo* and related genera.

Chapter 3.

Evolutionary Relationships between Red and Green Junglefowls

3.1 Introduction

There is little doubt that the domestication of various wild animals as beasts of burden, sources of protein and fat, and instruments of war and recreation has played a pivotal role in the cultural evolution of mankind. Of special interest has been the divine rites performed in association with various domesticated animals, particularly the chicken. For documentation of so recent an event as domestication, nuclear genes with their low mutation rate would be of little use. On the contrary, the mitochondrial genome appears particularly suitable. Its high mutation rate is expected to be constant, being relatively impervious to generation time differences between species. It may be recalled that an organism does not start its life with a single copy but with hundreds of thousands of copies of the mitochondrial genome harbored in the egg cytoplasm. Accordingly, generation changes do not constitute significant epochs in the life history of mitochondrial DNA. Furthermore, an extremely useful marker for measurement was established in recent studies on two hypervariable

subregions of the control region of human mitochondria. The average sequence divergence between all races of mankind was established as 2.0% and the rate of evolution was estimated to be 1% sequence divergence per 71,000-167,000 years (Cann *et al.*, 1987; Vigilant *et al.*, 1989). Under the assumption that the rate of human mtDNA divergence is almost comparable to that of domestic species mtDNA, it follows that any mitochondrial sequence divergence substantially above 2.0% within a given domesticated species creates a peculiar paradox of either domestication occurring before the emergence of mankind or at least domestication occurring within the African cradle before the exodus of certain bands to the Near East and outward.

Indeed, such a paradox was encountered in a recent study on the mitochondrial control region of various breeds of cattle. Two distinct mitochondrial lineages separated by a 5.01% sequence difference were observed. This paradox was resolved by the assumed presence of two subspecies of an ancestor of cattles, the aurochs (*Bos primigenius*), prior to the emergence of humans and the two subsequent independent domestication events after human divergence (Loftus *et al.*, 1994).

In view of the above data, I decided to study the control region light chain (L chain) of the avian mitochondria on various gallinaceous birds with regards to its restriction fragment length polymorphism (RFLP) as well as the sequence of the first 400 bases of the control region. In human studies,

64% of total polymorphism in the entire control region was found among the first 400 bases (di Rienzo & Wilson, 1991).

3.2 Materials and Methods

Materials.

Materials used for the present study are summarized in Table 3-1. Red junglefowls (*G. g. gallus* and *G. g. spadiceus*) were gifts from the Department of Forestry of the Thai government. Five specimens of *G. g. bankiva* were obtained from the Indonesian island of Java, and so are all of the green junglefowl (*Gallus varius*). Four additional Thai red junglefowl sampled were from those kept in the Tama Zoological Garden (Tokyo). Of various domestic breeds, samples from all those classified as "occidental breeds" were collected at The Domestic Fowl Trust of England. As to Asiatic breeds, those starting with "ayam" were all Indonesian breeds and were collected there. Others were either collected in their native habitats or obtained from one of the following three institutions in Japan: Yamashina Institute for Ornithology, The Research Institute of Evolutionary Biology, or Hiroshima Animal Husbandry Experimental Station.

Detection of RFLP.

After testing 36 restriction enzymes, the following four were chosen as

suitable: *Alu* I (recognition sequence, AGCT), *Mse* I (TTAA), *Mbo* II (TCTTC), And *Vsp* I (ATTAAT). RFLPs were detected on either 1.5% or 4.0% agarose gels after 30 min to 1 hr of electrophoresis at 80 V.

Most other methods, such as preparation of cell lysate and extraction of DNA, PCR and nucleotide sequencing are described in Chapter 2.

3.3 Results and Discussion

3.3.1 RFLP within the 1200- to 1300-base control region

As noted in *Materials and Methods*, four restriction enzymes recognized polymorphic cleavage sites within the control region, thus yielding different sized fragments readily distinguishable by gel electrophoresis. These four restriction enzymes were *Vsp* I, *Alu* I, *Mse* I, and *Mbo* II. Inasmuch as the last two enzymes recognized two polymorphic sites each, a total of six sites were involved in RFLP(Table. 3-1). The first four polymorphic sites are shown in Figure 3-1. Of the potential 64 (2^6) types involving the six sites, eight were found among domestic breeds and their wild ancestors, red junglefowls. Six additional types were seen among more distantly related green junglefowls. Thus, 14 of the 64 potential types are in existence today.

Table. 3-1 shows that regardless of whether they belong to breeds long established in the West (Europe and North America) or to breeds that remained in Asia, the predominant RFLP type among domesticated chickens

was type V, closely followed by type I. While type V was also found in more than half of the red junglefowls of the three subspecies sampled, types I, II, and IV have not thus far been found among red junglefowls. Conversely, type VII has been confined to one subspecies of Thai red junglefowl (*G. g. spadiceus*) in spite of the sampling of >27 diverse domestic breeds. Of particular interest was type VIII. Among domesticated chickens, this type was seen only in those breeds which had originated in Indonesia. At the same time, 1 of the 19 red junglefowls exhibiting RFLP type VIII was also of Javanese origin. The above data might be interpreted as an indication of multiple sites of domestication - i.e., Indonesia starting from the independent regional domestication of *G. gallus bankiva*. In the past, various population studies utilizing isozymes as well as blood group polymorphisms suggested such multiple and independent sites of domestication (Hashiguchi *et al.*, 1983).

The green junglefowl (*G. varius*) manifested its own polymorphism composed of six allelic forms, here designated as types A,B,C,D,E, and F. However, Table. 3-1 shows that while the second *Mbo* II site in all individuals of *G. gallus* was cleavable by the enzyme, the corresponding site in all 30 *G. varius* was not. If one excludes this second *Mbo* II site from consideration type I of *G. gallus* now becomes the same as type A of *G. varius* and the same applies to type II and type B. The above suggests that RFLP observed in *G. gallus* and *G. varius* has been a very ancient polymorphism

antedating the separation of *G. gallus* from *G. varius*.

3.3.2 Tandem duplication of 60-base unit within the control region as a genus-specific trait of Gallus

Before comparison of base sequences with regard to the first 400 bases of the control region L chain, the tandem duplication of one 60-base unit in members of the genus *Gallus* as a genus-specific trait should be noted. The control region base sequence of white leghorn (Desjardins & Morais, 1990) and that of the common quail of the Japanese subspecies *Coturnix coturnix japonica* (Desjardins & Morais, 1991) have been published, and it was shown that the control region of the latter was 41 bases shorter than that of the former. My own sequencing of type I white leghorn and Japanese quail produced only one discrepancy from the published sequences noted above. The base triplet CCC underlined in the first section of Figure 3-1 was missing from the published sequences of both white leghorn (Desjardins & Morais, 1990) and Japanese quail (Desjardins & Morais, 1991).

As shown in the fifth section of Figure 3-1 (marked "Original"), the 60-base unit containing the invariant tetradecamer AACTATGAATGGTT in its center is present as a single unit in the quail, whereas tandem duplication of this unit was observed in all 11 *G. gallus* as well as all four *G. varius* individuals. In Figure 3-1, a copy of the original located immediately

upstream is marked "1st copy." Also I found this duplication to be present in the third and fourth members of the genus *Gallus*: the grey junglefowl (*Gallus sonneratii*) and Cingalese junglefowl (*Gallus lafayettei*). Yet, their closest relatives, various pheasants of the genus *Phasianus* were quail-like, having this 60-base unit in a solitary state. Among members of the family Phasianidae, pheasants were thought to be far more closely related to the chicken than quails are, as evidenced by the fact that pheasant-chicken hybrids are fully viable, albeit sterile, whereas only 0.15-2.0% of incubated eggs produce live chicken-Japanese quail hybrids (Haley, 1965). Yet, the sequence comparison between the original and its first copy on every one of the 15 sequenced individuals of *G. gallus* and *G. varius* indicated that the average difference was 20%. Interestingly the sequence difference between originals of the Japanese quail and of *Gallus* was 25%. On the basis of the above noted evidence, it appears that duplication of the 60-base-long unit has taken place immediately after the genus *Gallus* came into being. Tandem repeats within the control region of mitochondrial DNA have previously been reported in two papers: 79-base tandem repeats in three subspecies of the masked shrew (*Sorex sinreus* : Insectivora) (Stewart & Baker, 1994) and 10-base tandem repeats in canine mitochondrial DNA (Tsuchida *et al.*, 1994).

Once duplication started, further duplication would have been inevitable (Ohno, 1970). Indeed, one extra copy of the 60-base unit was found in three green junglefowl of RFLP type C, while two extra copies were

found in one green junglefowl of RFLP type E (Table 3-1 and Figure 3-1).

One each of these individuals with one and two extra copies was sequenced.

When sequence comparisons were made between the original of green junglefowl nos. 32 and 50 and their own "second extra copies," a uniform sequence difference of 13.1% was noted. The above data revealed that the initial further duplication that produced a second extra copy from the original was a rather ancient affair, probably antedating the speciation of *G. varius*. Indeed, the presence of the second extra copy was also noted in certain individuals of *G. sonnerati* as well as *G. layayettei*. The generation of the "third extra copy" by green junglefowl no. 50, on the other hand, was a very recent event, for it differed only by a single base substitution from the second extra copy of the same individual (Figure 3-1).

3.3.3 Sequence Differences Between *G. varius* and *G. gallus* and Affinity of All Domestic Breeds to the Thai Red Junglefowl (*G. g. gallus*).

Of the four *G. varius* individuals sequenced, two (nos. 6 and 32) were of the same RFLP type C. In spite of the fact that the latter was endowed with the second extra copy, these two demonstrated the least sequence divergence at 1.50%. Furthermore, all the substitutions were transitions (Figure 3-1). In view of the considerable antiquity of the second extra copy

already discussed, this probably means the recent loss of the second extra copy by the lineage represented by no. 6. The difference between these two RFLP type C individuals and no. 2 of RFLP type D increased to 2.25%, while a 3.20% sequence difference separated no. 50 of RFLP type E from the rest. Furthermore, these differences included a few transversions (Figures 3-1 and 3-2).

In contrast to the green junglefowl (*G. varius*), which is a local species confined to the Indonesian Islands, the red junglefowl (*G. gallus*) inhabits a very large area: the Asian mainland stretching from northeastern India in the west to the western coast of China to the east. In addition, its range includes various Indonesian Islands where it is sympatric with *G. varius* as well as Hainan Island in the South China Sea. It is no surprise that *G. gallus* has often been subdivided into five subspecies (Zeuner, 1963).

As shown in Figure 3-1, when dealing with different subspecies, the same RFLP type was no indication of genetic similarity. Both red junglefowl no. 15 and the domestic breed *ayam pelung* no. 76 were typed as RFLP type VIII and they were from the same Indonesian island. Yet, a 5.75% sequence divergence separated the two. Furthermore, 9 of the 23 substitutions were transversions (Figure 3-1). The above clearly excluded the involvement of *G. gallus bankiva* from the domestication event. In sharp contrast, all three Thai red junglefowls (two *G. g. gallus* and one *G. g. spadiceus*) were very close to all breeds of domestic chicken. The closest affinity, of only 0.5% (one

each of transition and deletion) difference, was seen between Thai red junglefowl no. 11 of RFLP type V and a member of the Indonesian breed, *ayam cemani*, of the same RFLP type (Figure 3-3). From phylogenetic trees constructed by the Unweighted Pairwise Grouping (UPG) method (Figure 3-2) and the Neighbor Joining (NJ) method (Figure 3-4), of three subspecies of red junglefowls, *G. g. gallus* (Thai nos. 8 and 11) was far more closely related to *G. g. spadiceus* (Thai no. 3) from the adjacent area than to *G. g. bankiva* from Java (Indonesian no. 15). Nevertheless, a transversion was involved in the difference between the first two and RFLP type VII was unique to *G. g. spadiceus*.

3.3.4 Evolutionary relationships between junglefowls and domestic fowls

Because of findings at the well-known Mohenjo-Doro site in Pakistan, it was believed for a long time that the original domestication of the chicken occurred in the Indus Valley 4,000 years ago (Zeuner, 1963). Subsequently, however, earlier signs of domestication were found in unlikely places far removed from the habitat of junglefowls-e.g., Ukraine and Spain, dated at more than 4,000 years ago (West & Zhou, 1988). Indeed, the earliest time of domestication of the chicken was pushed back to nearly 8,000 years ago. Remains of domesticated chickens were evident in 16 neolithic sites along

the Huang He (Yellow River), Peiligan and Cishan, rivers in Northeast China (Figure 3-4), and some of these sites were dated to be at least 7,500 years old (West & Zhou, 1988). Inasmuch as the semiarid steppe environment of these loess highlands of North China has never been a suitable habitat for red junglefowls, the time and place of the original domestication is probably earlier and further to the south and the west. The present findings place the original site of domestication in the area inhabited by a single subspecies of the red junglefowl (*G. g. gallus*). However, as it would be shown shortly, our subsequent studies revealed no genetic distinction between continental populations of *G. g. gallus* and *G. g. spadiceus*. *G. g. gallus* inhabiting the island of Sumatra, on the other hand, was distinct from the first two.

In studying sequence divergence of the human mitochondrial DNA control region, Vigilant *et al.* (Vigilant *et al.*, 1989) noted that while all intraspecific base substitutions were transitions, the difference between humans and chimpanzees involved a number of transversions. Accordingly, the observed sequence divergence of 13.6% between the two species was converted to a 42% difference by regarding each transversion as an equivalent of 10 transitions in accordance with Nei (Nei, 1987). In the present study, I found that not only interspecific differences, but also intraspecific differences within *G. varius* involved transversions. This was also true of differences between three subspecies of *G. gallus* (*G. g. gallus* and *G. g. spadiceus* combined as one subspecies and *G. g. bankiva*). In sharp

contrast, all differences within one combined subspecies (*G. g. gallus* and *G. g. spadiceus*) and diverse domestic breeds were transitions (Figure 3-3).

Among the diverse domestic breeds presently studied, the greatest sequence divergence, 2.75%, separated two breeds of RFLP type I from that of RFLP type VIII. This was considerably more than the 1.25% that separated two individual *G. g. gallus* from Thailand. The sampling of this subspecies from distant areas such as Sumatra Island is expected to expand the scope of data on intrasubspecific diversity. It would thus appear that this subspecies alone may have yielded all the diverse breeds of domestic chicken, provided that not only the mainstream RFLP type V but also a number of hens of other RFLP types contributed to the domestication event.

3.4 Summary

The noncoding control region of the mitochondrial DNA of various gallinaceous birds was studied with regard to its RFLP and sequences of the first 400 bases.

Tandem duplication of the 60- base unit was established as a trait unique to the genus *Gallus*, which is shared neither by pheasant nor by quail. Unlike its close ally, green junglefowl (*Gallus varius*), the red junglefowl *Gallus gallus* is a genetically very diverse species; the 7.0% sequence divergence was seen between those from Thailand (*G. g. gallus* and *G. g. spadiceus*) and that from the Indonesian island of Java (*G. g. bankiva*). Furthermore, the divergence increased to 27.83% when each transversion was regarded as an equivalent of 10 transitions. On the other hand, a mere 0.5-3.0% difference (all transitions) separated various domestic breeds of the chicken from two subspecies of *G. g. gallus* of Thailand, thus indicating a single domestication event in the area inhabited by this subspecies with the red junglefowl being the origin of all domestic breeds. Only transitions separated six diverse domesticated breeds. Nevertheless, a 2.75% difference was seen between RFLP type I breeds (white leghorn and nagoya) and a RFLP type VIII breed (*ayam pelung*). The above data suggests that although the mitochondrion of RFLP type V was the main contributor to domestication, hens of other RFLP types also contributed to this event.

Chapter 4.

Evolutionary Origin and Dispersal Patterns of Domestic Fowl

4.1 Introduction

4.1.1 Purpose of Study

There is little doubt that the successive domestication of various wild animals contributed greatly to the sustenance and cultural developments of mankind. In particular, the domestication of chickens seems to have a long history. As I mentioned in chapter 3, the earliest sign of domestication of the chicken is estimated to be nearly 8,000 years ago, because remains of domesticated chickens were evident in 16 neolithic sites along the Huang He (Yellow River) in Northeast China and some of these sites were dated to be at least 7,500 years (West & Zhou, 1988). The domestication of chickens has also been observed at the well-known Mohenjo-Doro site in the Indus Valley in Pakistan (Zeuner, 1963). Moreover, earlier signs of domestication of the chicken were found in unlikely places, far distant from the habitat of junglefowls - e.g., Ukraine and Spain (West & Zhou, 1988).

Accordingly, the question of whether the domestication of junglefowls

occurred only once in a fixed site or occurred repeatedly in different locations had been raised time and again. In fact, various population studies utilizing isozyme polymorphism suggested multiple and independent sites of domestication (Hashiguchi *et al.*, 1983).

As I noted in chapter 3, I studied the noncoding region of the mitochondrial DNA of various gallinaceous birds with regard to RFLP and sequences of the first 400 bases. I then showed that while a 7.0% sequence divergence was seen between *G. g. bankiva* which inhabits the Indonesian island of Java and two other subspecies of *Gallus gallus*, both inhabitants of Thailand (*G. g. gallus* and *G. g. spadiceus*), a mere 0.5 - 3.0% difference separated three domestic breeds of the chicken from divergent geographical origins in two subspecies of *G. g. gallus* of Thailand. Thus, I indicated the strong possibility that a single domestication event took place in Thailand and its immediate surroundings (Fumihito *et al.*, 1994). In as much as the sequence divergence between two individuals of *G. g. gallus* from Thailand was 1.25%, it was concluded that this subspecies alone had been sufficient to yield all the diverse breeds of domestic chicken (Fumihito *et al.*, 1994). However, since this conclusion has been drawn from a limited number of *G. gallus* subspecies and domestic fowls, it was felt that the validity of the above conclusion should be verified by a more expanded survey.

4.1.2 Geographical distribution of junglefowls

With the aim of elucidating in more detail, the genealogical origin of the present domestic fowls in the world, I determined mitochondrial DNA (mtDNA) sequences of the D-loop regions for a total of 21 birds which belong to three subspecies of *Gallus gallus* (6 *G. g. gallus*, 3 *G. g. spadiceus* and 3 *G. g. bankiva*) and 9 domestic breeds and/or races (*G. g. domesticus*) such as white leghorn. Also, sequencing was done at the same regions of mtDNA for all other *Gallus* species (4 *G. varius*, 2 *G. lafayettei*, and 1 *G. sonneratii*). A phylogenetic tree was constructed using these sequence data. The phylogenetic tree constructed verified the previous conclusion that the genealogical origin of the domestic fowls is monophyletic descending from a combined continental population of *G. g. gallus* and *G. g. spadiceus*. I shall also discuss the possible significance of the initial dispersal pattern of the present domestic fowls, utilizing the phylogenetic tree.

4.2 Materials

The species which I examined in the present study are listed in Table 4-1. Three subspecies of *Gallus gallus* were represented by 6 specimens from *G. g. gallus*, 3 from *G. g. spadiceus*, and 3 from *G. g. bankiva*. Nine specimens were from diverse domesticated breeds. Moreover, we examined 3 other species of junglefowls of the genus *Gallus*. They were 4 specimens of *G. varius*, 2 of *G. lafayettei*, and 1 of *G. sonnerati*.

4.3 Results and Discussion

4.3.1 Evolutionary relationships among four species in the genus *Gallus*

The genus *Gallus* is comprised of four species, *G. gallus*, *G. varius*, *G. lafayettei*, and *G. sonnerati*. Their phylogenetic relationship shown in Figure 4.2 is consistent with the result of chapter 2 (Fumihito *et al.*, 1995) in which the dendrogram was constructed by using of only one specimen from each species. At first glance, *G. varius* appears to be the most divergent of the four species. However, the bootstrap value that sets this species apart is only 34.7%. Accordingly, it is likely that *G. varius*, *G. gallus*, *G. sonnerati*, and *G. lafayettei* branched out from a common stem almost simultaneously in the evolutionary time scale. Taking into account the phylogenetic tree in Figure 4.2 in my previous report (Fumihito *et al.*, 1995) together with the present results, I speculate that shortly after the split, *Gallus* of the Indian subcontinent yielded *G. sonnerati* and *G. lafayettei*.

In my first paper of this series (Fumihito *et al.*, 1994), I found that a 60-base-long unit containing the nearly invariant tetradecamer, AACTATGAATGGTT in the center of the D-loop region is duplicated only in members of the genus *Gallus* and no other phasianine birds. Furthermore, more than two copies of this unit were found in three of the four *Gallus* species. In the case of *G. varius*, while 2 copies were found in two

individuals, 3 and 4 copies were found in the remaining two. All specimens of *G. lafayettei* as well as *G. sonnerati* had 3 copies. Of particular significance was the fact that all 12 specimens of *G. gallus* have 2 copies, irrespective of subspecies. These observations on copy numbers of the 60-base-long unit are consistent with the above assumed order of divergence of the 4 species. All 9 domestic fowls examined had the same copy number as *G. gallus*, again implying that the genealogical origin of the domestic fowls is monophyletic descending from *G. gallus* alone.

