氏 名 寺社下 美 樹

学位(専攻分野) 博士(理学)

学 位 記 番 号 総研大乙第64号

学位授与の日付 平成11年3月24日

学位授与の要件 学位規則第4条第2項該当

学位論文題目 Control of the Synthesis and Functions of RNA

Polymerase Sigma Subunits in Escherichia coli

論文審查委員 主 查 教 授 嶋本伸雄

教 授 荒木 弘之

助 教 授 西村 昭子

助 教 授 和田 明(大阪医科大学)

論文内容の要旨

Selection of a σ subunit of RNA polymerase is a powerful mechanism for switching transcription of a group of genes. For understanding the regulation of σ subunit utilization in Escherichia coli, I determined the intracellular levels of four members among 8 known σ subunits of E. coli strains, W3110 and MC4100, at various growth phases by a quantitative Western immunoblot analysis. The level of σ^{70} is rather constant throughout the growth, and those of σ^{54} and σ^{28} are maintained at 10 and 50% the level of σ^{70} in both strains growing at exponential and stationary phases, respectively. At the exponential growth phase σ^{38} is undetectable but increases to up to 30% of σ^{70} at the stationary phase, supporting the concept that σ^{38} plays a key role in transcription of the stationary phase-expressed genes. Stress-coupled change in the intracellular level was observed for two σ subunits: an increase in σ^{38} level and a decrease in σ^{28} level upon exposure to heat shock at the exponential phase; and the increase in σ^{38} level under a high osmolality condition at both the exponential and the stationary phases. I also found that the composition of σ subunits is different between bacterial strains, even under the same strain name. For instance, the compositions of σ^{28} and σ^{38} differ among three W3110 strains.

The growth transition from exponential to stationary phase is accompanied by the replacement of RNA polymerase-associated σ^{70} subunit with σ^{38} . She found that the GST-fused σ^{70} in stationary-phase cell extracts exists as a complex with a new protein, and it was designated Rsd (Regulator of Sigma D). A gene located at 90 min on the *E. coli* chromosome encodes Rsd, and is transcribed from upstream and downstream promoters with σ^{38} and σ^{70} holoenzymes, respectively. Over-expressed and purified Rsd protein can form a complex with σ^{70} in vitro but not with any other σ subunits; σ^{54} , σ^{38} , σ^{32} , σ^{28} and σ^{24} . A binding assay with proteolytic fragments of σ^{70} suggested that Rsd interacts with the conserved region 4, the promoter -35 recognition domain, and its downstream of σ^{70} . The isolated Rsd inhibited transcription in vitro to various extents depending on the promoters used. The intracellular level of Rsd starts to increase during the transition from growing to stationary phase.

Experiments using strains with disrupted rsd gene and those with overproduced Rsd indicated that Rsd is a negative factor for transcription directed by σ^{70} and a positive factor for that by σ^{38} . The results indicate that Rsd is a stationary-phase $E.\ coli$ protein regulating the activity of the σ^{70} function, providing an evidence for the existence of the control of the activities as well as the levels of the σ subunits of RNA polymerase, contributing to a group switching of genes of $E.\ coli$. Taking all the observations together, she discussed possible models for the switching mechanism.

論文の審査結果の要旨

Selection of a σ subunit of RNA polymerase is a powerful mechanism for switching transcription of a group of genes. For understanding the regulation of σ subunit utilization in Escherichia coli, Ms. M. Jishage determined the intracellular levels of four members among 8 known σ subunits of E. coli strains, W3110 and MC4100, at various growth phases by a quantitative Western immunoblot analysis. The level of σ^{70} is rather constant throughout the growth, and those of σ^{54} and σ^{28} are maintained at 10 and 50% the level of σ^{70} in both strains growing at exponential and stationary phases, respectively. At the exponential growth phase σ^{38} is undetectable but increases to up to 30% of σ^{70} at the stationary phase, supporting the concept that σ^{38} plays a key role in transcription of the stationary phase-expressed genes. Stress-coupled change in the intracellular level was observed for two σ subunits: an increase in σ^{38} level and a decrease in σ^{28} level upon exposure to heat shock at the exponential phase; and the increase in σ^{38} level under a high osmolality condition at both the exponential and the stationary phases. She also found that the composition of σ subunits is different between bacterial strains, even under the same strain name. For instance, the compositions of σ^{28} and σ^{38} differ among three W3110 strains.

The growth transition from exponential to stationary phase is accompanied by the replacement of RNA polymerase-associated σ^{70} subunit with σ^{38} . She found that the GST-fused σ^{70} in stationary-phase cell extracts exists as a complex with a new protein, and it was designated Rsd (Regulator of Sigma D). A gene located at 90 min on the *E. coli* chromosome encodes Rsd, and is transcribed from upstream and downstream promoters with σ^{38} and σ^{70} holoenzymes, respectively. Over-expressed and purified Rsd protein can form a complex with σ^{70} in vitro but not with any other σ subunits; σ^{54} , σ^{38} , σ^{32} , σ^{28} and σ^{24} . A binding assay with proteolytic fragments of σ^{70} suggested that Rsd interacts with the conserved region 4, the promoter -35 recognition domain, and its downstream of σ^{70} . The isolated Rsd inhibited transcription *in vitro* to various extents depending on the promoters used. The intracellular level of Rsd starts to increase during the transition from growing to stationary phase.

Experiments using strains with disrupted rsd gene and those with overproduced Rsd indicated that Rsd is a negative factor for transcription directed by σ^{70} and a positive factor for that by σ^{38} . The results indicate that Rsd is a stationary-phase $E.\ coli$ protein regulating the activity of the σ^{70} function, providing an evidence for the existence of the control of the activities as well as the levels of the σ subunits of RNA polymerase, contributing to a group switching of genes of $E.\ coli$. Taking all the observations together, she discussed possible models for the switching mechanism.

The committee concluded that this dissertation fulfills the requirements for the degree of PhD (Science) for Miki Jishage.