4.3.2 A single cluster comprising a continental population of *G. g. gallus* and *G. g. spadiceus*

As shown in Figure 4-2, *G. g. gallus* and *G. g. spadiceus* do not form two separate clusters in the phylogenetic tree. It seems that *G. g. gallus* is divided into two separate populations; an island population from Sumatra which is distinct from the continental population. *G. g. spadiceus* is more closely allied with a continental population of *G. g. gallus* than an island population of *G. g. gallus* is to its continental relation. Thus, the subspecies status given to *G. g. spadiceus* might be questioned. On the other hand, *G. g. bankiva* which inhabits the Indonesian islands of Sumatra, Java and Bali is a very distinct entity that is clearly separate from *G. g. gallus* as well as *G. g. spadiceus*, thus, its subspecies status appears to be well deserved.

4.3.3 Geographical differentiation of *G. g. gallus* and *G. g. spadiceus* and domestic fowls

West and Zhou (West & Zhou, 1988) suggested that chickens were first domesticated in Southeast Asia and were taken north to become established in China, possibly spreading to European Celts via tribes of the Russian steppe. The phylogenetic tree (Figure 4-2) seems to be consistent with their contention for the following reasons.

According to the phylogenetic tree (Figure 4-2), domestic fowls including Indonesian races belong to the same cluster as continental populations of *G. g. gallus* and *G. g. spadiceus* sampled from Thailand and its adjacent areas. On the other hand, 3 specimens of *G. g. gallus* from South Sumatra form a separate cluster as already noted, yet native domestic fowls from the same island belong to the cluster in which a continental population of *G. g. gallus* and *G. g. spadiceus* from Thailand was included; e.g., two *ayam kokok balenggek* from West Sumatra. Accordingly, a domestication event in Sumatra from that of its own *G. g. gallus* can be excluded. It suggests that the original domestication took place in Thailand and its adjacent regions and subsequently dispersed to West Sumatra in Indonesia.

West Sumatra in Indonesia is known as Minangkabau. It is worth noting that the people of Minangkabau have developed a unique culture and

ayam kokok balenggek was adopted as a symbol of the Solok prefecture in Minangkabau (Fumihito & Tanaka, 1994). It is quite possible that this domestic fowl has been highly esteemed in Minangkabau because of its novelty, implying that *ayam kokok balenggek* had been brought to West Sumatra from elsewhere. These facts support the idea that Indonesian native domestic fowls have been transferred to Indonesia from Thailand and its adjacent regions.

4.3.4 Sequence differences between *G. g. bankiva* and other *Gallus gallus* subspecies

It is clear from Figure 4-2 that *G. g. bankiva* is a very distinct entity apart from *G. g. gallus* and *G. g. spadiceus* in the phylogenetic tree as already noted. In fact, the numbers of nucleotide substitutions separating *G. g. bankiva* from other two subspecies are quite large compared with the largest individual differences seen within *G. g. gallus* as well as *G. g. spadiceus*. Thus, *G. g. bankiva* is indeed deserving of its subspecies status within the species *Gallus gallus*. Because two other subspecies of *Gallus gallus*, *G. g. murghi* and *G. g. jabouillei*, were not included in the present study, the validity of their subspecies status still remains an unanswered question.

Nevertheless, the monophyletic origin of domestic fowls from a

continental population of *G. g. gallus* appears is evident. The Indus valley where it is evident the domestication of chicken took place at least 4,000 years ago, was and still is the domain of *G. g. murghi*. Yet, the present study indicates that if *G. g. murghi* contributed to an independent domestication event, *G. g. murghi* should be genetically close to some of the domestic fowls examined here. It naturally follows that *G. g. murghi* has to be genetically indistinct from *G. g. gallus* as well as *G. g. spadiceus* of Thailand, thus, relinquishing its subspecies status. The same applies to another subspecies not included in the present study; *G. g. jabouillei*.

4.3.5 Possible significance of the initial dispersal pattern of domestic fowls

As I have shown in the present study, it is likely that all domestic fowls have originated from a single domestication event in Thailand and its adjacent regions. The reason for the domestication of chickens is usually seen as the need to secure readily available sources of protein and fat in the form of eggs and meat. However, there are other aspects. We should not forget that since time immemorial, the chicken has often been associated with various religious services in different parts of the world; e.g. the cock as a harbinger of the sun to rise once again and cockfighting as a divine offering (Crawford, 1984 ; Stevens, 1991). The initial dispersion of domestic fowls might have been due to the religious significance attached to the

chicken. In particular, the fact that cockfighting is even now wide spread throughout Southeast Asian countries appears very significant in considering the reasons for the initial dispersion of domestic fowls from the original site of domestication.

I intend to trace this initial dispersion of domestic fowls with highly polymorphic DNA markers. Inasmuch as the religious significance attached to the chicken may have been the primary reason for the initial dispersion of domestic fowls, my future work would be in the realm of ethnozoology.

4.4 Summary

With the aim of elucidating in more detail the genealogical origin of the present domestic fowls of the world, I determined mitochondrial DNA (mtDNA) sequences of the D-loop regions for a total of 21 birds which belong to the red junglefowl (*Gallus gallus*) comprising of three subspecies (6 *Gallus gallus gallus*, 3 *Gallus gallus spadiceus* and 3 *Gallus gallus bankiva*) and 9 birds representing diverse domestic breeds (*Gallus gallus domesticus*). I also sequenced 4 green junglefowl (*Gallus varius*), 2 Lafayette's junglefowl (*Gallus lafayettei*) and 1 grey junglefowl (*Gallus sonneratii*). We then constructed a phylogenetic tree for these birds using of nucleotide sequences, choosing the Japanese quail (*Coturnix coturnix japonica*) as an outgroup. We found that a continental population of *G. g. gallus* was the real matriarchic origin of all the domestic poultries examined in the present study. It is also of particular interest to note that there were no discernible differences among *Gallus gallus* subspecies; *G. g. bankiva* being a notable exception. This was because *G. g. spadiceus* and a continental population of *G. g. gallus* formed a single cluster in the phylogenetic tree. One obvious and distinct feature that customarily separated *G. g. gallus* from *G. g. spadiceus* was a color of their car lobes: white for the former and red for the latter. The very fact that domestic breeds of the chicken are of the two kinds as to colors of their ear lobes is a clear reflection of the contribution made by *G. g.*

gallus as well as by *G. g. spadiceus* to their ancestry. An irony is that aside from this very distinct dimorphism, these two do not constitute distinct entities, thus, being undeserving of separate subspecies status. *G. g. bankiva*, on the other hand, was a distinct entity, thus, deserving its subspecies status. This implies that a continental population of *G. g. gallus* sufficed as the monophyletic ancestor of all domestic breeds. We also discussed the possible significance of the initial dispersal pattern of the present domestic fowls, utilizing the phylogenetic tree.

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TYP	RFLP	JUNGLEFOWLS	DOMESTIC FOWLS							
			V	A	Ms	Mb	Ms	Mb	ASIATIC	OCCIDENTAL
I	- - - - + +	THAI RED 3/14 INDONESIAN RED 3/5	NAGOYA	1/1						
			GIFU-JIDORI	2/2					WHITE LEGHORN 3/3	
			BLACK SILKY	1/3					INDIAN GAME 2/2	
			THAI BANTAM	5/8						
			AYAM KATAI	1/5						
II	- - - - - +		BLACK SILKY	1/3						
III	+ - - - + +	THAI RED 3/14	THAI BANTAM	3/8						
			WHITE LEGHORN(HIROSHIMA VAR.)	1/2						
IV	+ - - + + +	THAI RED 7/14 INDONESIAN RED 3/5	TOHMARU	1/2					BARRED PLYMOUTH ROCK 3/3	
			BLACK SILKY	1/3					WHITE PLYMOUTH ROCK 1/1	
			WHITE LEGHORN (HIROSHIMA VAR.)	1/2					RHODE ISLAND RED 1/1	
			TOHMARU	1/2					LIGHT SUSSEX 2/2	
			WHITE SILKY	1/1					BUFF COCHIN 2/2	
			DARK BRAHMA	1/1					PARTRIDGE COCHIN 2/2	
			MALAY GAME	2/2					SILVER GREY DORKING 2/2	
			SUMATRA GAME	2/2					JERSEY BLACK GIANT 2/2	
			AYAM BANGKOK	2/2					BROWN LEGHORN 1/1	
			AYAM BEKISAR	2/4					LA FRECHE 1/1	
			AYAM CEMANI	1/2					ARAUACANA 1/1	
			AYAM KEDU	2/9					HOUDAN 2/2	
			AYAM PELUNG	1/5					FAYOMI 1/1	
V	+ + - + + +	THAI RED 1/4 INDONESIAN RED 1/5	AYAM CEMANI	1/2						
VI	+ + - + - +	THAI RED 1/4 INDONESIAN RED 1/5	AYAM BEKISAR	2/4						
			AYAM KATAI	4/5						
VII	+ + + + + +	THAI RED 3/14	AYAM KEDU	7/9						
			AYAM PELUNG	4/5						
A	- - - - + -	GREEN 1/30								
B	- - - - - -	GREEN 1/30								
C	- - + - - -	GREEN 18/30 GREEN 3/30 (60-BASE-LONG INSERTION)								
D	- - + - + -	GREEN 2/30								
E	+ - + - - -	GREEN 3/30 GREEN 1/30 (120-BASE-LONG INSERTION)								
F	- + - - - -	GREEN 1/30								

Table 3-1 Distribution of 14 RFLP types among 121 individuals of *G. gallus* (red junglefowls and domestic breeds) and *G. varius* (green junglefowls) is shown. Mitochondrial control region amplified by PCR contained six polymorphic sites for four restriction enzymes. V, *Vsp* I; A, *Alu* I; Ms, *Mse* I; Mb, *Mbo* II. On the left, each RFLP type is defined as cleavable (+) or not cleavable (-) at each of the six sites. RFLP types of *G. gallus* are numbered in Roman numerals I-VIII, whereas those of *G. varius* are shown as A-F. Nevertheless, types I and II of the former and types A and B of the latter are related (see text). With regard to each wild species and subspecies as well as to each domestic breed, distribution is expressed as number of individuals of a particular RFLP type per total number studied. Aside from 14 Thai red junglefowls (10 *G. g. gallus* and 4 *G. g. spadiceus*) and 5 Indonesian red junglefowls (*G. g. bankiva*) and 30 green junglefowls, 72 individuals representing 26 diverse domestic breeds, 3 of them in 2 varieties each were studied. Although all domestic breeds are ultimately of Asiatic origin, those long established in Europe and the New World were classified as occidental in contrast to those that stayed in Asia.

Species / Subspecies / Races	Localities
<i>Gallus gallus bankiva</i> 15	Singaraja, Bali, Indonesia
<i>Gallus gallus bankiva</i> 18	West Java, Indonesia
<i>Gallus gallus bankiva</i> 19	Lanpung, East Sumatra, Indonesia
<i>Gallus gallus gallus</i> 8,10	Department of Forestry, Thailand
<i>Gallus gallus gallus</i> 11	Tama Zoological Garden, Tokyo, Japan
<i>Gallus gallus gallus</i> 39,41,58	Palembang, South Sumatra, Indonesia
<i>Gallus gallus spadiceus</i> 3,4,5	Department of Forestry, Thailand
<i>Gallus lafayettei</i> 1,2	Dehiwela Zoo, Dehiwela, Sri Lanka
<i>Gallus sonneratii</i>	India
<i>Gallus varius</i> 2,6	Singaraja, Bali, Indonesia
<i>Gallus varius</i> 32	Madura, Indonesia
<i>Gallus varius</i> 50	Banyuwangi, East Java, Indonesia
<i>Gallus gallus domesticus</i> (Ayam Cemani 1)	Surabaya Zoo, Surabaya, East Java, Indonesia
<i>Gallus gallus domesticus</i> (Ayam Kokok Balengkek 63,71)	Solok, West Sumatra, Indonesia
<i>Gallus gallus domesticus</i> (Ayam Pelung 76)	Bogor, Jawa, Indonesia
<i>Gallus gallus domesticus</i> (Barred Plymouth Rock 1)	The Domestic Fowl Trust, Worcestershire, UK
<i>Gallus gallus domesticus</i> (Nagoya)	Hiroshima Animal Husbandry Experimental Station, Hiroshima, Japan
<i>Gallus gallus domesticus</i> (Thai Bantam 8)	Thailand
<i>Gallus gallus domesticus</i> (White Leghorn 1)	Quotation from Desjardins & Morais (1990) J.Mol.Biol. 212,599-634
<i>Gallus gallus domesticus</i> (White Leghorn 2)	Hiroshima Animal Husbandry Experimental Station, Hiroshima, Japan
<i>Coturnix coturnix japonica</i>	Hiroshima, Japan

Table 4-1. List of the specimens and their localities.

FIGURE LEGENDS

Chapter 2

Figure 2-1. Multiple alignments and a phylogenetic tree for the mitochondrial control region sequences of five perdicine species, and multiple alignments for the mitochondrial control region sequences of nine of twelve phasianine species.

At the top, the 392 positions (Positions 1 to 192 and positions 241 to 441) of the mitochondrial control region sequences of the five perdicine species are aligned and shown in four rows. The major base of each position is shown by a large capital letter, while minor bases are shown by small capital letters adorned with asterisks. The largely invariant tetradecameric core of the 60-base-long unit is underlined with a solid bar, as is the subsequent, nearly equally invariant decameric unit. The length of the 60-base long unit is also indicated by a thin underline.

Based upon the above sequence comparison, a dendrogram of five perdicine species was drawn in accordance with the neighbor-joining method (Saitou & Nei, 1987) and is shown next. Only shared sites were dealt with and the length of each line indicates the distance from a branch point expressed as the number of base substitutions per site. The bootstrap probability of each estimated distance being correct is expressed as a

percentage at each branch point based upon 1,000 replication trials.

At the bottom, nine of twelve phasianine mitochondrial sequences are aligned and continued on the top of Figure 2-2.

Figure 2-2. Multiple alignments and a phylogenetic tree for the phasianine mitochondrial sequences.

The last row of the phasianine mitochondrial sequence alignments continued from Figure 2-1 is followed by the dendrogram of 12 phasianine species. At the bottom, the sequence of the Chinese bamboo partridge (*Bambusicola thoracica*) is shown aligned with the sequence of the green peacock (*Pavo muticus*) on one hand and that of a green junglefowl (*gallus varius*) on the other. At the very bottom, the identity and differences between four pertinent pairs of species are shown. Each identity is shown as a percentage and the number of identical sites per total of 392 is shown in parentheses. Differences are shown as numbers of gaps, transitions and transversions.

Figure 2-3. Dendrogram of postulated evolutionary relationships among the species of pheasants (Illustration after Johnsgard, 1986).

Figure 2-4. Evolutionary tree of phasianidae, based on karyotype analysis (Illustration after Yamashina, 1949).

Chapter 3

Figure 3-1. Multiple alignment for the mitochondrial control region sequences of nine gallinaceous birds.

L-chain sequences of the first 400 bases of the mitochondrial control region from nine gallinaceous birds representing two wild species (*G. gallus* and *G. varius*) and three domestic breeds are aligned using the published Japanese quail (*C. coturnix japonica*) sequence as the reference (Desjardins and Morais, 1991). These nine individuals and their RFLP types are identified on the left. At each polymorphic site, the majority base is shown in a large capital letter, whereas a minority base(s) is shown in small capital letters marked by asterisks. CCCbase triplets underlined in the first section were bases missed in two previous publications (Desjardins and Morais, 1990; Desjardins and Morais, 1991). Four polymorphic positions within three potential restriction sites are so indicated, *Vsp* I in the second section, *Alu* I and *Mse* I in the third section, and *Mbo* II in the bottom section. At these sites, cleavable sequences are underlined. The invariant 14-base sequence in the center of each 60-base unit is underlined and so is the invariant 10-base unit residing near the end (last section).

Figure 3-2. Phylogenetic tree constructed by Unweighted Pairwise Grouping (UPG) method, based on sequence divergence with regard to the

first 400 bases of the mitochondrial control region of four *G. varius* and two *G. g. gallus* and one each of *G. g. spadiceus* and *G. g. bankiva*. Japanese quail (Desjardins and Morais, 1991) was chosen as the outgroup. Sequence difference is shown as percentage at each branch point. Often larger percentages in parentheses are derived by regarding each transversion as an equivalent of 10 transitions.

Figure 3-3. Polymorphic sites in the mitochondrial control region sequences of junglefowls and domestic fowls.

With regard to two sequenced members of *G. g. gallus* (Thai nos. 8 and 11) and one of *G. g. spadiceus* (Thai no.3) and seven individuals representing six domestic breeds and four RFLP types, individual bases at 24 polymorphic positions are identified. Asterisk at position 317 marks a single instance of transversion (A to C). All other substitutions are transitions. Shown below alignments are sequence differences in percentages of pertinent pairs.

Figure 3-4. Phylogenetic tree constructed by Neighbor Joining (NJ) method. The values given beside the branches indicate the bootstrap probabilities.

Figure 3-5. First places where the domestication occurred in the world (Illustration after West & Zhou, 1988).

Chapter 4

Figure 4-1. Multiple alignment of mtDNA sequences for 32 specimens of junglefowls, domestic fowls and the common quail of Japanese subspecies.

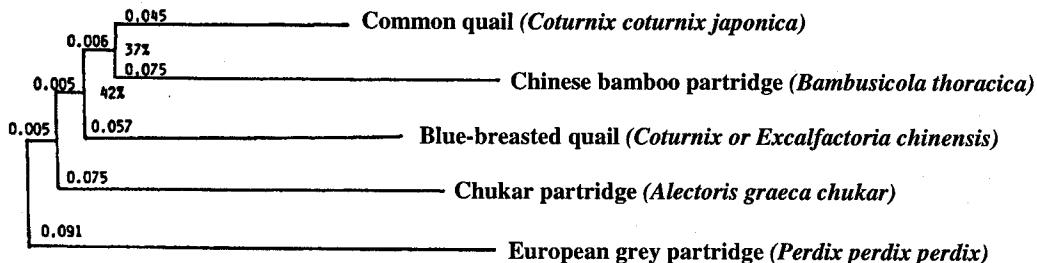
Figure 4-2. Phylogenetic tree constructed by the Neighbor Joining (NJ) method, using nucleotide sequence data. The values given beside the branches indicate the bootstrap probabilities.

1) Common quail (*Coturnix coturnix japonica*)
 2) Blue-breasted quail (*Coturnix or Excalfactoria chinensis*)
 3) Chinese bamboo partridge (*Bambusicola thoracica*)
 4) Chukar partridge (*Alectoris graeca chukar*)
 5) European grey partridge (*Perdix perdix p.*)

1) ATḠATAATCGTCATACATTATATTCACATATĀTATGGTACCGGTAATATATATĀ-CGTACTAAACCATTATATGTATACGG&CATTACAT̄-ATT̄CCC-CATTCTC
 2) ATḠATAATCGTCATACATTATATTCACATATĀTATGGTACCGGTAATATATATĀ-CGTACTAAACCATTATATGTATACGGCATTAĀB-ATT̄GCC-CATTCTC
 3) ATḠATAATCGTCATACATTATATC̄CACATATĀ-TATGGTACCGGTAATATATĀ-CGTACTAAACCATTATAC̄TATACAGCATTAC̄-TATAPCCCACATTCTC
 4) ATḠATAATCGTCATATTTATATC̄CACATATĀ-TATGGTACCGGTAATATATĀ-CGTACTAAACCATTATATGTAC̄CAGGACATAĀBATTAGCC-CATTCTC
 5) ATḠATAACḠCGTCATATTTATATC̄CACATATĀ-TATGGTACCGGTAATATATĀ-CGTACTAAACCATTATATGTAC̄CAGGACATAĀBATTAGCC-CATTCTC

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 1) CCCATGTAC CTTC̄CACTAAC=AGḠCACCAA-CTTAGAATGGT̄CAGGACATAĀ-CTTAGAATGGT̄ĀĀTĀT̄=TĀCT̄-C̄C̄CATTTGGTTATGCT̄ḠACGTACAGAGATGG
 2) CCATGTAC CTAĀ-C̄C̄AAACAGḠ-CAGCATAĀ-AGATGGT̄ĀCAGGACATAĀ-CTTĀ-ĀĀTĀT̄=ATT̄GGTTATGCTGCTGTĀCAGATGG
 3) CCCATGTAC AAT̄-CACTAACAGḠ-CACC=TĀ-CTTAGAATGGT̄CAGGACATAĀ-CTTĀ-ĀĀTĀT̄=ATT̄GGTTATGCTḠCAGGACAGATGG
 4) CCAACGḠ CTC̄-C̄C̄C̄CAGḠ-CACC=TĀ-CTTAGAATGGT̄CAGGACATAĀ-CTTĀ-ĀĀTĀT̄=TĀ-TḠB̄ACT-CCACAT̄-TGGTTATGCT̄ḠACGTACAGATGG
 5) CTCAGTGC XXAḠ-C̄C̄CAGḠ-CAGGACATAĀ-CTTĀ-ĀĀTĀT̄=ATT̄GGTTACAGGACATAĀ-CTTĀ-ĀĀTĀT̄=ĀC̄-CCACAT̄-TGGTTATGCTḠCAGGACAGATGG

1) ATTTATTGATCGTACACCTTCAGGAGAGATCĀCAACCCCTḠCTGTAATḠB̄TTCT̄-TGACTAGCTCAGGCCATTCTTCCCCCTACACCCCTGCCCCTC
 2) ATTTATTGATCḠCACCTTCAGGAGAGATCĀCAACCCCTGCCCCTḠCTGTAATḠB̄TTCT̄-TGACTAGB̄CTAGGCCATTCTTCCCCCTACACCCCTGCCCCTC
 3) ATTTATTGATCGTACACCTTCAGGAGAGATCĀCAACCCCTGCCCCTḠCTGTAATḠB̄TTCT̄-TGACTAGB̄CTAGGCCATTCTTCCCCCTACACCCCTGCCCCTC
 4) ATTTATTGATCḠCACCTTCAGGAGAGATCĀCAACCCCTGCCCCTḠCTGTAATḠB̄TTCT̄-TGACTAGB̄CTAGGCCATTCTTCCCCCTACACCCCTGCCCCTC
 5) ATTTATTGATCḠCACCTTCAGGAGAGATCĀCAACCCCTGCCCCTḠCTGTAATḠB̄TTCT̄-TGACTAGB̄CTAGGCCATTCTTCCCCCTACACCCCTGCCCCTC



6) Grey junglefowl (*Gallus sonneratii*)
 7) Cingalese junglefowl (*Gallus lafayetii*)
 8) Green pheasant (*Phasianus versicolor*)
 9) Golden Pheasant (*Chrysolophus pictus*)
 10) Mrs. Hume's Pheasant (*Syrmaticus humiae*)
 11) Silver Pheasant (*Lophura nycthemera*)
 12) Great argus (*Argusianus argus*)
 13) Burmese peacock-pheasant (*Polyplectron bicalcaratum*)
 14) Common peafowl (*Pavo cristatus*)

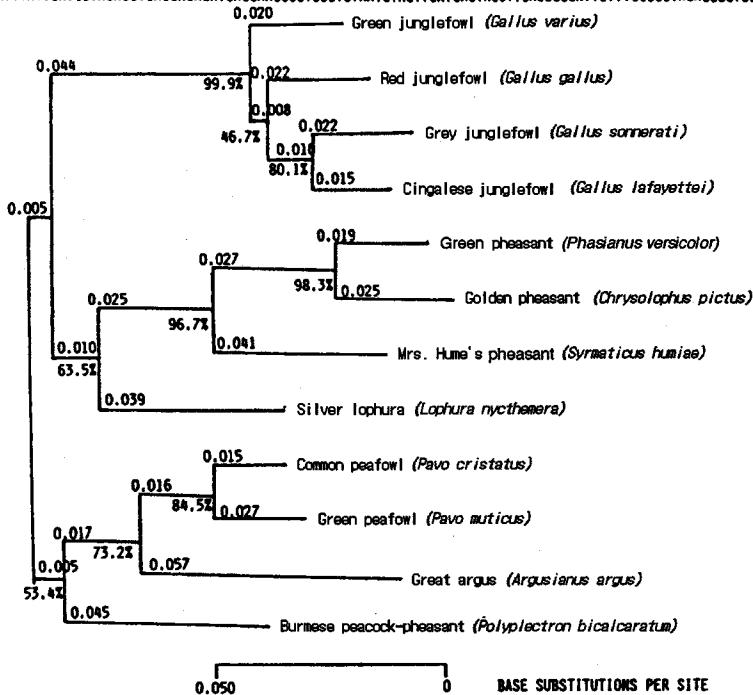
6) ATACTATGATAATCGTCATACATTATATACACATATĀ-TTATGGTACCGGTAATATATĀ-CGTACTAAACCATTATATGTACGG&CATTAĀC̄FATAT̄CCC-C
 7) ATACTATGATAATCGTCATACATTATATACACATATĀ-TTATGGTACCGGTAATATATĀ-CGTACTAAACCATTATATGTACGG&CATTAĀC̄FATAT̄CCC-C
 8) ATACTATGATAATCGTCATACATTATATACACATATĀ-TTATGGTACCGGTAATATATĀ-CGTACTAAACCATTATATGTACGG&CATTACACCAC̄-CCC-C
 9) ATACTATGATAATCGTCATACATTATATACACATATĀ-TTATGGTACCGGTAATATATĀ-CGTACTAAACCATTATATGTACGG&CATTACACCAC̄-CCC-C
 10) ATACTATGATAATCGTCATACATTATATACACATATĀ-TTATGGTACCGGTAATATATĀ-CGTACTAAACCATTATATGTACGG&CATTAĀC̄FATAT̄CCC-C
 11) ATACTATGATAATCGTCATACATTATACACATATĀ-TTATGGTACCGGTAATATATĀ-CGTACTAAACCATTATATGTACGG&CATTAĀC̄FATAT̄CCC-C
 12) ATACTATGATAATCGTCATACATTATACACATATĀ-TTATGGTACCGGTAATATATĀ-CGTACTAAACCATTATATGTACGG&CATTACACCAC̄-CCC-C
 13) ATACTATGATAATCGTCATACATTATACACATATĀ-TTATGGTACCGGTAATATATĀ-CGTACTAAACCATTATATGTACGG&CATTACACCAC̄-CCC-C
 14) ATACTATGATAATCGTCATACATTATACACATATĀ-TTATGGTACCGGTAATATATĀ-CGTACTAAACCATTATATGTACGG&CATTACACCAC̄-CCC-C

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 6) ATTCCTCCC=ATGTAC CTC̄-C̄C̄C̄CAACTAGTCACCA=TAACTATGAATGGT̄CAGGACATḠB̄CTTĀĀĀTĀT̄=ATGTC-TĀC̄C̄CATTTGGTTATGCTGCTGTĀ
 7) ATTCCTCCC=ATGTAC CTC̄-C̄C̄C̄C̄TAACTATGAATGGT̄CAGGACATḠB̄CTTĀĀĀTĀT̄=ATGTC-TĀC̄C̄CATTTGGTTATGCTGCTGTĀ
 8) ATTCCTCCC=ATGTAC CTC̄-C̄C̄C̄C̄ATGTAAATGGT̄CAGGACATḠB̄CTTĀĀĀTĀT̄=ATGTC-TĀC̄C̄CATTTGGTTATGCTGCTGTĀ
 9) ATTCCTCCC=ATGTAC CTC̄-C̄C̄C̄C̄ATGTAAATGGT̄CAGGACATḠB̄CTTĀĀĀTĀT̄=ATGTC-TĀC̄C̄CATTTGGTTATGCTGCTGTĀ
 10) ATTCCTCCC=ATGTAC CTC̄-C̄C̄C̄C̄ATGTAAATGGT̄CAGGACATḠB̄CTTĀĀĀTĀT̄=ATGTC-TĀC̄C̄CATTTGGTTATGCTGCTGTĀ
 11) ATTCATCCC=ATGTAC CTC̄-C̄C̄C̄C̄ATGTAAATGGT̄CAGGACATḠB̄CTTĀĀĀTĀT̄=ATGTC-TĀC̄C̄CATTTGGTTATGCTGCTGTĀ
 12) ATTCATCCC=ATGTAC CTC̄-C̄C̄C̄C̄ATGTAAATGGT̄CAGGACATḠB̄CTTĀĀĀTĀT̄=ATGTC-TĀC̄C̄CATTTGGTTATGCTGCTGTĀ
 13) ATTCATCCC=ATGTAC CTC̄-C̄C̄C̄C̄ATGTAAATGGT̄CAGGACATḠB̄CTTĀĀĀTĀT̄=ATGTC-TĀC̄C̄CATTTGGTTATGCTGCTGTĀ
 14) ATTCATCCC=ATGTAC CTC̄-C̄C̄C̄C̄ATGTAAATGGT̄CAGGACATḠB̄CTTĀĀĀTĀT̄=ATGTC-TĀC̄C̄CATTTGGTTATGCTGCTGTĀ

Figure 2-1.

441 (572)

6) CAGATGGATTATTGATCGT~~C~~ACTCTACAGAGAGATCAGCAACCCCTGCCCTGTAAATGACTTCATGAC~~A~~G~~T~~TCAGGCCCATCTTTCCCCCTAACACCCCTGCCCT~~A~~CTTG
 7) CAGATGGATTATTGATCGT~~C~~ACTCTACAGAGAGATCAGCAACCCCTGCCCTGTAAATGACTTCATGAC~~A~~G~~T~~TCAGGCCCATCTTTCCCCCTAACACCCCTGCCCT~~A~~CTTG
 8) CAGATGGATTATTGATCGT~~C~~ACTCTACAGAGAGATCAGCAACCCCTGCCCTGTAAATGACTTCATGAC~~A~~G~~T~~TCAGGCCCATCTTTCCCCCTAACACCCCTGCCCT~~A~~CTTG
 9) CAGATGGATTATTGATCGT~~C~~ACTCTACAGAGAGATCAGCAACCCCTGCCCTGTAAATGACTTCATGAC~~A~~G~~T~~TCAGGCCCATCTTTCCCCCTAACACCCCTGCCCT~~A~~CTTG
 10) CAGATGGATTATTGATCGT~~C~~ACTCTACAGAGAGATCAGCAACCCCTGCCCTGTAAATGACTTCATGAC~~A~~G~~T~~TCAGGCCCATCTTTCCCCCTAACACCCCTGCCCT~~A~~CTTG
 11) CAGATGGATTATTGATCGT~~C~~ACTCTACAGAGAGATCAGCAACCCCTGCCCTGTAAATGACTTCATGAC~~A~~G~~T~~TCAGGCCCATCTTTCCCCCTAACACCCCTGCCCT~~A~~CTTG
 12) CAGATGGATTATTGATCGT~~C~~ACTCTACAGAGAGATCAGCAACCCCTGCCCTGTAAATGACTTCATGAC~~A~~G~~T~~TCAGGCCCATCTTTCCCCCTAACACCCCTGCCCT~~A~~CTTG
 13) CAGATGGATTATTGATCGT~~C~~ACTCTACAGAGAGATCAGCAACCCCTGCCCTGTAAATGACTTCATGAC~~A~~G~~T~~TCAGGCCCATCTTTCCCCCTAACACCCCTGCCCT~~A~~CTTG
 14) CAGATGGATTATTGATCGT~~C~~ACTCTACAGAGAGATCAGCAACCCCTGCCCTGTAAATGACTTCATGAC~~A~~G~~T~~TCAGGCCCATCTTTCCCCCTAACACCCCTGCCCT~~A~~CTTG



15) Green peafowl (*Pavo muticus*) A~~A~~~~C~~~~T~~TTTTTTAACCTAACCCCCCTACT~~G~~TG~~T~~&CCCCCA~~G~~GGGGGGTATAC
 3) Chinese bamboo partridge (*Bambusicola thoracica*) A~~A~~~~T~~TTTCTTTAACCTAACCCCCCTT~~G~~TG~~T~~GTACCCCCCTTCCCCCAGGGGAGGTATAC
 16) Green junglefowl (*Gallus varius*) A~~A~~~~T~~TTT~~A~~ACCC~~A~~CCCCCTACT~~G~~TG~~T~~ACCCCCCTTCCCCCAGGGGGTATAC
 15) TATGCATAATCGTCATACATTATACCACAT~~A~~C~~G~~TAATACTATACGTA~~A~~ACCCATTATATGTA~~G~~ACGGACATTAC~~A~~TAT~~E~~TTCCCCATT~~A~~TC
 3) TATG~~T~~ATAATCGTCATACATTATACCACATATATTATG~~T~~ACCGGTAATACTATACGTA~~A~~ACCCATTAT~~A~~GTACGACATTACTCTATAT~~E~~CC~~A~~TTTCTC
 16) TATGCATAATCGTCATACATTATACCACATATATTATG~~T~~ACCGGTAATACTATACGTA~~A~~ACCCATTATATG~~T~~ACGGACATTAC~~A~~CTA~~T~~TTCCCCATTCTC
 15) CCCA~~G~~T¹⁹² 241(307) C~~E~~ACAC~~C~~ACAAG~~G~~ACCT AACTATGAATGGT~~T~~ ACAGGACATAA~~C~~TT~~E~~-TAT~~A~~~~G~~ CTCT~~E~~CCCATT~~T~~GGTTATGCTCG~~G~~GT~~A~~~~T~~CAGATGGAT
 3) CCCATGTAC A~~T~~~~T~~CACTAACAGT~~G~~ACCT AACTATGAATGGT~~T~~ ACAGGACATAA~~C~~TT~~E~~-TAT~~A~~~~G~~ CTCTAC~~C~~ATT~~T~~GGTTATGCTCG~~G~~GT~~A~~~~T~~CAGATGGAT
 16) CCCATGTAC C~~T~~AC~~C~~CTAACAGT~~G~~ACCT AACTATGAATGGT~~T~~ ACAGGACATAA~~C~~TT~~E~~-TAT~~A~~~~G~~ AT~~T~~CA~~C~~CCATT~~T~~GGTTATGCTCG~~G~~GT~~A~~~~T~~CAGATGGAT

438(503)

15) TTATTGATCGTACACCTCACGAGAGATCAGCAACCCCTGCCGTAAATGACTTCATGACT~~G~~~~T~~TCAGGCCATTCTTCCCCCTAACACCCCTG~~G~~
 3) TTATTGATCGTACACCTCACGAGAGATCAGCAACCCCTGCCGTAAATGACTTCATGACT~~G~~~~T~~TCAGGCCATTCTTCCCCCTAACACCCCTG~~G~~
 16) TTATTGATCGT~~C~~ACACCTCACGAGAGATCAGCAACCCCTGCCGTAAATGACTTCATGAC~~A~~G~~T~~TCAGGCCATTCTTCCCCCTAACACCC~~T~~G~~G~~

	IDENTITY	GAPS	DIFFERENCES	TRANSVERSIONS
<i>Bambusicola thoracica</i>	85.71% (336/392)	13	20	23
<i>Coturnix coturnix japonica</i>				
<i>Pavo muticus</i>	86.82% (336/387)	5	21	25
<i>Gallus varius</i>				
<i>Bambusicola thoracica</i>	88.37% (342/387)	2	20	23
<i>Pavo muticus</i>				
<i>Bambusicola thoracica</i>	89.15% (345/387)	3	20	19
<i>Gallus varius</i>				

Figure 2-2.

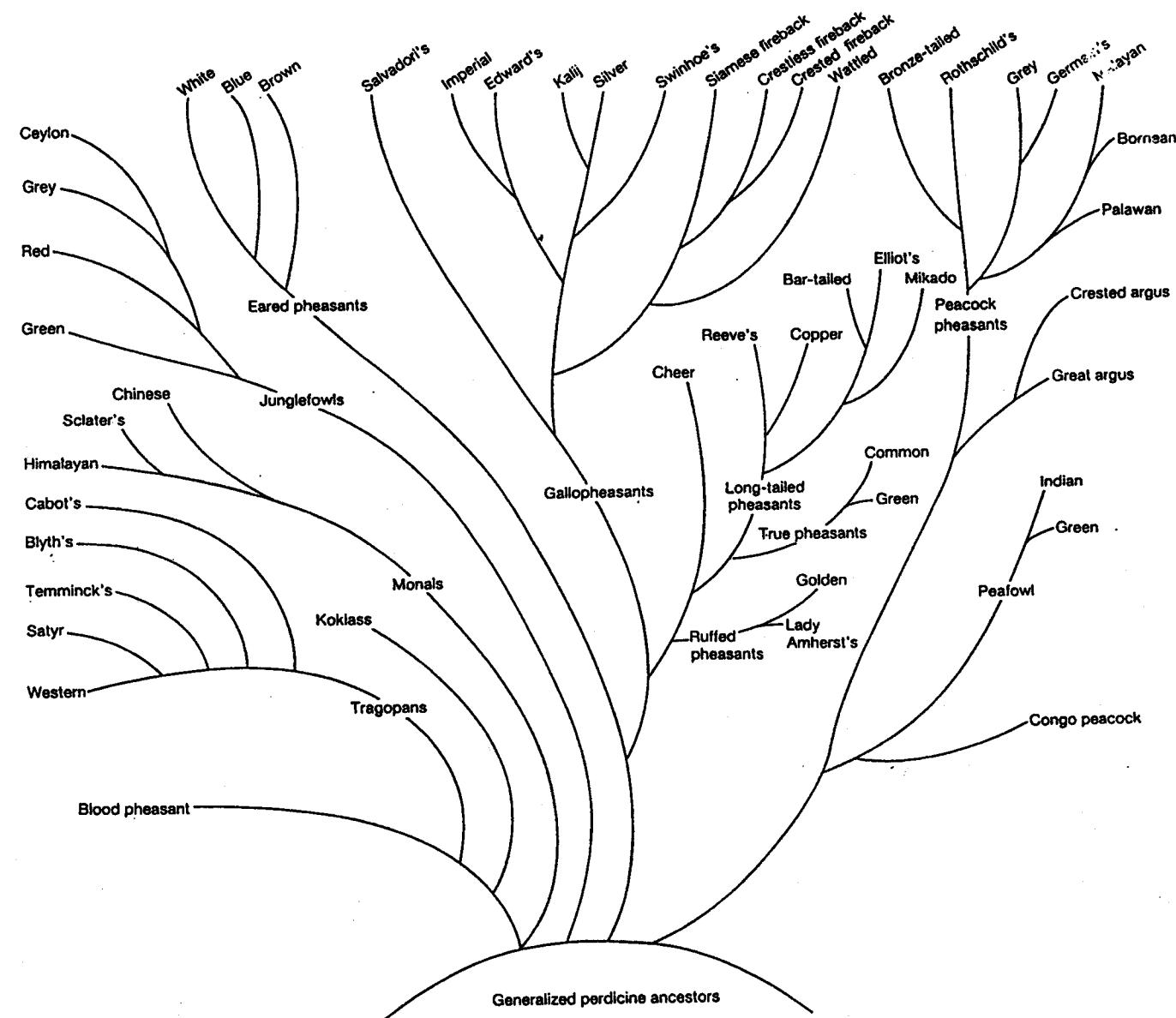


Figure 2-3.

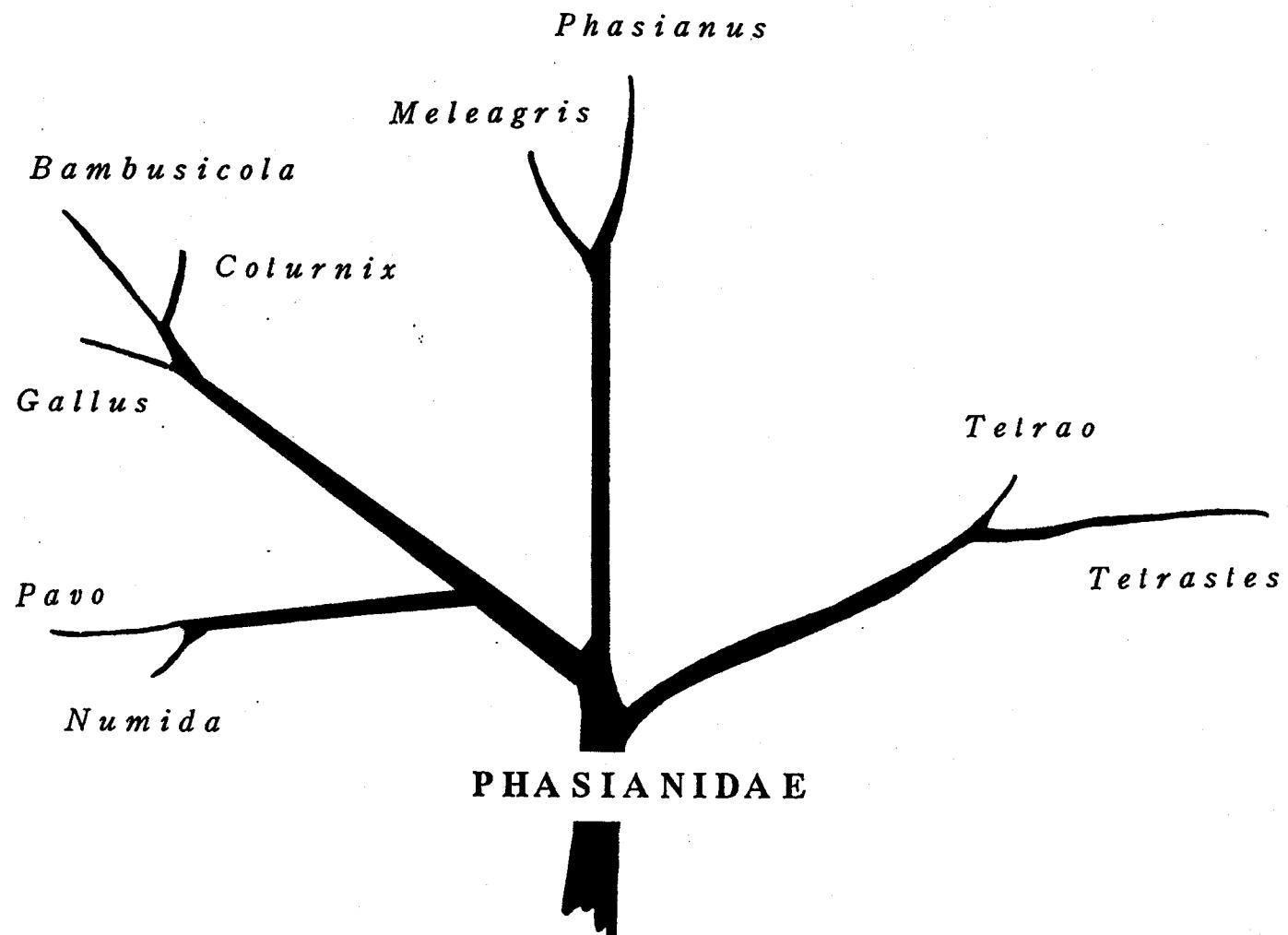


Figure 2-4.

RFPL

- 1) AYAM PELUNG #76 (VIII) AATTTTATTTTAACCTA**ACTCCCTACTAAGTGTACCCCCCTTCCCCCAGGGGGGGTATACTATGCATAATCGTCATA**CATTATAT 94
 2) NAGOYA # 1 (I) AATTTTATTTTAACCTA**ACTCCCTACTAAGTGTACCCCCCTTCCCCCAGGGGGGGTATACTATGCATAATCGTCATA**CATTATAT 94
 3) BARRED PLYMOUTH ROCK # 1 (V) AATTTTATTTTAACCTA**ACTCCCTACTAAGTGTACCCCCCTTCCCCCAGGGGGGGTATACTATGCATAATCGTCATA**CATTATAT 94
 4) THAI RED JUNGLEFOWL #11 (V) AATTTTATTTTAACCTA**ACTCCCTACTAAGTGTACCCCCCTTCCCCCAGGGGGGGTATACTATGCATAATCGTCATA**CATTATAT 94
 5) THAI RED JUNGLEFOWL # 3 (VII) AATTTTATTTTAACCTA**ACTCCCTACTAAGTGTACCCCCCTTCCCCCAGGGGGGGTATACTATGCATAATCGTCATA**CATTATAT 94
 6) INDN. RED JUNGLEFOWL #15 (VIII) AATTTTATTTTAACCTA**ACTCCCTACTAAGTGTACCCCCCTTCCCCCAGGGGGGGTATACTATGCATAATCGTCATA**CATTATAT 94
 7) GREEN JUNGLEFOWL #32 (C) AATTTTATTTTAACCT**AACTCCCTACTAAGTGTACCCCCCTTCCCCCAGGGGGGGTATACTATGCATAATCGTCATA**CATTATAT 94
 8) GREEN JUNGLEFOWL # 2 (D) AATTTTATTTTAACCT**AACTCCCTACTAAGTGTACCCCCCTTCCCCCAGGGGGGGTATACTATGCATAATCGTCATA**CATTATAT 94
 9) GREEN JUNGLEFOWL #50 (E) AATTTTATTTTAACCT**AACTCCCTACTAAGTGTACCCCCCTTCCCCCAGGGGGGGTATACTATGCATAATCGTCATA**CATTATAT 94
 10) JAPANESE QUAIL REF. AACACT-TTTTTAACCTA**ACTCCCTACTAAGTGTACCCCCCTTCCCCCAGGGGGGGTATACTATGCATAATCGTCATA**CATTATAT 93

VSP

- 1) DMB (VIII) ACCACATATATTGTA**CCGGTAATACTATGACTAAACCCATTATGTATACTGGCATTAACTATATTCCACATTCTCCAA**TGTCCATTCTATGCATG 206
 2) DMB (I) ACCACATATATTGTA**CCGGTAATACTATGACTAAACCCATTATGTATACTGGCATTAACTATATTCCACATTCTCCAA**TGTCCATTCTATGCATG 206
 3) DMB (V) ACCACATATATTGTA**CCGGTAATACTATGACTAAACCCATTATGTATACTGGCATTAACTATATTCCACATTCTCCAA**TGTCCATTCTATGCATG 206
 4) T RJF (V) ACCACATATATTGTA**CCGGTAATACTATGACTAAACCCATTATGTATACTGGCATTAACTATATTCCACATTCTCCAA**TGTCCATTCTATGCATG 206
 5) T RJF(VII) ACCACATATATTGTA**CCGGTAATACTATGACTAAACCCATTATGTATACTGGCATTAACTATATTCCACATTCTCCAA**TGTCCATTCTATGCATG 206
 6) I RJF(VIII) ACCACATATATTGTA**CCGGTAATACTATGACTAAACCCATTATGTATACTGGCATTAACTATCTCCAA**TGTCCATTCTATGCATG 206
 7) GJF (C) ACCACATATATTGTA**CCGGTAATACTATGACTAAACCCATTATGTATACTGGCATTAACTATCTCCAA**TGTCCATTCTATGCATG 206
 8) GJF (D) ACCACATATATTGTA**CCGGTAATACTATGACTAAACCCATTATGTATACTGGCATTAACTATCTCCAA**TGTCCATTCTATGCATG 206
 9) GJF (E) ACCACATATATTGTA**CCGGTAATACTATGACTAAACCCATTATGTATACTGGCATTAACTATCTCCAA**TGTCCATTCTATGCATG 206
 10) J QU AACACATATATTGTA**CCGGTAATACTATGACTAAACCCATTATGTATACTGGCATTAACTATCTCCAA**TGTCCATTCTATGCATG 203

ALU

- 1) DMB (VIII) ATCCAGGACAT-**AC-TTACCTCCCCTAGACAGCT** 244
 2) DMB (I) ATCCAGGACAT-**AC-CCATTACCCCTCCCATAGACAGCT** 244
 3) DMB (V) ATCCAGGACAT-**AC-TCATTCACCCCTCCCATAGACAGCT** 244
 4) T RJF (V) ATCCAGGACAT-**AC-TCATTCACCCCTCCCATAGACAGCT** 244
 5) T RJF(VII) ATCCAGGACAT-**AC-TCATTCACCCCTCCCATAGACAGCT** 244
 6) I RJF(VIII) ATCCAAGTCAT-**TC-TTACCTACATAGACAGCT** 244
 7) GJF (C) ATCCAAGTCAT**ACATC-GTC-C-TACCCATATACTAA**CT 243
 8) GJF (D) ATCCAAGTCAT**ACATC-GTC-C-TACCCATATACTAA**CT 243
 9) GJF (E) ATCCAAGTCAT**ACATC-GTC-C-TACCCATATACTAA**CT 243
 10) J QU CTCGAAGACAT-**AAACCATAC-GTTCACCTAGTAATAGA-** 240

NE

1ST COPY

- CCAAACCACTACCAAGTC**ACCTAACTATGAATGGTTACAGGACATAAATCTCACTCTCATGCT** 307
 CCAAACCACTACCAAGTC**ACCTAACTATGAATGGTTACAGGACATAAATCTCACTCTCATGCT** 307
 CCAAACCACTACCAAGTC**ACCTAACTATGAATGGTTACAGGACATAAATCTCACTCTCATGCT** 307
 CCAAACCACTACCAAGTC**ACCTAACTATGAATGGTTACAGGACATAAATCTCACTCTCATGCT** 307
 CCAAACCACTACCAAGTC**ACCTAACTATGAATGGTTACAGGACATAAATCTCACTCTCATGCT** 307
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 CCAAACCACTACCAAGTC**ACCTAACTATGAATGGTTACAGGACATAAATCTCACTCTCATGCT** 306
 CCAAACCACTACCAAGTC**ACCTAACTATGAATGGTTACAGGACATAAATCTCACTCTCATGCT** 306

2ND EXTRA COPY

- 7) GJF (C) CTACCCCTAACAGGT**ACCTAACTATGAATGGTTACAGGACATACTCTAATGTAGTGCT** 367
 9) GJF (E) CTACCCCTAACAGGT**ACCTAACTATGAATGGTTACAGGACATACTCTAATGTAGTGCT** 367

3RD EXTRA COPY

- 7) GJF (C) -----
 9) GJF (E) CTACCCCTAACAGGT**ACCTAACTATGAATGGTTACAGGACATACTCTAATATTAGTGCT** 428

MboII

ORIGINAL

- 1) DMB (VIII) CTCCCCC-AACAGTCACC-**TAACATGAATGGTTACAGGACATACATTAACTACCATGTT** 368 -CTAACCCATTGGTTATG**CTCG-CCGTATCAG** 399
 2) DMB (I) CTCCCCC-AACAGTCACC-**TAACATGAATGGTTACAGGACATACATTAACTACCATGTT** 368 -CTAACCCATTGGTTATG**CTCG-CCGTATCAG** 399
 3) DMB (V) CTCCCCC-AACAGTCACC-**TAACATGAATGGTTACAGGACATACATTAACTACCATGTT** 368 -CTAACCCATTGGTTATG**CTCG-CCGTATCAG** 399
 4) T RJF(V) CTCCCCC-AACAGTCACC-**TAACATGAATGGTTACAGGACATACATTAACTACCATGTT** 368 -CTAACCCATTGGTTATG**CTCG-CCGTATCAG** 400
 5) T RJF(VII) CTCCCCC-AACAGTCACC-**TAACATGAATGGTTACAGGACATACATTAACTACCATGTT** 368 -CTAACCCATTGGTTATG**CTCG-CCGTATCAG** 399
 6) I RJF(VIII) CTCCCCC-AACAGTCACC-**TAACATGAATGGTTACAGGACATACATTAACTACCATGTT** 367 -CTAACCCATTGGTTATG**CTCG-CCGTATCAG** 398
 7) GJF (C) CTACCCCT-AACAGGT**ACCTAACTATGAATGGTTACAGGACATACATTAACTACCATGTT** 430 -CTAACCCATTGGTTATG**CTCG-CCGTACAG** 456
 8) GJF (D) CTACCCCT-AACAGGT**ACCTAACTATGAATGGTTACAGGACATACATTAACTACCATGTT** 367 -CTAACCCATTGGTTATG**CTCG-CCGTACAG** 398
 9) GJF (E) CTACCCCT-AACAGGT**ACCTAACTATGAATGGTTACAGGACATACATTAACTACCATGTT** 489 -CTAACCCATTGGTTATG**CTCG-CCGTACAG** 516
 10) J QU CTTCCACTAACAGGACACC**TAACATGAATGGTTACAGGACATAAGCTTA-CTA-AATCT** 302 TAGCTCCATTGGTTATG**TAG-ACGTACAG** 334

Figure 3-1.

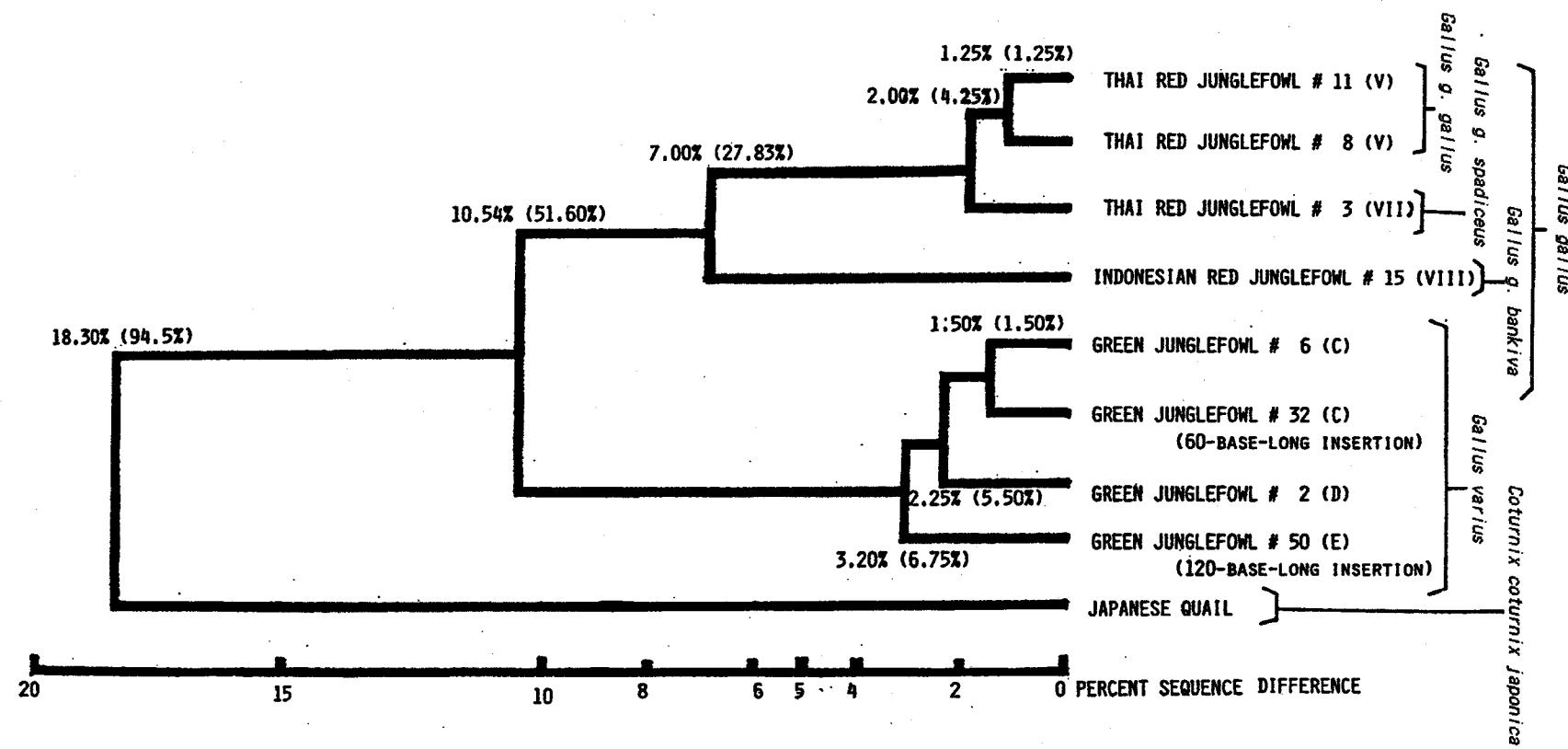


Figure 3-2.

	167	171	210	217	220	221	225	243	246	254	256	261	265	281	282	306	309	310	315	317	327	342	391'	394'
THAI RED JUNGLEFOWL # 8 (V)	T	C	C	T	T	C	C	C	T	C	G	T	C	T	T	C	A	T	G	-	-			
THAI RED JUNGLEFOWL #11 (V)	T	T	C	T	T	C	C	C	T	C	T	C	G	C	C	T	T	C	A	C	A	T	-	
AYAM CEMANI (V)	T	T	C	T	T	C	C	C	T	C	T	C	G	C	C	T	T	C	A	T	A	-	-	
BARRED PLYMOUTH ROCK (V)	T	T	C	C	T	C	C	C	T	C	T	C	A	C	T	T	T	C	A	T	A	-	-	
WHITE LEGHORN(HIROSHIMA VAR.) (V)	T	T	C	C	T	C	C	C	T	T	C	T	C	A	C	T	T	T	C	A	T	A	-	
WHITE LEGHORN(HIROSHIMA VAR.) (IV)	T	T	C	C	T	C	C	T	C	T	C	T	C	A	C	T	T	T	C	A	T	A	-	
THAI RED JUNGLEFOWL # 3 (VII)	T	C	C	T	T	C	C	C	C	C	T	T	A	C	T	T	T	T	C	T	G	-	-	
AYAM PELUNG (VIII)	T	T	C	T	T	T	C	C	C	T	C	T	C	G	C	C	C	T	C	A	T	A	-	
WHITE LEGHORN (I)	C	T	T	T	T	C	T	T	C	T	T	C	C	A	C	T	T	T	C	C	A	T	A	
NAGOYA (I)	C	T	C	T	C	C	T	T	C	T	T	C	C	A	C	T	T	C	C	A	T	A	-	
THAI RED JUNGLEFOWL # 8 (V)																								
THAI RED JUNGLEFOWL #11 (V)																								
THAI RED JUNGLEFOWL # 8 (V)																								
THAI RED JUNGLEFOWL # 3 (VII)																								
THAI RED JUNGLEFOWL #11 (V)																								
THAI RED JUNGLEFOWL # 3 (VII)																								
THAI RED JUNGLEFOWL # 8 (V)																								
AYAM CEMANI (V)																								
THAI RED JUNGLEFOWL #11 (V)																								
BARRED PLYMOUTH ROCK (V)																								
THAI RED JUNGLEFOWL # 8 (V)																								
AYAM PELUNG (VIII)																								
THAI RED JUNGLEFOWL # 3 (VII)																								
WHITE LEGHORN (I)																								
NAGOYA (I)																								
THAI RED JUNGLEFOWL # 8 (V)																								
THAI RED JUNGLEFOWL #11 (V)																								
THAI RED JUNGLEFOWL # 8 (V)																								
AYAM CEMANI (V)																								
THAI RED JUNGLEFOWL #11 (V)																								
AYAM PELUNG (VIII)																								
THAI RED JUNGLEFOWL # 8 (V)																								
AYAM CEMANI (V)																								
BARRED PLYMOUTH ROCK (V)																								
THAI RED JUNGLEFOWL # 3 (VII)																								
WHITE LEGHORN (I)																								
AYAM PELUNG (VIII)																								

Figure 3-3.

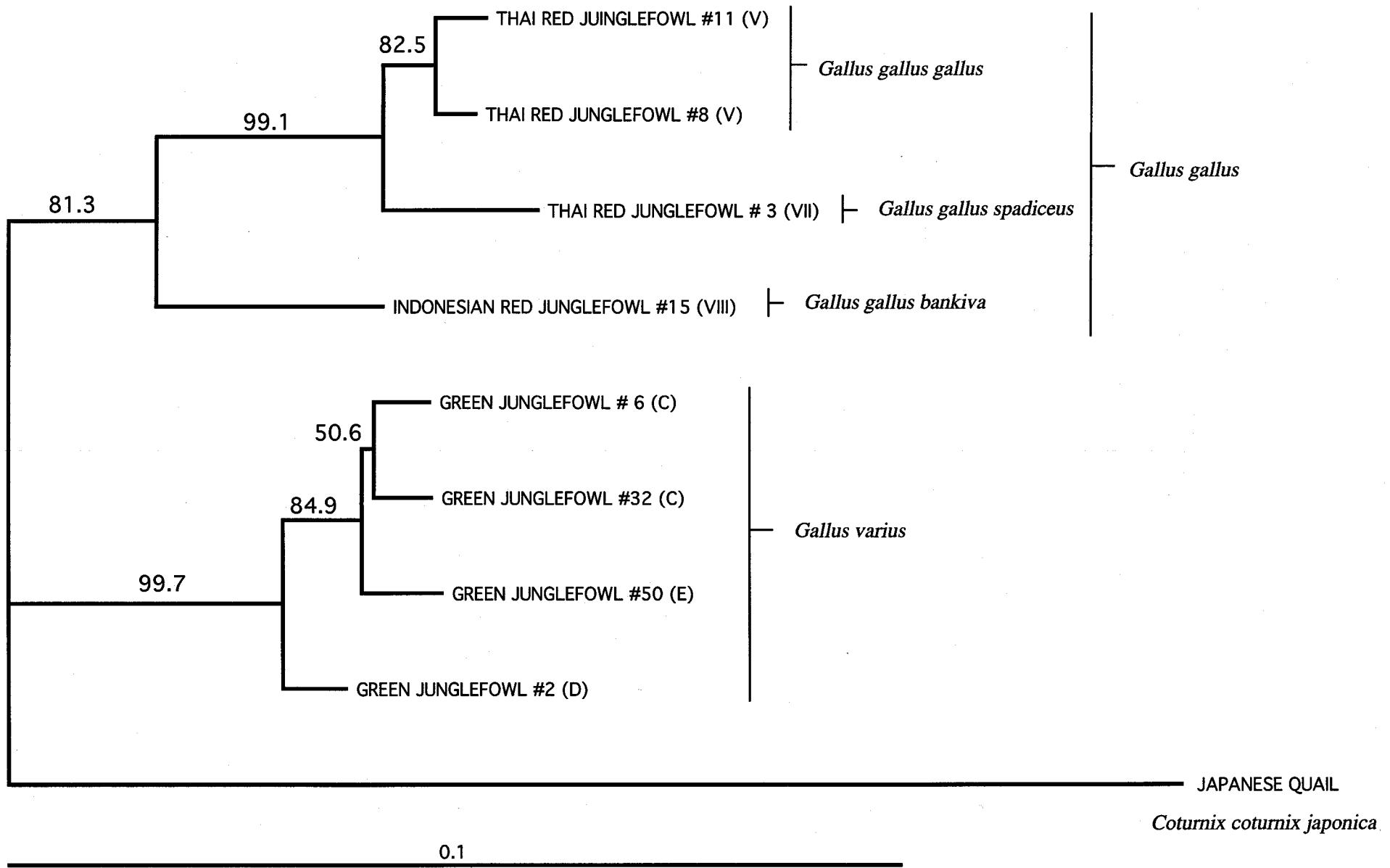


Figure 3-4.

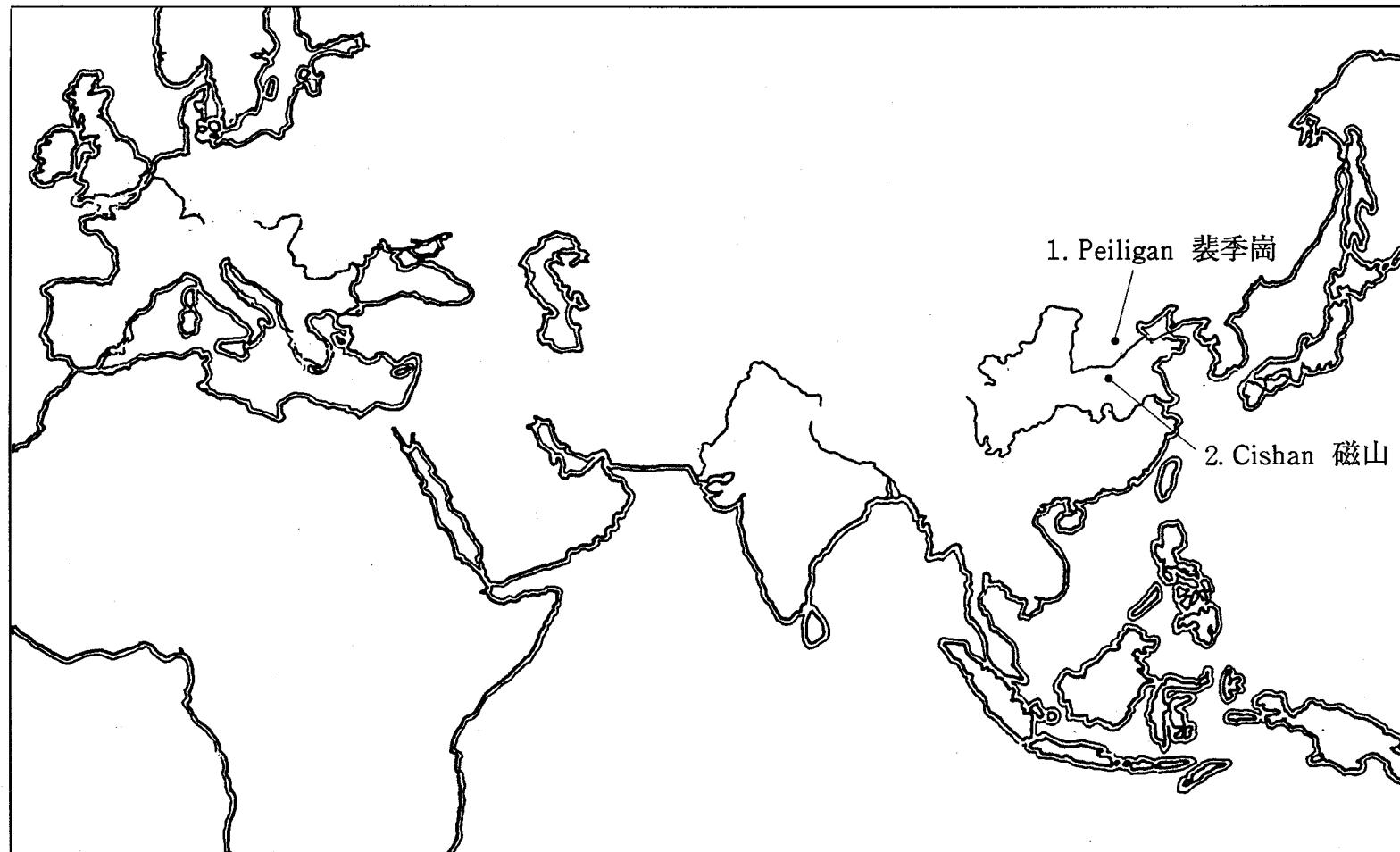


Figure 3-5.

Figure 4-1.

361	GG_bankiva15	-TTAACTAC C-ATGATTCT AACCCATTG GTTATGCTCG T-GTATCAGA T-----	420	GG_bankiva15	-----
GG_bankiva18	-TTAACTAC C-ATGATTCT AACCCATTG GTTATGCTCG T-GTATCAGA TGATTATT		GG_bankiva18	-----	
GG_bankiva19	-TTAACTAC C-ATGATTCT AACCCATTG GTTATGCTCG T-GTATCAGA TGATTATT		GG_bankiva19	-----	
GG_gallus11	-TTAACTAC C-ATG-TTCT AACCCATTG GTTATGCTCG TCGTATCAGA TGATTATT		GG_gallus11	-----	
GG_gallus39	-CTTAACAC C-ATG-TTCT AACCCATTG GTTATGCTCG TCGTATCAGA TGATTATT		GG_gallus39	TCAGGCCCAT TCTTCCCCC TACACCCCTC GCCCTACTTG -----	
GG_gallus41	-CTTAACAC C-ATG-TTCT AACCCATTG GTTATGCTCG TCGTATCAGA TGATTATT		GG_gallus41	TCAGGCCCAT TCTTCCCCC TACACCCCTC GCCCTACTT-----	
GG_gallus58	-CTTAACAC C-ATG-TTCT AACCCATTG GTTATGCTCG TCGTATCAGA TGATTATT		GG_gallus58	-----	
GG_gallus8	-TTAACTAC C-ATG-TTCT AACCCATTG GTTATGCTCG CGTATCAGA TGATTATT		GG_gallus8	-----	
GG_gallus10	-TTAACTAC T-ATG-TTCT AACCCATTG GTTATGCTCG CGTATCAGA TGATTATT		GG_gallus10	TCAGGCCCAT TCTTCCCCC TACACCCCTC GCCCTACTTG CCTTCAC-----	
GG_spadiceus3	-TTAACTAC C-ATG-TTCT AACCCATTG GTTATGCTCG CGTATCAGA TGATTATT		GG_spadiceus3	-----	
GG_spadiceus4	-TTAACTAC C-ATG-TTCT AACCCATTG GTTATGCTCG CGTATCAGA TGATTATT		GG_spadiceus4	TCAGGCCCAT TCTTCCCCC TACACCCCTC GCCCTACTTG CCTTCACC-----	
GG_spadiceus5	-TTAACTAC C-ATG-TTCT AACCCATTG GTTATGCTCG CGTATCAGA TGATTATT		GG_spadiceus5	TCAGGCCCAT TCTTCCCCC TACACCCCTC GCCCTA-----	
G_lafayettei1	CTCCAACTCT T-ATG-TTCT AACCCATTG GTTATGCTCG TCGTATCAGA TGATTATT		G_lafayettei1	TCAGGCCCAT TCTTCCCCC TACACCCCTC GCCCTACTTG CCTTCAC-----	
G_lafayettei2	CTCCAACTCT T-ATG-TTCT AACCCATTG GTTATGCTCG TCGTATCAGA TGATTATT		G_lafayettei2	-----	
G_sonneratii	CCTAAATCT T-ATG-TTCT AACCCATTG GTTATGCTCG TCGTATCAGA TGATTATT		G_sonneratii	TCAGGCCCAT TCTTCCCCC TACACCCCTC GCCCTACTTG CCTTCAC-----	
G_varius2	-TTAACTAC C-ATG-ATCT AACCCATTG GTTATGCTCG TCGTACAGA TGATTATT		G_varius2	-----	
G_varius32	-TCTAACTAC C-ATG-ATCT AACCCATTG GTTATGCTCG TCGTACAGA TGATTATT		G_varius32	-----	
G_varius50	-TCTAACTAC C-ATG-ATCT AACCCATTG GTTATGCTCG T-GTACAGT-----		G_varius50	-----	
G_varius6	-TCTAACTAC C-ATG-ATCT AACCCATTG GTTATGCTCG TGTACACAGA TGATTATT		G_varius6	TCAGGCC-AT TCTTCCCC- TACACCCCTG CCTACTTG-----	
Cemani	-TTAACTAC C-ATG-TTCT AACCCATTG GTTATGCTCG CGTATCAGA TGATTATT		Cemani	TCAGGCCAT TCTT-----	
Ayam63	-TTAACTAC C-ATG-TTCT AACCCATTG GTTATGCTCG CGTATCAGA TGATTATT		Ayam63	-----	
Ayam71	-TTAACTAC C-ATG-TTCT AACCCATTG GTTATGCTCG CGTATCAGA TGATTATT		Ayam71	-----	
Pelung	-TTAACTAC C-ATG-TTCT AACCCATTG GTTATGCTCG CGTATCAGA TGATTATT		Pelung	-----	
Bantam8	-TTAACTAC C-ATG-TTCT AACCCATTG GTTATGCTCG CGTATCAGA TGATTATT		Bantam8	TCAGGCC-----	
Barred_Plymous	-TTAACTAC C-ATG-TTCT AACCCATTG GTTATGCTCG CGTATCAGA CGATTATT		Barred_Plymous	TCAGGCCAT TCTTCCCCC TACACCC-----	
Nagoya	-TTAACTAC C-ATG-TTCT AACCCATTG GTTATGCTCG CGTATCAGA CGATTATT		Nagoya	TCAGGCCAT TCTTCCCCC TACACCC-----	
White_Leghorn1	-TTAACTAC C-ATG-TTCT AACCCATTG GTTATGCTCG CGTATCAGA TGATTATT		White_Leghorn1	TCAGGCCAT TCTTCCCCC TACACCCCTC GCCCTACTTG CCTTC-----	
White_Leghorn2	-TTAACTAC C-ATG-TTCT AACCCATTG GTTATGCTCG CGTATCAGA -----		White_Leghorn2	-----	
White_Leghorn3	-TTAACTAC C-ATG-TTCT AACCCATTG GTTATGCTCG CGTATCAGA -----		White_Leghorn3	-----	
C_coturnix	-CTTACTAAA TACTT--AGC TCCCCATTG GTTATGCTAG AGCTACAGA TGATTATT	//	C_coturnix	TCAGGCCAT TCTTCCCCC TACACCCCTC GCCCCCTTG CCTCTGGTC	
421	GG_bankiva15	-----	480	GG_bankiva15	-----
GG_bankiva18	GATCGTTCAC CTACGGAGAG AGCGCAACC CCTG-----		GG_bankiva18	-----	
GG_bankiva19	GATCGTTCAC CTACG-----		GG_bankiva19	-----	
GG_gallus11	G-----		GG_gallus11	-----	
GG_gallus39	GATCGTTCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC		GG_gallus39	GATCGTTCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC	
GG_gallus41	GATCGTTCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC		GG_gallus41	GATCGTTCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC	
GG_gallus58	GATCGTTCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC		GG_gallus58	GATCGTTCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC	
GG_gallus8	GATCGTTCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC		GG_gallus8	GATCGTTCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC	
GG_gallus10	GATCGTTCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC		GG_gallus10	GATCGTTCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC	
GG_spadiceus3	-----		GG_spadiceus3	-----	
GG_spadiceus4	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC		GG_spadiceus4	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC	
GG_spadiceus5	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC		GG_spadiceus5	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC	
G_lafayettei1	GATCGTTCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC		G_lafayettei1	GATCGTTCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC	
G_lafayettei2	GATCG-----		G_lafayettei2	GATCG-----	
G_sonneratii	GATCGTTCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC		G_sonneratii	GATCGTTCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC	
G_varius2	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC-----		G_varius2	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC-----	
G_varius32	GATCG-----		G_varius32	GATCG-----	
G_varius50	-----		G_varius50	-----	
G_varius6	GATCGTC-AC CTACGGAGAG ATCAGCAACC CCTGCC-GTA ATG-TACTT- ATGACCACTC		G_varius6	GATCGTC-AC CTACGGAGAG ATCAGCAACC CCTGCC-GTA ATG-TACTT- ATGACCACTC	
Cemani	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC		Cemani	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCTGTA ATG-TACTTC ATGACCACTC	
Ayam63	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCT-----		Ayam63	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCT-----	
Ayam71	GATCGTCCAC CTACGGAG-----		Ayam71	GATCGTCCAC CTACGGAG-----	
Pelung	-----		Pelung	-----	
Bantam8	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCC-GTA ATG-TACTTC ATGACCACTC		Bantam8	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCC-GTA ATG-TACTTC ATGACCACTC	
Barred_Plymous	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCC-GTA ATG-TACTTC ATGACCACTC		Barred_Plymous	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCC-GTA ATG-TACTTC ATGACCACTC	
Nagoya	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCC-GTA ATG-TACTTC ATGACCACTC		Nagoya	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCC-GTA ATG-TACTTC ATGACCACTC	
White_Leghorn1	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCC-GTA ATG-TACTTC ATGACCACTC		White_Leghorn1	GATCGTCCAC CTACGGAGAG ATCAGCAACC CCTGCC-GTA ATG-TACTTC ATGACCACTC	
White_Leghorn2	-----		White_Leghorn2	-----	
White_Leghorn3	-----		White_Leghorn3	-----	
C_coturnix	GATCGTACAC CTACGGAGAG ATCAGCAACC CCTGCC-GTA ATGCTATTCC GTGACTAGCT		C_coturnix	GATCGTACAC CTACGGAGAG ATCAGCAACC CCTGCC-GTA ATGCTATTCC GTGACTAGCT	

Figure 4-1. (continued)

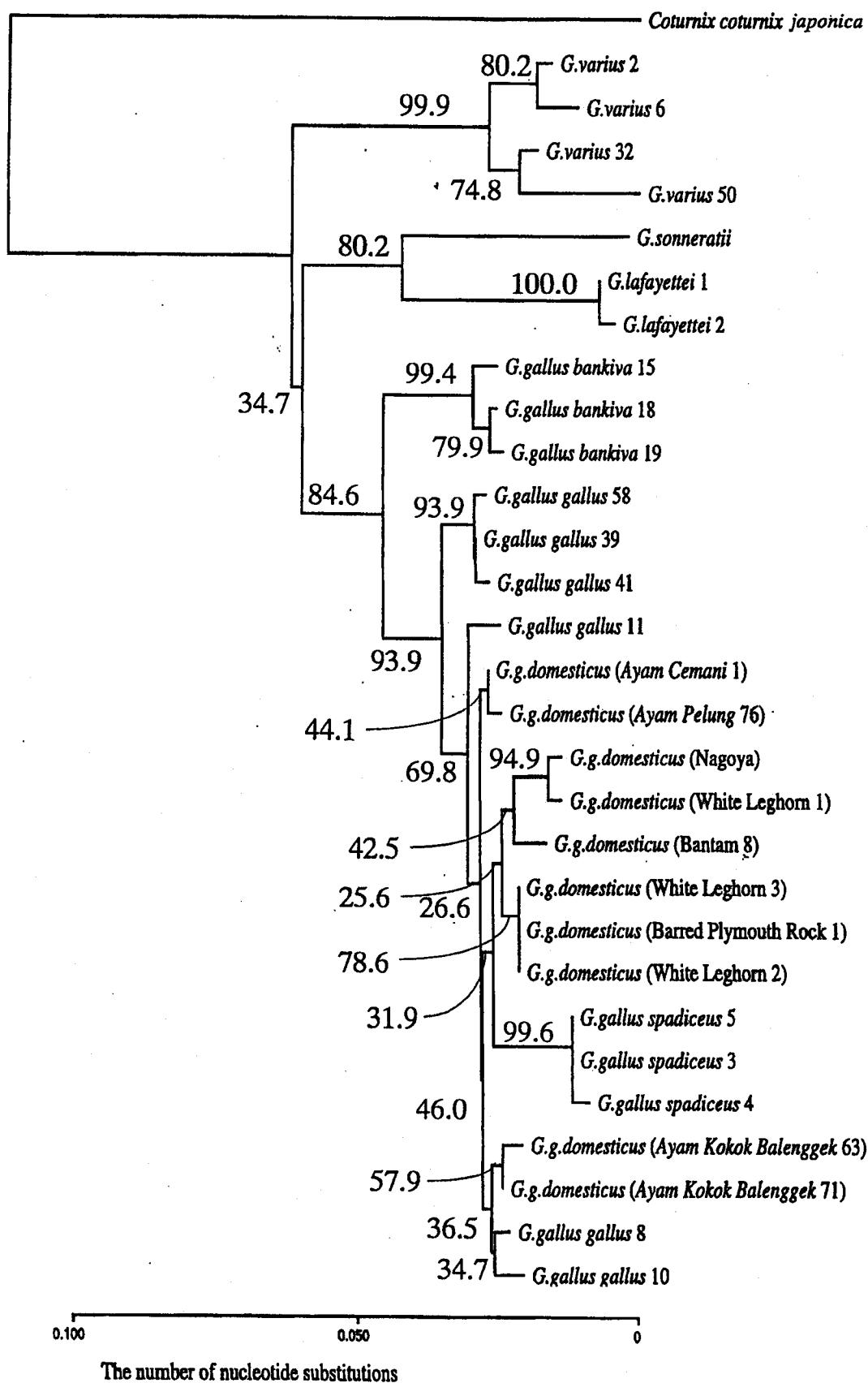


Figure 4-2.

Appendix A

**Nucleotide Sequences Determined
in this Study**

LOCUS CHKMTB01 400 bp ds-DNA VRT 17-JUN-1996
 DEFINITION Red Junglefowl mitochondrial DNA for D-loop region.
 ACCESSION D82897
 KEYWORDS D-loop region.
 SOURCE Gallus gallus (sub_species:bankiva, isolate:15) mitochondrial DNA.
 ORGANISM Gallus gallus
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
 Neognathae; Galliformes; Phasianidae.
 REFERENCE 1 (bases 1 to 400)
 AUTHORS Fumihiro,A.
 TITLE Direct Submission
 JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
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 (E-mail:nkojima@lab.nig.ac.jp)
 STANDARD full staff_review
 REFERENCE 2 (bases 1 to 400)
 AUTHORS Fumihiro,A.
 JOURNAL Unpublished (1996)
 STANDARD full staff_review
 COMMENT
 FEATURES Location/Qualifiers
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 /mitochondrion
 /organism="Gallus gallus"
 /sequenced_mol="DNA"
 /sub_species="bankiva"
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 61 ggtatactat gcataatcgt gcatacattt atataccca tatattttgg taccggtaat
 121 atatactata tatgtactaa acccattaa tgatatacggg cattaatcta catccctcat
 181 ttctcccaat gtccatccca tgcatgatcc aagtcatctt tagtcataattt cccataaagc
 241 agctctoacc actaccaaga cacctaacta tgaatggta caggacataa ctcttactct
 301 catgttcctc ccccaacaag tcacctaact atgaatggtt acaggacatac catttaacta
 361 ccatgattct aaccatgg tgatgtctcg tgcatacgat
 //
 LOCUS CHKMTB02 442 bp ds-DNA VRT 17-JUN-1996
 DEFINITION Red Junglefowl mitochondrial DNA for D-loop region.
 ACCESSION D82898
 KEYWORDS D-loop region.
 SOURCE Gallus gallus (sub_species:bankiva, isolate:18) mitochondrial DNA.
 ORGANISM Gallus gallus
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
 Neognathae; Galliformes; Phasianidae.
 REFERENCE 1 (bases 1 to 442)
 AUTHORS Fumihiro,A.
 TITLE Direct Submission
 JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
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 STANDARD full staff_review
 REFERENCE 2 (bases 1 to 442)
 AUTHORS Fumihiro,A.
 JOURNAL Unpublished (1996)
 STANDARD full staff_review
 COMMENT
 FEATURES Location/Qualifiers
 source 1..442
 /isolate="18"
 /mitochondrion
 /organism="Gallus gallus"
 /sequenced_mol="DNA"
 /sub_species="bankiva"
 BASE COUNT 129 a 117 c 58 g 138 t 0 others
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 61 ggtatactat gcataatcgt gcatacattt atataccca tatattttgg taccggtaat
 121 atatactata tatgtactaa acccattaa tgatatacggg cattaatcta catccctcat
 181 ttctcccaat gtccatccca tgcatgatcc aagtcatctt tagtcataattt cccataaagc
 241 gctctaccca ctaccaagac acctaactat gaatggtac aggacataac tctcacttc
 301 atgttttcc cccaaacaatg caccctaacta tgaatggtt caggacatac atttaactat
 361 catgatttca acccatgg tgatgtctgt gtatcgatg gatttttgta tggttcacct
 421 cacgagagag cagcaaaaaa tg

Locus CHKMTB03 424 bp ds-DNA VRT 17-JUN-1996
 Definition Red Junglefowl mitochondrial DNA for D-loop region.
 Accession D82899
 Keywords D-loop region.
 Source Gallus gallus (sub_species:bankiva, isolate:19) mitochondrial DNA.
 Organism Gallus gallus
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
 Neognathae; Galliformes; Phasianidae.
 Reference 1 (bases 1 to 424)
 Authors Fumihiro,A.
 Title Direct Submission
 Journal Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
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 Reference 2 (bases 1 to 424)
 Authors Fumihiro,A.
 Journal Unpublished (1996)
 Standard full staff_review
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 Features Location/Qualifiers
 source 1..424
 /isolate="19"
 /mitochondrion
 /organism="Gallus gallus"
 /sequenced_mol="DNA"
 /sub_species="bankiva"
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 121 atatactata tatgtactaa acccattata tgatatacggt cattaatcta cattccatcat
 181 ttctcccaat gtccattcca tgcattgtcc aagtcatctt agtcatatcc cccataatga
 241 gcttaacca ctaccaagac actaactat gaatgttac aggacataac tcttacttc
 301 atgttctcc cccaaacaatg cacctaacta tgaatgttg caggacatac atttaaatct
 361 catgattcta acccatttg ttatgtcggt gtatcatatg gattttatga tgcgttccat
 421 caac
 //
 Locus CHKMTG04 448 bp ds-DNA VRT 17-JUN-1996
 Definition Red Junglefowl mitochondrial DNA for D-loop region.
 Accession D82900
 Keywords D-loop region.
 Source Gallus gallus (sub_species:gallus, isolate:8) mitochondrial DNA.
 Organism Gallus gallus
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
 Neognathae; Galliformes; Phasianidae.
 Reference 1 (bases 1 to 448)
 Authors Fumihiro,A.
 Title Direct Submission
 Journal Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
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 Standard full staff_review
 Reference 2 (bases 1 to 448)
 Authors Fumihiro,A.
 Journal Unpublished (1996)
 Standard full staff_review
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 Features Location/Qualifiers
 source 1..448
 /isolate="8"
 /mitochondrion
 /organism="Gallus gallus"
 /sequenced_mol="DNA"
 /sub_species="gallus"
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 61 ggataactat gcataatcggt gcatacattt atataccaca tatattttgg taccggtaat
 121 atatactata tatgtactaa acccattata tgatatacggt cattaatcta cattccatcat
 181 ttctcccaat gtccattcca tgcattgtcc aaggacatact cattacccat ccccatatga
 241 agtcccaac cactaccaag tcaactact atgaatgggtt gttaggacata atctcactc
 301 tcatgtctt ccccaacaatg gtaccaaacat tgcattgttg tgcaggacat acattnaac
 361 accatgtttt aaccatattt gttatgtcggt cgatgtatca tgatgtttt gatcgttcc
 421 tcacgagaga tcagcaacc ctgcctgt
 //

LOCUS CHMGTG05 518 bp ds-DNA VRT 17-JUN-1996
 DEFINITION Red Junglefowl mitochondrial DNA for D-loop region.
 ACCESSION D82901
 KEYWORDS D-loop region.
 SOURCE Gallus gallus (sub_species:gallus, isolate:10) mitochondrial DNA.
 ORGANISM Gallus gallus
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
 Neognathae; Galliformes; Phasianidae.
 REFERENCE 1 (bases 1 to 518)
 AUTHORS Fumihiro,A.
 TITLE Direct Submission
 JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
 Akishinonomiya Fumihiro, Yamashina Institute for Ornithology; 115
 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan
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 STANDARD full staff_review
 REFERENCE 2 (bases 1 to 518)
 AUTHORS Fumihiro,A.
 JOURNAL Unpublished (1996)
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 FEATURES Location/Qualifiers
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 /mitochondrion
 /organism="Gallus gallus"
 /sequenced_mol="DNA"
 /sub_species="gallus"
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 181 ttctcccaat gtccatctt tgcatacgtt aggacatctt cattacccctt ccccatagac
 241 agctccaaac cactaccagg taacctactt atgaatggt gcaggacata aatttcactc
 301 tcatgtctt ccccaacaa gtcaccaaac tatgaatggt tgccggacat acatthaact
 361 attatgtttt aaccatttg gtatgtctg cctgtatcaga tggattttt atgtgtccac
 421 ctcacggagat atcacaacc ctgtgttgc atgtacttca tgaccgtct caggccatt
 481 ctttccccctt acacccttgtt ccctacttgc ctccaccc
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 LOCUS CHMGTG06 412 bp ds-DNA VRT 17-JUN-1996
 DEFINITION Red Junglefowl mitochondrial DNA for D-loop region.
 ACCESSION D82902
 KEYWORDS D-loop region.
 SOURCE Gallus gallus (sub_species:gallus) mitochondrial DNA, clone:11.
 ORGANISM Gallus gallus
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
 Neognathae; Galliformes; Phasianidae.
 REFERENCE 1 (bases 1 to 412)
 AUTHORS Fumihiro,A.
 TITLE Direct Submission
 JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
 Akishinonomiya Fumihiro, Yamashina Institute for Ornithology; 115
 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan
 (E-mail:nkojima@lab.nig.ac.jp)
 STANDARD full staff_review
 REFERENCE 2 (bases 1 to 412)
 AUTHORS Fumihiro,A.
 JOURNAL Unpublished (1996)
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 COMMENT
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 /sub_species="gallus"
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 repeat_unit 245..307
 repeat_unit 308..368
 BASE COUNT 120 a 112 c 52 g 128 t 0 others
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 121 atatactata tatgtactaa accattata tgatacggg cattaatcta tattccacat
 181 ttctcccaat gtccatctt tgcatacgtt aggacatctt cattacccctt ccccatagac
 241 agctccaaac cactaccagg taacctactt atgaatggt gcaggacata aatttcactc
 301 tcatgtctt ccccaacaa gtcaccaaac tatgaatggt tgccggacat acatthaact
 361 accatgtttt aaccatttg gtatgtctg cctgtatcaga tggattttt atgtgtccac
 421 ctcacggagat atcacaacc ctgtgttgc atgtacttca tgaccgtct caggccatt
 481 ctttccccctt acacccttgtt ccctacttgc ctccaccc
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LOCUS CHKMTG07 508 bp ds-DNA VRT 17-JUN-1996
DEFINITION Red Junglefowl mitochondrial DNA for D-loop region.
ACCESSION D82903
KEYWORDS D-loop region.
SOURCE Gallus gallus (sub_species:gallus, isolate:39) mitochondrial DNA.
ORGANISM Gallus gallus
Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
Neognathae; Galliformes; Phasianidae.
REFERENCE 1 (bases 1 to 509)
AUTHORS Fumihiato,A.
TITLE Direct Submission
JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
Akishinonoya Fumihiato, Yamashina Institute for Ornithology; 115
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STANDARD full staff_review
REFERENCE 2 (bases 1 to 509)
AUTHORS Fumihiato,A.
JOURNAL Unpublished (1996)
STANDARD full staff_review
COMMENT
FEATURES Location/Qualifiers
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/mitochondrion
/organism="Gallus gallus"
/sequenced_mol="DNA"
/sub_species="gallus"
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121 atatactata tatgtactaa acccattata tgatatacggg cattaatcta tattccacat
181 ttctcccaat gtccatttc tgcattgtcc aagacatact cattaccctt ccccatat
241 agctccaaac cactaccgaq acacctaact atgaaatggt acaggacata aatctcaatc
301 tcattgtctt ccccaacaa gtcaccaac tatgaatggt tacaggacat atacttaact
361 accatgttct aaccatattt gttatgtcg tgcattatcgg tggattttt gatcgttac
421 ctcacgagag atcagaacc ctgtgtgtt atgtatctca tgaccgtct caggccatt
481 ctttccccct acacccctcg ccctacttg
//
LOCUS CHKMTG08 508 bp ds-DNA VRT 17-JUN-1996
DEFINITION Red Junglefowl mitochondrial DNA for D-loop region.
ACCESSION D82904
KEYWORDS D-loop region.
SOURCE Gallus gallus (sub_species:gallus, isolate:41) mitochondrial DNA.
ORGANISM Gallus gallus
Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
Neognathae; Galliformes; Phasianidae.
REFERENCE 1 (bases 1 to 508)
AUTHORS Fumihiato,A.
TITLE Direct Submission
JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
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(E-mail:nkojima@lab.nig.ac.jp)
STANDARD full staff_review
REFERENCE 2 (bases 1 to 508)
AUTHORS Fumihiato,A.
JOURNAL Unpublished (1996)
STANDARD full staff_review
COMMENT
FEATURES Location/Qualifiers
source 1..508
/isolate="41"
/mitochondrion
/organism="Gallus gallus"
/sequenced_mol="DNA"
/sub_species="gallus"
BASE COUNT 143 a 148 c 63 g 154 t 0 others
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121 atatactata tatgtactaa acccattata tgatatacggg cattaatcta tattccacat
181 ttctcccaat gtccatttc tgcattgtcc aagacatact cattaccctt ccccatat
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301 tcattgtctt ccccaacaa gtcaccaac tatgaatggt tacaggacat atacttaact
361 accatgttct aaccatattt gttatgtcg tgcattatcgg tggattttt gatcgttac
421 ctcacgagag atcagaacc ctgtgtgtt atgtatctca tgaccgtct caggccatt
481 ctttccccct acacccctcg ccctacttg
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LOCUS CHKMTG09 434 bp ds-DNA VRT 17-JUN-1996
DEFINITION Red Junglefowl mitochondrial DNA for D-loop region.
ACCESSION D82905
KEYWORDS D-loop region.
SOURCE Gallus gallus (sub_species:gallus, isolate:58) mitochondrial DNA.
ORGANISM Gallus gallus
Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
Neognathae; Galliformes; Phasianidae.
REFERENCE 1 (bases 1 to 434)
AUTHORS Fumihiito,A.
TITLE Direct Submission
JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
Akishinonoya Fumihiito, Yamashina Institute for Ornithology; 115
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STANDARD full staff_review
REFERENCE 2 (bases 1 to 434)
AUTHORS Fumihiito,A.
JOURNAL Unpublished (1996)
STANDARD full staff_review
COMMENT
FEATURES Location/Qualifiers
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/isolate="58"
/mitochondrion
/organism="Gallus gallus"
/sequenced_mol="DNA"
/sub_species="gallus"
BASE COUNT 130 a 116 c 54 g 134 t 0 others
ORIGIN
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121 atatactata tatgtactaa accattata tgatatacggg cattaatcta tattccacat
181 ttctcccaat gtccattcta tgcattgtaccc agagacatcattcaccc ccccatagac
241 agccccaaac cactaccaag acacccaaact atgaatggt acaggacata aatctcaatc
301 tcattgtttt ccccaacaa gtacccaaac tatgaatggt tacaggacat atactaaatc
361 actatgtttt aaccatgtt gttatgtctg tgcattacca tggattttt gatcgtttcc
421 ctcacgagag atca
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LOCUS CHKMTS10 400 bp ds-DNA VRT 17-JUN-1996
DEFINITION Red Junglefowl mitochondrial DNA for D-loop region.
ACCESSION D82906
KEYWORDS D-loop region.
SOURCE Gallus gallus (sub_species:spadiceus, isolate:3) mitochondrial DNA.
ORGANISM Gallus gallus
Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
Neognathae; Galliformes; Phasianidae.
REFERENCE 1 (bases 1 to 400)
AUTHORS Fumihiito,A.
TITLE Direct Submission
JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
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(E-mail:nkojima@lab.nig.ac.jp)
STANDARD full staff_review
REFERENCE 2 (bases 1 to 400)
AUTHORS Fumihiito,A.
JOURNAL Unpublished (1996)
STANDARD full staff_review
COMMENT
FEATURES Location/Qualifiers
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/isolate="3"
/mitochondrion
/organism="Gallus gallus"
/sequenced_mol="DNA"
/sub_species="spadiceus"
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repeat_unit 308..368
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61 ggatatactat gcataatcggt gcatacattt atataccaca tatattatgg taccggtaat
121 atatactata tatgtactaa accattata tgatatacggg cattaatcta tattccacat
181 ttctcccaat gtccattcta tgcattgtaccc agagacatcattcaccc ccccatagac
241 agctccaaac caccaccaag tcacttaact atgaatggt acaggacata aatctcaatc
301 tcattgtttt ccccaacaa gtacccaaac tatgaatggt tacaggacat atatataact
361 accatgtttt aaccatgtt gttatgtctg tgcaggacat atatataact
421 //

LOCUS CHKMTS11 518 bp ds-DNA VRT 17-JUN-1996
DEFINITION Red Junglefowl mitochondrial DNA for D-loop region.
ACCESSION D82907
KEYWORDS D-loop region.
SOURCE Gallus gallus (sub_species:spadiceus, isolate:4) mitochondrial DNA.
ORGANISM Gallus gallus
Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
Neognathae; Galliformes; Phasianidae.
REFERENCE 1 (bases 1 to 518)
AUTHORS Fumihiro,A.
TITLE Direct Submission
JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
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STANDARD full staff_review
REFERENCE 2 (bases 1 to 518)
AUTHORS Fumihiro,A.
JOURNAL Unpublished (1996)
STANDARD full staff_review
COMMENT
FEATURES Location/Qualifiers
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/isolate="4"
/mitochondrion
/organism="Gallus gallus"
/sequenced_mol="DNA"
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361 accatgttct aaccatgttgc ttatgtctcg ccgtatcaga tggtattttt gatgtccac
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LOCUS CHKMTS12 505 bp ds-DNA VRT 17-JUN-1996
DEFINITION Red Junglefowl mitochondrial DNA for D-loop region.
ACCESSION D82908
KEYWORDS D-loop region.
SOURCE Gallus gallus (sub_species:spadiceus, isolate:5) mitochondrial DNA.
ORGANISM Gallus gallus
Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
Neognathae; Galliformes; Phasianidae.
REFERENCE 1 (bases 1 to 505)
AUTHORS Fumihiro,A.
TITLE Direct Submission
JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
Akishinonoya Fumihiro, Yamashina Institute for Ornithology; 115
Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan
(E-mail:nkojima@lab.nig.ac.jp)
STANDARD full staff_review
REFERENCE 2 (bases 1 to 505)
AUTHORS Fumihiro,A.
JOURNAL Unpublished (1996)
STANDARD full staff_review
COMMENT
FEATURES Location/Qualifiers
source 1..505
/isolate="5"
/mitochondrion
/organism="Gallus gallus"
/sequenced_mol="DNA"
/sub_species="spadiceus"
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241 agtcctaaac caccaccaaag tcacttaact atgaatgggt acaggacatca aatctcaatc
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361 accatgttct aaccatgttgc ttatgtctcg ccgtatcaga tggtattttt gatgtccac
421 ctcacgagag atcagaaccc ctgtctgtt aatgtatcca tgaccgtttt caggccatt
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LOCUS	CHKMTL13	578 bp ds-DNA	VRT	17-JUN-1996	LOCUS	CHKMTL14	476 bp ds-DNA	VRT	17-JUN-1996										
DEFINITION	Cingalese mitochondrial DNA for D-loop region.				DEFINITION	Cingalese mitochondrial DNA for D-loop region.													
ACCESSION	D82909				ACCESSION	D82910													
KEYWORDS	D-loop region.				KEYWORDS	D-loop region.													
SOURCE	Gallus lafayettei (isolate:1) mitochondrial DNA.				SOURCE	Gallus lafayettei (isolate:2) mitochondrial DNA.													
ORGANISM	Gallus lafayettei Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.				ORGANISM	Gallus lafayettei Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.													
REFERENCE	1 (bases 1 to 578)				REFERENCE	1 (bases 1 to 476)													
AUTHORS	Fumihiro,A.				AUTHORS	Fumihiro,A.													
TITLE	Direct Submission				TITLE	Direct Submission													
JOURNAL	Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases. Akishinonomiya Fumihiro, Yamashina Institute for Ornithology; 115 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan (E-mail:nkojima@lab.nig.ac.jp)				JOURNAL	Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases. Akishinonomiya Fumihiro, Yamashina Institute for Ornithology; 115 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan (E-mail:nkojima@lab.nig.ac.jp)													
STANDARD	full staff_review				STANDARD	full staff_review													
REFERENCE	2 (bases 1 to 578)				REFERENCE	2 (bases 1 to 476)													
AUTHORS	Fumihiro,A.				AUTHORS	Fumihiro,A.													
JOURNAL	Unpublished (1996)				JOURNAL	Unpublished (1996)													
STANDARD	full staff_review				STANDARD	full staff_review													
COMMENT					COMMENT														
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LOCUS CHKMTS15 580 bp ds-DNA VRT 17-JUN-1996
 DEFINITION Grey Junglefowl mitochondrial DNA for D-loop region.
 ACCESSION D82911
 KEYWORDS D-loop region.
 SOURCE Gallus gallus (sub_species:sonnerati, isolate:1) mitochondrion DNA.
 ORGANISM Gallus gallus
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
 Neognathae; Galliformes; Phasianidae.
 REFERENCE 1 (bases 1 to 580)
 AUTHORS Fumihiro,A.
 TITLE Direct Submission
 JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
 Akishinonoya Fumihiro, Yamashina Institute for Ornithology; 115
 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan
 (E-mail:nkojimo@lab.nig.ac.jp)
 STANDARD full staff_review
 REFERENCE 2 (bases 1 to 580)
 AUTHORS Fumihiro,A.
 JOURNAL Unpublished (1996)
 STANDARD full staff_review
 COMMENT
 FEATURES Location/Qualifiers
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 /sequenced_mol="DNA"
 /sub_species="sonnerati"
 BASE COUNT 153 a 170 c 78 g 179 t 0 others
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361 ccatgtctg tccccttaca agtaccccaa ctatgtatgg tcaaggaca tgaacctaaa
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LOCUS CHKMTV16 457 bp ds-DNA VRT 17-JUN-1996
 DEFINITION Gallus varius mitochondrial DNA for D-loop region.
 ACCESSION D82912
 KEYWORDS D-loop region.
 SOURCE Gallus varius mitochondrion DNA, clone:2.
 ORGANISM Gallus varius
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
 Neognathae; Galliformes; Phasianidae.
 REFERENCE 1 (bases 1 to 457)
 AUTHORS Fumihiro,A.
 TITLE Direct Submission
 JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
 Akishinonoya Fumihiro, Yamashina Institute for Ornithology; 115
 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan
 (E-mail:nkojimo@lab.nig.ac.jp)
 STANDARD full staff_review
 REFERENCE 2 (bases 1 to 457)
 AUTHORS Fumihiro,A.
 JOURNAL Unpublished (1996)
 STANDARD full staff_review
 COMMENT
 FEATURES Location/Qualifiers
 source 1..457
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Locus CHKMTV17 502 bp ds-DNA VRT 17-JUN-1996
 Definition Green Junglefowl mitochondrial DNA for D-loop region.
 Accession D82913
 Keywords D-loop region.
 Source Gallus varius (isolate:6) mitochondrion DNA.
 Organism Gallus varius
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.
 Reference 1 (bases 1 to 502)
 Authors Fumihiro,A.
 Title Direct Submission
 Journal Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
 Akishinonomiya Fumihiro, Yamashina Institute for Ornithology; 115
 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan
 (E-mail:nkojima@lab.nig.ac.jp)
 Standard full staff_review
 Reference 2 (bases 1 to 502)
 Authors Fumihiro,A.
 Journal Unpublished (1996)
 Standard full staff_review
 Comment
 Features Location/Qualifiers
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 /mitochondrion
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 /sequenced_mol="DNA"
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 181 ttctccccat gtacattcca tgaatgtaccc aagtcattac gtcgtccata ccctacatcc
 241 aactctatac cactaacccg tacttaact atgaatgttt acaggacata cctcttaatcc
 301 tagtgcctta cccttaacccg gtcacctaac tatgaatgtt tacaggacat acatctaact
 361 accatgtatc aaccctattt gttatgtctg tgcgtccata tggattttt gatcgtaacc
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 481 ccctacacc cttgcctact tg
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 Locus CHKMTV18 474 bp ds-DNA VRT 17-JUN-1996
 Definition Green Junglefowl mitochondrial DNA for D-loop region.
 Accession D82914
 Keywords D-loop region.
 Source Gallus varius (isolate:32) mitochondrion DNA.
 Organism Gallus varius
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.
 Reference 1 (bases 1 to 474)
 Authors Fumihiro,A.
 Title Direct Submission
 Journal Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
 Akishinonomiya Fumihiro, Yamashina Institute for Ornithology; 115
 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan
 (E-mail:nkojima@lab.nig.ac.jp)
 Standard full staff_review
 Reference 2 (bases 1 to 474)
 Authors Fumihiro,A.
 Journal Unpublished (1996)
 Standard full staff_review
 Comment
 Features Location/Qualifiers
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 /mitochondrion
 /organism="Gallus varius"
 /sequenced_mol="DNA"
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 241 aactctatac cactaacccg tacttaact atgaatgttt acaggacata cctcttaatcc
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 361 cagtgcctta cccttaacccg gtcacctaac tatgaatgtt tacaggacat acatctaact
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LOCUS CHKMTV19 520 bp ds-DNA VRT 17-JUN-1996
 DEFINITION Green Junglefowl mitochondrial DNA for D-loop region.
 ACCESSION D82915
 KEYWORDS D-loop region.
 SOURCE Gallus varius mitochondrion DNA, clone:50.
 ORGANISM Gallus varius
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.
 REFERENCE 1 (bases 1 to 520)
 AUTHORS Fumihito,A.
 TITLE Direct Submission
 JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
 Akishinonomiya Fumihito, Yamashina Institute for Ornithology; 115
 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan
 (E-mail:nkojima@lab.nig.ac.jp)
 STANDARD full staff_review
 REFERENCE 2 (bases 1 to 519)
 AUTHORS Fumihito,A.
 JOURNAL Unpublished (1996)
 STANDARD full staff_review
 COMMENT
 FEATURES Location/Qualifiers
 source 1..519
 /clone="50"
 /mitochondrion
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 /sequenced_mol="DNA"
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 repeat_unit 307..367
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 301 agtgctctac ccccaacagg tcaccttaact atgaatgggt acaggacata cctctaatgt
 361 tagtgctcta ccccaacagg gtacatccaa tgatgttgc tacaggacat acctctaata
 421 tttagtgctcc tacccttcaac aggtcaccta accatgtatg gttacaggacat atacatctaa
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 LOCUS CHKMTD20 483 bp ds-DNA VRT 17-JUN-1996
 DEFINITION Red Junglefowl mitochondrial DNA for D-loop region.
 ACCESSION D82916
 KEYWORDS D-loop region.
 SOURCE Gallus gallus (sub_species:domesticus, strain:Ayam Cemani, isolate:1) mitochondrion DNA.
 ORGANISM Gallus gallus
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.
 REFERENCE 1 (bases 1 to 483)
 AUTHORS Fumihito,A.
 TITLE Direct Submission
 JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
 Akishinonomiya Fumihito, Yamashina Institute for Ornithology; 115
 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan
 (E-mail:nkojima@lab.nig.ac.jp)
 STANDARD full staff_review
 REFERENCE 2 (bases 1 to 483)
 AUTHORS Fumihito,A.
 JOURNAL Unpublished (1996)
 STANDARD full staff_review
 COMMENT
 FEATURES Location/Qualifiers
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 /mitochondrion
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 241 agtcacaaac cactaccagg taaccttaact atgaatgggt gcaggacata aatctctac
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 421 ctcaegagag atcagacaa cctgcgttta atgtactta tgaccatct caggccattc
 481 ttt
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LOCUS CHKMTD21 446 bp ds-DNA VRT 17-JUN-1996
 DEFINITION Red Junglefowl mitochondrial DNA for D-loop region.
 ACCESSION D82917
 KEYWORDS D-loop region.
 SOURCE *Gallus gallus* (*sub_species:domesticus*, strain:Ayamkokok Balenggek, isolate:63) mitochondrial DNA.
 ORGANISM *Gallus gallus*
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.
 REFERENCE 1 (bases 1 to 446)
 AUTHORS Fumihiro,A.
 TITLE Direct Submission
 JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
 Akishinonomiya Fumihiro, Yamashina Institute for Ornithology; 115 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan
 (E-mail:nkojima@lab.nig.ac.jp)
 STANDARD full staff_review
 REFERENCE 2 (bases 1 to 446)
 AUTHORS Fumihiro,A.
 JOURNAL Unpublished (1996)
 STANDARD full staff_review
 COMMENT
 FEATURES source Location/Qualifiers
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 /mitochondrion
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 /sub_species="domesticus"
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 181 ttctcccaat gtccattcta tgcatgtatcc agggcatact cattcaccc ccccatagac
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 301 tcatgtctt cccccaacaa gtccatcaa tctgtatggt tgccaggacat acatttact
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 LOCUS CHKMTD22 428 bp ds-DNA VRT 17-JUN-1996
 DEFINITION Red Junglefowl mitochondrial DNA for D-loop region.
 ACCESSION D82918
 KEYWORDS D-loop region.
 SOURCE *Gallus gallus* (*sub_species:domesticus*, strain:Ayamkokok Balenggek, isolate:71) mitochondrial DNA.
 ORGANISM *Gallus gallus*
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.
 REFERENCE 1 (bases 1 to 428)
 AUTHORS Fumihiro,A.
 TITLE Direct Submission
 JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
 Akishinonomiya Fumihiro, Yamashina Institute for Ornithology; 115 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan
 (E-mail:nkojima@lab.nig.ac.jp)
 STANDARD full staff_review
 REFERENCE 2 (bases 1 to 428)
 AUTHORS Fumihiro,A.
 JOURNAL Unpublished (1996)
 STANDARD full staff_review
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 FEATURES source Location/Qualifiers
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 /mitochondrion
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 /sequenced_mol="DNA"
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 /sub_species="domesticus"
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LOCUS	CHKMTD23	400 bp ds-DNA	VRT	17-JUN-1996	LOCUS	CHKMTD24	502 bp ds-DNA	VRT	17-JUN-1996										
DEFINITION	Red Junglefowl mitochondrial DNA for D-loop region.				DEFINITION	Chicken mitochondrial DNA for D-loop region.													
ACCESSION	D82919				ACCESSION	D82920													
KEYWORDS	D-loop region.				KEYWORDS	D-loop region.													
SOURCE	Gallus gallus (sub_species:domesticus, strain:Pelung) mitochondrial DNA, clone:1.				SOURCE	Gallus gallus (sub_species:domesticus, strain:Barred Plymouth Rock) mitochondrial DNA, clone:1.													
ORGANISM	Gallus gallus				ORGANISM	Gallus gallus													
Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.					Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.														
REFERENCE	1 (bases 1 to 400)				REFERENCE	1 (bases 1 to 502)													
AUTHORS	Fumihiato,A.				AUTHORS	Fumihiato,A.													
TITLE	Direct Submission				TITLE	Direct Submission													
JOURNAL	Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases. Akishinonomiya Fumihiato, Yamashina Institute for Ornithology; 115 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan (E-mail:nkojima@lab.nig.ac.jp)				JOURNAL	Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases. Akishinonomiya Fumihiato, Yamashina Institute for Ornithology; 115 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan (E-mail:nkojima@lab.nig.ac.jp)													
STANDARD	full staff_review				STANDARD	full staff_review													
REFERENCE	2 (bases 1 to 400)				REFERENCE	2 (bases 1 to 502)													
AUTHORS	Fumihiato,A.				AUTHORS	Fumihiato,A.													
JOURNAL	Unpublished (1996)				JOURNAL	Unpublished (1996)													
STANDARD	full staff_review				STANDARD	full staff_review													
COMMENT					COMMENT														
FEATURES	Location/Qualifiers				FEATURES	Location/Qualifiers													
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LOCUS CHKMTD25 491 bp ds-DNA VRT 17-JUN-1996
DEFINITION Red Junglefowl mitochondrial DNA for D-loop region.
ACCESSION D82921
KEYWORDS D-loop region.
SOURCE Gallus gallus (sub_species:domesticus, strain:Nagoya) mitochondrial DNA, clone:1.
ORGANISM Gallus gallus
Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.
REFERENCE 1 (bases 1 to 491)
AUTHORS Fumihiito,A.
TITLE Direct Submission
JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
Akishinonomiya Fumihiito, Yamashina Institute for Ornithology; 115 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan
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STANDARD full staff_review
REFERENCE 2 (bases 1 to 442)
AUTHORS Fumihiito,A.
JOURNAL Unpublished (1996)
STANDARD full staff_review
COMMENT
FEATURES Location/Qualifiers
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LOCUS CHKMTD26 476 bp ds-DNA VRT 17-JUN-1996
DEFINITION Red Junglefowl mitochondrial DNA for D-loop region.
ACCESSION D82922
KEYWORDS D-loop region.
SOURCE Gallus gallus (sub_species:domesticus, strain:Bantam, isolate:8-3) mitochondrial DNA.
ORGANISM Gallus gallus
Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.
REFERENCE 1 (bases 1 to 476)
AUTHORS Fumihiito,A.
TITLE Direct Submission
JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
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(E-mail:nkojima@lab.nig.ac.jp)
STANDARD full staff_review
REFERENCE 2 (bases 1 to 476)
AUTHORS Fumihiito,A.
JOURNAL Unpublished (1996)
STANDARD full staff_review
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FEATURES Location/Qualifiers
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LOCUS CHKMTD27 400 bp ds-DNA VRT 17-JUN-1996
DEFINITION Red Junglefowl mitochondrial DNA for D-loop region.
ACCESSION D82923
KEYWORDS D-loop region.
SOURCE Gallus gallus (*sub_species:domesticus*, strain:White leghorn) mitochondrial DNA, clone:2.
ORGANISM Gallus gallus
Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.
REFERENCE 1 (bases 1 to 400)
AUTHORS Fumihiito,A.
TITLE Direct Submission
JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
Akishinonomiya Fumihiito, Yamashina Institute for Ornithology; 115 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan
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STANDARD full staff_review
REFERENCE 2 (bases 1 to 400)
AUTHORS Fumihiito,A.
JOURNAL Unpublished (1996)
STANDARD full staff_review
COMMENT
FEATURES Location/Qualifiers
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LOCUS QULMTJ28 465 bp ds-DNA VRT 17-JUN-1996
DEFINITION Quail mitochondrial DNA for D-loop region.
ACCESSION D82924
KEYWORDS D-loop region.
SOURCE Coturnix coturnix (*sub_species:japonica*) mitochondrial DNA.
ORGANISM Coturnix coturnix
Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.
REFERENCE 1 (bases 1 to 465)
AUTHORS Fumihiito,A.
TITLE Direct Submission
JOURNAL Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
Akishinonomiya Fumihiito, Yamashina Institute for Ornithology; 115 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan
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STANDARD full staff_review
REFERENCE 2 (bases 1 to 465)
AUTHORS Fumihiito,A.
JOURNAL Unpublished (1996)
STANDARD full staff_review
COMMENT
FEATURES Location/Qualifiers
source 1..465
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241 ctttccacta acaggacacc ataaactatga atgggtcgag gacataaqt tactaaatcc
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421 ccccttacac ccctcgcccc tcttgctt ttgcgcctt ggttc

Locus CHKMTD29 400 bp ds-DNA VRT 17-JUN-1996
 Definition Red Junglefowl mitochondrial DNA for D-loop region.
 Accession D82925
 Keywords D-loop region.
 Source Gallus gallus (sub_species:domesticus, strain:White leghorn) mitochondrial DNA, clone:3.
 Organism Gallus gallus
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.
 Reference 1 (bases 1 to 400)
 Authors Fumihiro,A.
 Title Direct Submission
 Journal Submitted (10-JAN-1996) to the DDBJ/EMBL/GenBank databases.
 Akishinonomiya Fumihiro, Yamashina Institute for Ornithology; 115 Tsutsumine-aza, Abiko-shi, Chiba 270-11, Japan
 (E-mail:nkojima@lab.nig.ac.jp)
 Standard full staff_review
 Reference 2 (bases 1 to 400)
 Authors Fumihiro,A.
 Journal Unpublished (1996)
 Standard full staff_review
 Comment
 Features Location/Qualifiers
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 /organism="Gallus gallus"
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 /strain="White leghorn"
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 241 agctctaaac cactaccgaq taaccttaact atgatgttt acaggacata aatctcaact
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 Locus AGUMTK 423 bp ds-DNA ORG 26-FEB-1996
 Definition Great argus pheasant mitochondrial gene, D-loop region.
 Accession D66898
 Keywords D-loop; hypervariable region.
 Source Argusianus argus adult blood mitochondrial DNA.
 Organism Mitochondrion Argusianus argus
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.
 Reference 1 (bases 1 to 423)
 Authors Miyake,T.
 Title Direct Submission
 Journal Submitted (14-SEP-1995) to the DDBJ/EMBL/GenBank databases.
 Miyake, Wakunaga Pharmaceutical Co., Ltd., Institute for Biotechnology Research, 1624 Shimokotachi, Koda-cho, Takata-gun, Hiroshima 739-11, Japan
 Standard full staff_review
 Reference 2 (bases 1 to 423)
 Authors Akishinonomiya,F., Miyake,T., Takada,M., Ohno,S. and Kondo,N.
 Title The genetic link between the Chinese bamboo partridge (*Bambusicola thraupica*) and the chicken and junglefowls of the genus *Gallus*.
 Journal Proc. Natl. Acad. Sci. U.S.A. 92, 11053-11056 (1995)
 Standard full staff_review
 Comment Submitted (14-Sep-1995) to DDBJ by:
 Tetsuo Miyake
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 Japan
 Phone: 0826-45-2331
 Fax: 0826-45-4351
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 421 ctt
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LOCUS AUKMTC 444 bp ds-DNA ORG 26-FEB-1996
 DEFINITION Chukar partridge mitochondrial gene, D-loop region.
 ACCESSION D66890
 KEYWORDS D-loop; hypervariable region.
 SOURCE Alectoris chukar adult blood mitochondrial DNA.
 ORGANISM Mitochondrion Alectoris chukar
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.
 REFERENCE 1 (bases 1 to 444)
 AUTHORS Miyake,T.
 TITLE Direct Submission
 JOURNAL Submitted (14-SEP-1995) to the DDBJ/EMBL/GenBank databases. Tetsuo Miyake, Wakunaga Pharmaceutical Co., Ltd., Institute for Biotechnology Research, 1624 Shimokotachi, Koda-cho, Takata-gun, Hiroshima 739-11, Japan
 STANDARD full staff_review
 REFERENCE 2 (bases 1 to 444)
 AUTHORS Akishinonomiya,F., Miyake,T., Takada,M., Ohno,S. and Kondo,N.
 TITLE The genetic link between the Chinese bamboo partridge (*Bambusicola thoracica*) and the chicken and junglefowls of the genus *Gallus*.
 JOURNAL Proc. Natl. Acad. Sci. U.S.A. 92, 11053-11056 (1995)
 STANDARD full staff_review
 COMMENT Submitted (14-Sep-1995) to DDBJ by:
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 Japan
 Phone: 0826-45-2331
 Fax: 0826-45-4351
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 LOCUS BBAMTB 451 bp ds-DNA ORG 26-FEB-1996
 DEFINITION Chinese bamboo partridge mitochondrial gene, D-loop region.
 ACCESSION D66889
 KEYWORDS D-loop; hypervariable region.
 SOURCE *Bambusicola thoracica* adult blood mitochondrial DNA.
 ORGANISM Mitochondrion *Bambusicola thoracica*
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.
 REFERENCE 1 (bases 1 to 451)
 AUTHORS Miyake,T.
 TITLE Direct Submission
 JOURNAL Submitted (14-SEP-1995) to the DDBJ/EMBL/GenBank databases. Tetsuo Miyake, Wakunaga Pharmaceutical Co., Ltd., Institute for Biotechnology Research, 1624 Shimokotachi, Koda-cho, Takata-gun, Hiroshima 739-11, Japan
 STANDARD full staff_review
 REFERENCE 2 (bases 1 to 451)
 AUTHORS Akishinonomiya,F., Miyake,T., Takada,M., Ohno,S. and Kondo,N.
 TITLE The genetic link between the Chinese bamboo partridge (*Bambusicola thoracica*) and the chicken and junglefowls of the genus *Gallus*.
 JOURNAL Proc. Natl. Acad. Sci. U.S.A. 92, 11053-11056 (1995)
 STANDARD full staff_review
 COMMENT Submitted (14-Sep-1995) to DDBJ by:
 Tetsuo Miyake
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 Takatagun, Hiroshima 739-11
 Japan
 Phone: 0826-45-2331
 Fax: 0826-45-4351
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LOCUS CH2MTH 475 bp ds-DNA **ORG** 26-FEB-1996
DEFINITION Golden pheasant mitochondrial gene, D-loop region.
ACCESSION D66895
KEYWORDS D-loop; hypervariable region.
SOURCE Chrysolophus pictus adult blood mitochondrial DNA.
ORGANISM Mitochondrion Chrysolophus pictus
Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
Neognathae; Galliformes; Phasianidae.
REFERENCE 1 (bases 1 to 475)
AUTHORS Miyake,T.
TITLE Direct Submission
JOURNAL Submitted (14-SEP-1995) to the DDBJ/EMBL/GenBank databases. Tetsuo Miyake, Wakunaga Pharmaceutical Co., Ltd., Institute for Biotechnology Research, 1624 Shimokotachi, Koda-cho, Takata-gun, Hiroshima 739-11, Japan
STANDARD full staff_review
REFERENCE 2 (bases 1 to 475)
AUTHORS Akishinonomiya,F., Miyake,T., Takada,M., Ohno,S. and Kondo,N.
TITLE The genetic link between the Chinese bamboo partridge (*Bambusicola thraecica*) and the chicken and junglefowls of the genus *Gallus*.
JOURNAL Proc. Natl. Acad. Sci. U.S.A. 92, 11053-11056 (1995)
STANDARD full staff_review
COMMENT Submitted (14-Sep-1995) to DDBJ by:
Tetsuo Miyake
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Phone: 0826-45-2331
Fax: 0826-45-4351
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LOCUS CHKMTA 502 bp ds-DNA **ORG** 26-FEB-1996
DEFINITION Green junglefowl mitochondrial gene, D-loop region.
ACCESSION D64163
KEYWORDS D-loop; hypervariable region.
SOURCE Gallus varius adult blood mitochondrial DNA.
ORGANISM Mitochondrion Gallus varius
Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
Neognathae; Galliformes; Phasianidae.
REFERENCE 1 (bases 1 to 502)
AUTHORS Miyake,T.
TITLE Direct Submission
JOURNAL Submitted (15-Sep-1995) to the DDBJ/EMBL/GenBank databases. Tetsuo Miyake, Wakunaga Pharmaceutical Co., Ltd., Institute for Biotechnology Research, 1624 Shimokotachi, Koda-cho, Takata-gun, Hiroshima 739-11, Japan
STANDARD full staff_review
REFERENCE 2 (bases 1 to 502)
AUTHORS Akishinonomiya,F., Miyake,T., Takada,M., Ohno,S. and Kondo,N.
TITLE The genetic link between the Chinese bamboo partridge (*Bambusicola thraecica*) and the chicken and junglefowls of the genus *Gallus*.
JOURNAL Proc. Natl. Acad. Sci. U.S.A. 92, 11053-11056 (1995)
STANDARD full staff_review
COMMENT Submitted (15-Sep-1995) to DDBJ by:
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Japan
Phone: 0826-45-2331
Fax: 0826-45-4331
FEATURES
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181 ttctccatg tacatatacc tcgaatgcgc ccaaaatcata caagctccct cccaaaaaca
241 aactctatac cactaacggg tcacttaact atgaatggt acaggacata cctctaatac
301 tagtgctta cccttaacacg gtccatgttgcacccatc atgtatgtt tacaggacata acatctaact
361 accatgttgc aaccatgttgc ttatgttgc tgcgtccatc tggtttttt gatgttgcacc
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481 cccctacacc ctgttgcataat tg

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LOCUS CHIKMTE 580 bp ds-DNA **ORG** 26-FEB-1996
DEFINITION Grey junglefowl mitochondrial gene, D-loop region.
ACCESSION D66892
KEYWORDS D-loop; hypervariable region.
SOURCE Gallus sonneratii adult blood mitochondrial DNA.
ORGANISM Mitochondrion Gallus sonneratii
Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
Neognathae; Galliformes; Phasianidae.
REFERENCE 1 (bases 1 to 580)
AUTHORS Miyake,T.
TITLE Direct Submission
JOURNAL Submitted (14-SEP-1995) to the DDBJ/EMBL/GenBank databases. Tetsuo Miyake, Wakunaga Pharmaceutical Co., Ltd., Institute for Biotechnology Research, 1624 Shimokotachi, Koda-cho, Takata-gun, Hiroshima 739-11, Japan
STANDARD full staff_review
REFERENCE 2 (bases 1 to 580)
AUTHORS Akishinonomiya,F., Miyake,T., Takada,M., Ohno,S. and Kondo,N.
TITLE The genetic link between the Chinese bamboo partridge (*Bambusicola thraecica*) and the chicken and junglefowls of the genus *Gallus*.
JOURNAL Proc. Natl. Acad. Sci. U.S.A. 92, 11053-11056 (1995)
STANDARD full staff_review
COMMENT Submitted (14-Sep-1995) to DDBJ by:
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Phone: 0826-45-2331
Fax: 0826-45-4351
FEATURES
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181 ttctcccat gtccatctca tgaatggtcc aagtcatctt attcaccta cccatcatg
241 gcttatatcc actaccaggc cacctaacta tgaatggttt caggacatac acttaacttt
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361 ccataatcg tccccatcaa agtccatcaa ctatgtatgg tcaaggaca tgaacctaaa
421 tccttatgtt cttaaccattt tggttatgtt cgtgtatca gtatgttta ttgtatgttc
481 accttacacgag agatcagcaaa cccctgcctg taatgttactt catgaccatgtt ctcaggccca
541 ttctttccccc ctacaccctt cgcctactt gccttcacc
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LOCUS CHIKMTF 578 bp ds-DNA **ORG** 26-FEB-1996
DEFINITION Sri Lanka junglefowl mitochondrial gene, D-loop region.
ACCESSION D66893
KEYWORDS D-loop; hypervariable region.
SOURCE Gallus lafayettei adult blood mitochondrial DNA.
ORGANISM Mitochondrion Gallus lafayettei
Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
Neognathae; Galliformes; Phasianidae.
REFERENCE 1 (bases 1 to 578)
AUTHORS Miyake,T.
TITLE Direct Submission
JOURNAL Submitted (14-SEP-1995) to the DDBJ/EMBL/GenBank databases. Tetsuo Miyake, Wakunaga Pharmaceutical Co., Ltd., Institute for Biotechnology Research, 1624 Shimokotachi, Koda-cho, Takata-gun, Hiroshima 739-11, Japan
STANDARD full staff_review
REFERENCE 2 (bases 1 to 578)
AUTHORS Akishinonomiya,F., Miyake,T., Takada,M., Ohno,S. and Kondo,N.
TITLE The genetic link between the Chinese bamboo partridge (*Bambusicola thraecica*) and the chicken and junglefowls of the genus *Gallus*.
JOURNAL Proc. Natl. Acad. Sci. U.S.A. 92, 11053-11056 (1995)
STANDARD full staff_review
COMMENT Submitted (14-Sep-1995) to DDBJ by:
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Phone: 0826-45-2331
Fax: 0826-45-4351
FEATURES
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181 ttctcccat gtccatctca tgaatggtcc aagtcatctt attcaccta cccatcatg
241 aatcttacc actaacaatgtt cccatctaacta tgaatggttt caggacatac acttaacttt
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421 cttctatgtt cttaaccattt tggttatgtt cgtgtatca gtatgttta ttgtatgttc
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LOCUS	LOUMTJ	442 bp ds-DNA	ORG	26-FEB-1996	LOCUS	PBTMTL	461 bp ds-DNA	ORG	26-FEB-1996
DEFINITION	Silver lophura mitochondrial gene, D-loop region.		DEFINITION	Burmese peacock-pheasant mitochondrial gene, D-loop region.					
ACCESSION	D66897		ACCESSION	D66899					
KEYWORDS	D-loop; hypervariable region.		KEYWORDS	D-loop; hypervariable region.					
SOURCE	Lophura nycthemera adult blood mitochondrial DNA.		SOURCE	Polyplectron bicalcaratum adult blood mitochondrial DNA.					
ORGANISM	Mitochondrion Lophura nycthemera		ORGANISM	Mitochondrion Polyplectron bicalcaratum					
Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.			Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.						
REFERENCE	1 (bases 1 to 442)		REFERENCE	1 (bases 1 to 461)					
AUTHORS	Miyake,T.		AUTHORS	Miyake,T.					
TITLE	Direct Submission		TITLE	Direct Submission					
JOURNAL	Submitted (14-SEP-1995) to the DDBJ/EMBL/GenBank databases. Tetsuo Miyake, Wakunaga Pharmaceutical Co., Ltd., Institute for Biotechnology Research, 1624 Shimokotachi, Koda-cho, Takata-gun, Hiroshima 739-11, Japan		JOURNAL	Submitted (14-SEP-1995) to the DDBJ/EMBL/GenBank databases. Tetsuo Miyake, Wakunaga Pharmaceutical Co., Ltd., Institute for Biotechnology Research, 1624 Shimokotachi, Koda-cho, Takata-gun, Hiroshima 739-11, Japan					
STANDARD	full staff_review		STANDARD	full staff_review					
REFERENCE	2 (bases 1 to 442)		REFERENCE	2 (bases 1 to 461)					
AUTHORS	Akishinononya,F., Miyake,T., Takada,M., Ohno,S. and Kondo,N.		AUTHORS	Akishinononya,F., Miyake,T., Takada,M., Ohno,S. and Kondo,N.					
TITLE	The genetic link between the Chinese bamboo partridge (<i>Bambusicola thraupica</i>) and the chicken and junglefowls of the genus <i>Gallus</i> .		TITLE	The genetic link between the Chinese bamboo partridge (<i>Bambusicola thraupica</i>) and the chicken and junglefowls of the genus <i>Gallus</i> .					
JOURNAL	Proc. Natl. Acad. Sci. U.S.A. 92, 11053-11056 (1995)		JOURNAL	Proc. Natl. Acad. Sci. U.S.A. 92, 11053-11056 (1995)					
STANDARD	full staff_review		STANDARD	full staff_review					
COMMENT	Submitted (14-Sep-1995) to DDBJ by: Tetsuo Miyake Dept. of Institute for Biotechnology Research Wakunaga Pharmaceutical Co.,Ltd. 1624 Shimokotachi, Kodachou Takatagun, Hiroshima 739-11 Japan Phone: 0826-45-2331 Fax: 0826-45-4351		COMMENT	Submitted (14-Sep-1995) to DDBJ by: Tetsuo Miyake Dept. of Institute for Biotechnology Research Wakunaga Pharmaceutical Co.,Ltd. 1624 Shimokotachi, Kodachou Takatagun, Hiroshima 739-11 Japan Phone: 0826-45-2331 Fax: 0826-45-4351					
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	/tissue_type="blood"			/tissue_type="blood"					
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BASE COUNT	129 a 132 c 61 g 120 t 0 others		BASE COUNT	122 a 138 c 67 g 134 t 0 others					
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LOCUS	PHSMTG	455 bp ds-DNA	ORG	26-FEB-1996	LOCUS	PVQMTB	439 bp ds-DNA	ORG	26-FEB-1996
DEFINITION	Green pheasant mitochondrial gene, D-loop region.				DEFINITION	Green peafowl mitochondrial gene, D-loop region.			
ACCESSION	D66894		ACCESSION	D64164					
KEYWORDS	D-loop; hypervariable region.				KEYWORDS	D-loop; hypervariable region.			
SOURCE	Phasianus versicolor adult blood mitochondrial DNA.				SOURCE	Pavo muticus adult blood mitochondrial DNA.			
ORGANISM	Mitochondrion Phasianus versicolor Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.				ORGANISM	Mitochondrion Pavo muticus Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.			
REFERENCE	1 (bases 1 to 455)				REFERENCE	1 (bases 1 to 439)			
AUTHORS	Miyake,T.				AUTHORS	Miyake,T.			
TITLE	Direct Submission				TITLE	Direct Submission			
JOURNAL	Submitted (14-SEP-1995) to the DDBJ/EMBL/GenBank databases. Tetsuo Miyake, Wakunaga Pharmaceutical Co., Ltd., Institute for Biotechnology Research, 1624 Shimokotachi, Koda-cho, Takata-gun, Hiroshima 739-11, Japan				JOURNAL	Submitted (15-SEP-1995) to the DDBJ/EMBL/GenBank databases. Tetsuo Miyake, Wakunaga Pharmaceutical Co., Ltd., Institute for Biotechnology Research, 1624 Shimokotachi, Koda-cho, Takata-gun, Hiroshima 739-11, Japan			
STANDARD	full staff_review				STANDARD	full staff_review			
REFERENCE	2 (bases 1 to 455)				REFERENCE	2 (bases 1 to 439)			
AUTHORS	Akishinonomiya,F., Miyake,T., Takada,M., Ohno,S. and Kondo,N.				AUTHORS	Akishinonomiya,F., Miyake,T., Takada,M., Ohno,S. and Kondo,N.			
TITLE	The genetic link between the Chinese bamboo partridge (<i>Bambusicola thraupica</i>) and the chicken and junglefowls of the genus <i>Gallus</i> .				TITLE	The genetic link between the Chinese bamboo partridge (<i>Bambusicola thraupica</i>) and the chicken and junglefowls of the genus <i>Gallus</i> .			
JOURNAL	Proc. Natl. Acad. Sci. U.S.A. 92, 11053-11056 (1995)				JOURNAL	Proc. Natl. Acad. Sci. U.S.A. 92, 11053-11056 (1995)			
STANDARD	full staff_review				STANDARD	full staff_review			
COMMENT	Submitted (14-Sep-1995) to DDBJ by: Tetsuo Miyake Dept. of Institute for Biotechnology Research Wakunaga Pharmaceutical Co.,Ltd. 1624 Shimokotachi, Kodachou Takatagun, Hiroshima 739-11 Japan Phone: 0826-45-2331 Fax: 0826-45-4351				COMMENT	Submitted (15-Sep-1995) to DDBJ by: Tetsuo Miyake Intitute for Biotechnology Research Wakunaga Pharmaceutical Co.,Ltd. 1624 Shimokotachi, Kodachou Takatagun, Hiroshima 739-11 Japan Phone: 0826-45-2331 Fax: 0826-45-4351			
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	61	ggtatatact gtataatcgat gcatacattt atataccaca tatatactat ggtaccgtta	61	gtataactatg cataatcgatg catabatata tataccacat acattatggt cacatgtata					
	121	ctatataatata taatcgatact aaaccatata tatgtatgcg gacatttacac ctgtaccctta	121	ctatataatata acgttactaaat ccattatataat gtatgcggatc atttacatat ctccccat					
	181	tttctcccaa ttttcccaa ttgtacaaatc atgcacttcc ccaggatattt aatgtcttta cccctgtaaa	181	tatccccacg ttttcccaa atgtatctt agatcatatcca cttatataat cccatcccc					
	241	tgatctacaa cttccagggtt accataccat gaaatgttac aggacatata tcgtatata	241	gttcccaaaa cccacaaggcc acctaactat gatgttac aggttacccat gttcatat					
	301	ggtattatccc cttttgggtt tgctcgatgtt accagatggaa ttatgttgcg tttatgttgc	301	cgatctccccc ccattttgggtt atgcgtcgatc tatcatgttgg attttatgttgcg tttatgttgc					
	361	cgatgtatca ccaaaaaatcg ctatataatgtt atccatgtac tagtttcagg cccatttttt	361	acgttggatc agcaacccctt gctctgtatgtt tacttcatgtatgttgcg tttatgttgc					
	421	cccccttacac ccctcgcccccc ttgttgcgtt ttgcgtt	421	tcccccatac cccctcgcc					

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Locus PVQMTM 443 bp ds-DNA Org 26-FEB-1996
 Definition Common peafowl mitochondrial gene, D-loop region.
 Accession D66900
 Keywords D-loop; hypervariable region.
 Source Pavo cristatus adult blood mitochondrial DNA.
 Organism Mitochondrion Pavo cristatus
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.
 Standard full staff_review
 Reference 1 (bases 1 to 443)
 Authors Akishinonomiya,F., Miyake,T., Takada,M., Ohno,S. and Kondo,N.
 Title The genetic link between the Chinese bamboo partridge (*Bambusicola thraupica*) and the chicken and junglefowls of the genus *Gallus*.
 Journal Proc. Natl. Acad. Sci. U.S.A. 92, 11053-11056 (1995)
 Standard full staff_review
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 421 ttccccctac accccctgc ctt
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 Locus PXDMTD 483 bp ds-DNA Org 26-FEB-1996
 Definition European grey partridge mitochondrial gene, D-loop region.
 Accession D66891
 Keywords D-loop; hypervariable region.
 Source Perdix perdix adult blood mitochondrial DNA.
 Organism Mitochondrion Perdix perdix
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes; Neognathae; Galliformes; Phasianidae.
 Reference 1 (bases 1 to 483)
 Authors Miyake,T.
 Title Direct Submission
 Journal Submitted (14-SEP-1995) to the DDBJ/EMBL/GenBank databases. Tetsuo Miyake, Wakunaga Pharmaceutical Co., Ltd., Institute for Biotechnology Research, 1624 Shimokotachi, Koda-cho, Takata-gun, Hiroshima 739-11, Japan
 Standard full staff_review
 Reference 2 (bases 1 to 483)
 Authors Akishinonomiya,F., Miyake,T., Takada,M., Ohno,S. and Kondo,N.
 Title The genetic link between the Chinese bamboo partridge (*Bambusicola thraupica*) and the chicken and junglefowls of the genus *Gallus*.
 Journal Proc. Natl. Acad. Sci. U.S.A. 92, 11053-11056 (1995)
 Standard full staff_review
 Comment Submitted (14-Sep-1995) to DDBJ by:
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 Fax: 0826-45-4351
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 301 ctatcaccc cacatgttgtt tatgtctgac gtacccatg gattttatgtt tgccacact
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 481 tcc
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Locus QULM7A 459 bp ds-DNA ORG 26-FEB-1996
 Definition Blue-breasted quail mitochondrial gene, D-loop region.
 Accession D66888
 Keywords D-loop; hypervariable region.
 Source Coturnix sinesis adult blood mitochondrial DNA.
 Organism Mitochondrion Coturnix sinesis
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
 Neognathae; Galliformes; Phasianidae.
 Reference 1 (bases 1 to 459)
 Authors Miyake,T.
 Title Direct Submission
 Journal Submitted (14-SEP-1995) to the DDBJ/EMBL/GenBank databases. Tetsuo Miyake, Wakunaga Pharmaceutical Co., Ltd., Institute for Biotechnology Research, 1624 Shimokotachi, Koda-cho, Takata-gun, Hiroshima 739-11, Japan
 Standard full staff_review
 Reference 2 (bases 1 to 459)
 Authors Akishinonomiya,F., Miyake,T., Takada,M., Ohno,S. and Kondo,N.
 Title The genetic link between the Chinese bamboo partridge (*Bambusicola thoracica*) and the chicken and junglefowls of the genus *Gallus*.
 Journal Proc. Natl. Acad. Sci. U.S.A. 92, 11053-11056 (1995)
 Standard full staff_review
 Comment Submitted (14-Sep-1995) to DDBJ by:
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 Fax: 0826-45-4351
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 181 ctcccaatgt acattaatgc atgtctctag acattaaact ctcttacca gccccaggat
 241 ctaacccaaa caaggcacca tagacatgaa tggttacagg acataacctt aatacataat
 301 gtcatccac atttggtat gtcgtcgta tcagatgtt ttatgtatg gacacccatc
 361 gagatgtcg caacccatgc tgcgtatgtt tatcatgact agttcaggc ccatttttc
 421 cccctacacc ctcgeccctc ctgcgtt tgcgtct
 Locus SIYMTI 440 bp ds-DNA ORG 26-FEB-1996
 Definition Mrs. Hume's pheasant mitochondrial gene, D-loop region.
 Accession D66896
 Keywords D-loop; hypervariable region.
 Source Syrmaticus humiae adult blood mitochondrial DNA.
 Organism Mitochondrion Syrmaticus humiae
 Eukaryota; Animalia; Chordata; Vertebrata; Aves; Neornithes;
 Neognathae; Galliformes; Phasianidae.
 Reference 1 (bases 1 to 440)
 Authors Miyake,T.
 Title Direct Submission
 Journal Submitted (14-SEP-1995) to the DDBJ/EMBL/GenBank databases. Tetsuo Miyake, Wakunaga Pharmaceutical Co., Ltd., Institute for Biotechnology Research, 1624 Shimokotachi, Koda-cho, Takata-gun, Hiroshima 739-11, Japan
 Standard full staff_review
 Reference 2 (bases 1 to 440)
 Authors Akishinonomiya,F., Miyake,T., Takada,M., Ohno,S. and Kondo,N.
 Title The genetic link between the Chinese bamboo partridge (*Bambusicola thoracica*) and the chicken and junglefowls of the genus *Gallus*.
 Journal Proc. Natl. Acad. Sci. U.S.A. 92, 11053-11056 (1995)
 Standard full staff_review
 Comment Submitted (14-Sep-1995) to DDBJ by:
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 Japan
 Phone: 0826-45-2331
 Fax: 0826-45-4351
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 241 ctatgttcgg agccctcaag tccatataac tatgtatgtt tacaggacat actatcta
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 361 ctccacgagat atccaccaacc cctgtctgtt atgtattccg tgaccgact caggccatc
 421 cttttccctt acacccctcg
 //

Appendix B

**Photographs of the Phasianid Birds
that are the Same Species from which
the Blood Samples are Taken**



Coturnix coturnix japonica



Coturnix or Excalfactoria chinensis



Bambusicola thoracica



Alectoris graeca



Perdix perdix perdix



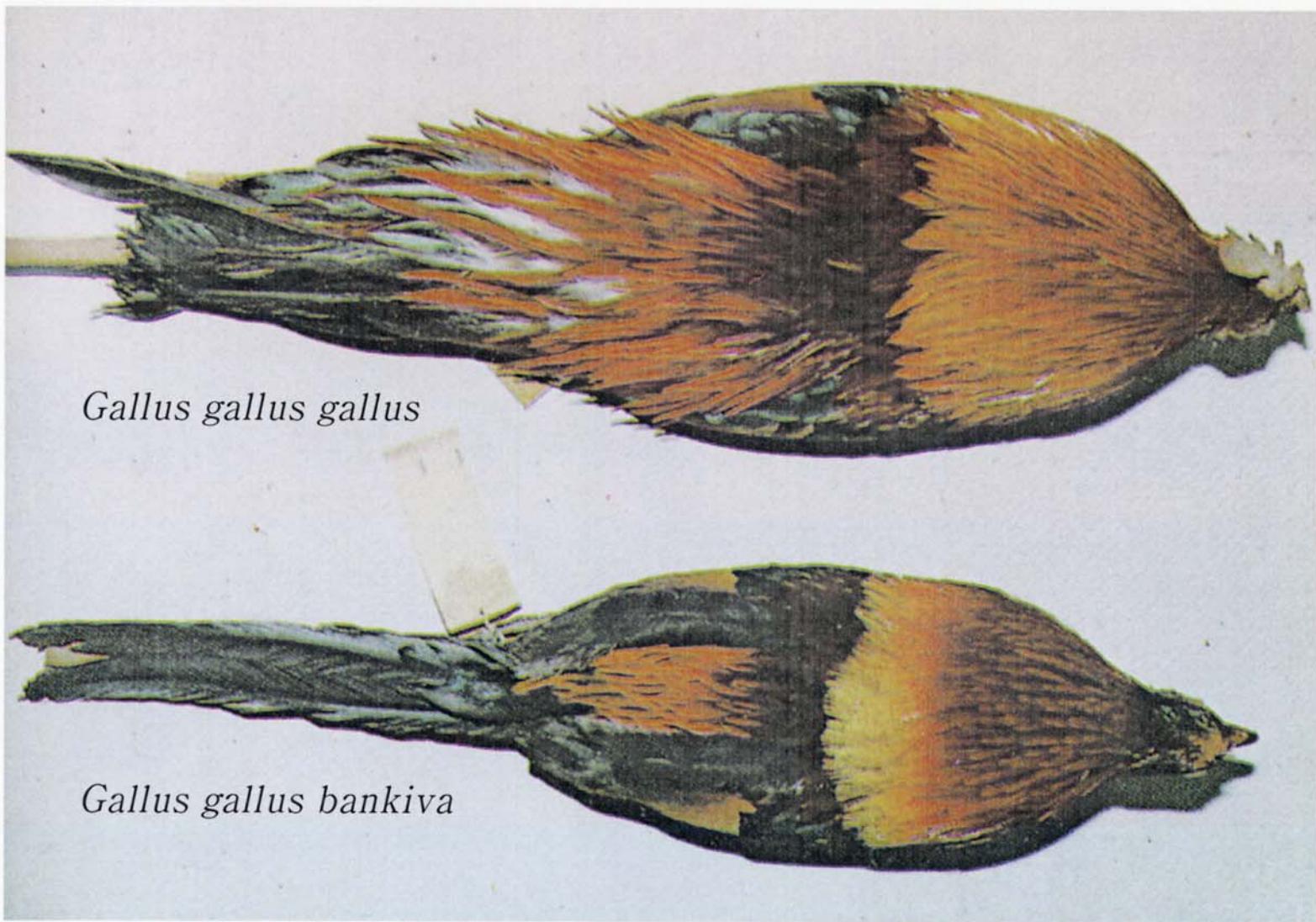
Gallus varius



Gallus gallus



Gallus gallus bankiva



Morphological comparison between *Gallus gallus gallus* and *Gallus gallus bankiva*



Gallus sonnerati



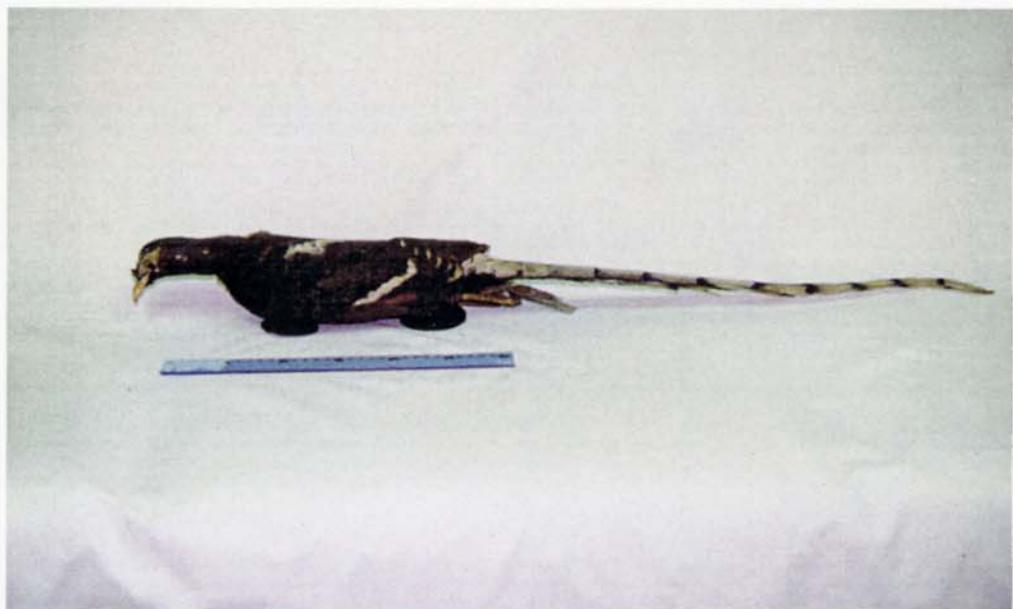
Gallus lafayettei



Phasianus versicolor



Chrysolophus pictus



Syrmaticus humiae



Lophura nympha



Pavo muticus



Pavo muticus



Pavo cristatus



Polyplectron bicalcaratum



Argusianus argus



Argusianus argus